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Variations in milk protein fractions affect the efficiency of the cheese-making process

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ABSTRACT

The aim of this study was to assess the influence of the amounts of the α_{S1} -, α_{S2} -, β -, and κ -casein (CN) and the α -lactalbumin and β -lactoglobulin protein fractions on the efficiency of the cheese-making process independently of their genetic polymorphisms. The study was carried out on milk samples from 1,271 Brown Swiss cows from 85 herds classified into 4 categories according to management, feeding, and housing characteristics (traditional and modern systems). To assess the efficiency of the cheese-making process, we processed the milk samples according to a laboratory cheese-making procedure (1,500 mL/sample) and obtained the following measures: (1) 3 percentage cheese yields (%CY_{curd}, %CY_{solids}, %CY_{water}), (2) 2 daily cheese yields obtained by multiplying %CY (curd and total solids) by daily milk yields (dCY_{curd}, dCY_{solids}), (3) 4 measures of nutrient recovery in the curd (REC_{fat}, REC_{protein}, REC_{solids}, REC_{energy}), and (4) 2 measures of cheese-making efficiency in terms of the ratio between the observed and theoretical %CY (Ef-%CY_{curd}, Ef-%CY_{solids}). All the aforementioned traits were analyzed by fitting 2 linear mixed models with protein fractions as fixed effects expressed as percentage in the milk (model M-%milk) and as percentage of the total casein content (model M-%cas) together with the effects of total casein content (only in model M-%cas), daily milk yield (only in model M-%milk; not for dCY traits), dairy system, herd (random effect), days in milk, parity, and vat. The efficiency of overall cheese yield (Ef-%CY_{curd}) was mostly positively associated with β -CN content in the milk, whereas Ef-%CY_{solids} was greater with higher amounts of κ -CN and α_{S1} -CN (M-%milk) due to the strong influ-

ence of both fractions on the recovery rate of milk components in the curd (fat and total solids, protein with α_{S1} -CN only) when expressed as percentage of milk and of total casein; only β -CN was more important for REC_{protein}. In contrast, we found β -lactoglobulin to be highly negatively related to all the traits related to the cheese-making process and to the daily cheese yield per cow, whereas α -lactalbumin was positively associated with the latter traits. Additional research on this topic is needed, with particular focus on the genetic and genomic aspects of the role of protein fractions in the cheese-making process and on the associations between genetic polymorphisms in milk protein and milk nutrient recovery in the curd.

Key words: milk protein, cheese-making, cheese yield, nutrient recovery

INTRODUCTION

The cheese production industry has a crucial need for an integrated approach to monitor all the relationships between milk quality and cheese-making so that it can guarantee a high level of efficiency (in terms of cheese yield and recovery of milk nutrients in the curd) across the entire process (Banks, 2007). Such an approach would also include gathering information on the milk quality of individual animals for use in current practices related to the genetic improvement of dairy cattle populations and for adjusting the milk-to-cheese economic value (i.e., milk payment systems).

Among the various factors involved, there is universal acknowledgment of the role played by the composition of protein fractions in the cheese-making process from milk gelation to cheese ripening (Emmons et al., 2003; Guinee, 2003; Caroli et al., 2009). As cheese consists of a paracasein reticulum in which fat globules and part of the soluble phase of milk are entrapped (Guinee, 2003; Rybak, 2014), the concentration of casein in milk is positively correlated with the quantity of cheese produced per unit of milk (percentage cheese yield). Moreover,

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physicochemical characteristics and structural properties of milk casein micelles (average size, proportion of caseins, concentration of colloidal calcium phosphate, casein genetic variants) influence the rheological properties of the resulting rennet curd and, consequently, its capacity to retain milk constituents—mainly water, casein, and fat—in cheese (cheese-making efficiency; Bittante et al., 2012). In fact, when empirical predictive formulas for cheese yield using milk composition are to be developed, the coefficients of casein are higher than its own weight (Emmons et al., 1990).

Most studies at the cow level limit their analyses to the effect of protein fractions on the coagulation process (Macheboeuf et al., 1993; Hallén et al., 2007) or sample only a small number of animals (Wedholm et al., 2006; Hallén et al., 2010) due to the time-consuming and labor-intensive nature of cheese-making trials. Furthermore, there is a lack of knowledge about the role of proteins in milk nutrient recovery in the curd. Recently, we proposed a laboratory model cheese-manufacturing procedure that allows several cheese-making traits to be measured. This method was used to process more than 1,000 individual Brown Swiss milk samples, which revealed high variability in cheese-making traits related to dairy system (Bittante et al., 2015) and animal characteristics (Stocco et al., 2018). The aim of the present study was to examine the influence of the amounts of protein fractions (caseins and whey proteins), expressed as percentage in milk, and their proportions to total casein content on the efficiency of the cheese-making process and on the daily cheese production of individual cows.

MATERIALS AND METHODS

Animals, Dairy Systems, and Milk Sample Collection

This study falls within the scope of the Cowability–Cowplus project. Herd selection, dairy farming systems, and sampling procedure are described in detail in Bittante et al. (2015). Briefly, milk samples were taken from a total of 1,264 Brown Swiss cows (3 subsamples per cow) during the evening milking. Cows were selected from the clinically healthy cows of 85 herds (a maximum of 15 cows per herd) located in Trento Province (Italy) to represent different parities and lactation stages. The herds were chosen from 610 farms and classified into 4 dairy farming systems (3 modern and 1 traditional).

One subsample of milk (50 mL) was transported to the Milk Quality Laboratory of the Trento Breeders Association (Trento, Italy) for milk composition analysis. The other subsamples were taken to the Milk

Laboratory of the Department of Agronomy, Food, Natural Resources, Animals and Environment of the University of Padua (Legnaro, Padua, Italy) for cheese-making and protein fraction analyses. All phases, from sampling to storage of milk samples, were standardized to maximize reproducibility among herds and collection dates. The Superbrown Consortium of Trento (Trento, Italy) provided information on the cows and herds.

Milk Analyses and Processing

Gross Composition Traits. Individual raw full-fat milk samples (50 mL) of all cows sampled were analyzed within 20 h of milking for gross composition (protein, casein, fat, lactose, and TS) using a MilkoScan FT6000 (Foss, Hillerød, Denmark) calibrated according to the following reference methods: fat (ISO, 2010b; ISO1211/IDF 1; gravimetric method, Röse-Gottlieb), protein (ISO, 2014; ISO 8968-1/IDF 20-1; titrimetric method, Kjeldahl), casein (ISO, 2004; ISO 17997-1/IDF 29; titrimetric method, Kjeldahl), lactose (ISO, 2002; ISO 5765-1/IDF 79-1; enzymatic method), and TS (ISO, 2010a; ISO 6731/IDF 21; determination of TS content). Somatic cell count was measured with a Fossomatic FC counter (Foss) and log-transformed to SCS (Ali and Shook, 1980). Milk pH, adjusted for sample temperature, was obtained with a Crison Basic 25 electrode (Crison, Barcelona, Spain).

Milk Protein Fractions. Individual milk samples of all cows sampled (2 aliquots of 1 mL each per cow) were mixed with preservative (bronopol, 2-bromo-2-nitropropan-1,3-diol, 0.6:100 vol/vol) to prevent microbial development and frozen at -20°C in portable chilling devices immediately after collection and then stored at -80°C until analysis. Frozen individual milk aliquots were prepared following the method proposed by Bobe et al. (1998). The contents of the casein ($\alpha_{\text{S1-}}$, $\alpha_{\text{S2-}}$, β -, and κ -CN) and the whey protein (β -LG and α -LA) fractions were assessed separately for each of their major genetic variants by the reversed-phase HPLC method (Bonfatti et al., 2008). The phosphorylated form of the $\alpha_{\text{S1-}}$ -CN (with 9 phosphorylated serine residues instead of 8) was also obtained (Bonfatti et al., 2011b). The genetic variants of each protein fraction (A and B for κ -CN; A1, A2, and B for β -CN; A and B for β -LG) were recorded for each milk sample, and their concentrations were summed to obtain the total concentration of the protein fraction.

Cheese-Making Efficiency. Individual raw full-fat milk samples (2,000 mL) were processed within 20 h of milking using the model cheese-making procedure (1,500 mL of individual milk sample/vat) described by Stocco et al. (2018). Using this method, we were

able to collect 7 traits related to the cheese-making process. The first 3 traits were measures of percentage cheese yield (%CY) representing the ratios between the weight of the milk processed and the weight of the curd, the curd DM, and the water retained in the curd (%CY_{curd}, %CY_{solids}, and %CY_{water}, respectively):

$$\%CY_{\text{curd, solids, water}} = \frac{\text{weight of curd, DM, or water in curd (g)}}{\text{weight of milk (g)}} \times 100.$$

The next 3 traits were milk component recoveries in the curd (**REC**) representing the ratios between the weight of the fat, protein, and DM in the curd and the weight of the corresponding component in the milk (**REC**_{fat}, **REC**_{protein}, and **REC**_{solids}, respectively):

$$\%REC_{\text{fat, protein, solids}} = \frac{\text{weight of curd fat, protein, or DM (g)}}{\text{weight of milk fat, protein, or DM (g)}} \times 100.$$

We also estimated energy recovery in the curd (%; NRC, 2001). The 2 cheese-making efficiency indicators (**Ef-%CY**_{curd} and **Ef-%CY**_{solids}) were estimated as the ratio between the actual values of %CY_{curd} and %CY_{solids}, respectively, obtained in the laboratory from the model cheeses and the theoretical %CY (**Th-%CY**_{curd} and **Th-%CY**_{solids}; Supplemental Table S1; <https://doi.org/10.3168/jds.2018-14503>) calculated on the basis of milk composition using the predictive formulas proposed by Van Slyke and Price (1949):

$$\text{Ef-\%CY}_{\text{curd}} = \%CY_{\text{curd}} / \text{Th-\%CY}_{\text{curd}};$$

$$\text{Ef-\%CY}_{\text{solids}} = \%CY_{\text{solids}} / \text{Th-\%CY}_{\text{solids}}.$$

The daily cheese yield traits (**dCY**_{curd} and **dCY**_{solids}; kg/d) were calculated by multiplying the various %CY (curd and TS, respectively) by the daily milk yield (**dMY**; kg/d).

Statistical Analyses

The quantitative effects of the milk protein fractions (α_{S1-} , α_{S2-} , $\beta-$, and κ -CN; β -LG; and α -LA), expressed as percentage in milk, on the cheese-making traits were analyzed using the MIXED procedure of SAS (SAS Institute Inc., Cary, NC) according to the following linear model (**M-%milk**):

$$\begin{aligned} y_{ijklmnopqrstuvwxy} = & \mu + \text{dairy system}_i \\ & + \text{herd}_j(\text{dairy system})_i + \text{vat}_k + \text{DIM}_l + \text{parity}_m \\ & + \text{dMY}_n + \alpha_{S1\text{-CN}}_o + \alpha_{S1\text{-CN ph}}_p + \alpha_{S2\text{-CN}}_q + \beta\text{-CN}_r \\ & + \kappa\text{-CN}_s + \beta\text{-LG}_t + \alpha\text{-LA}_u + \beta\text{-CN-GT}_v \\ & + \kappa\text{-CN-GT}_w + \beta\text{-LG-GT}_x + e_{ijklmnopqrstuvwxy} \end{aligned}$$

where $y_{ijklmnopqrstuvwxy}$ is the observed trait (cheese-making traits: %CY, REC, Th-%CY, Ef-%CY, and dCY); μ is the overall mean; dairy system_{*i*} is the fixed effect of the *i*th class of dairy system (*i* = 1 to 4); herd_{*j*} (dairy system)_{*i*} is the random effect of the *j*th herd within the *i*th class of dairy system; vat_{*k*} is the fixed effect of the *k*th vat (*k* = 1 to 15 classes); DIM_{*l*} is the fixed effect of the *l*th 60-d class of lactation (*l* = 6 classes); parity_{*m*} is the fixed effect of the *m*th class of parity order (*m* = 1 to ≥ 5); dMY_{*n*} is the fixed effect of the *n*th class of dMY (*n* = 7 classes; this effect was not included in the statistical model when dCY traits were analyzed); $\alpha_{S1\text{-CN}}_o$ is the fixed effect of the *o*th class of $\alpha_{S1\text{-CN}}$ content (*o* = 7 classes); $\alpha_{S1\text{-CN ph}}_p$ is the fixed effect of the *p*th class of phosphorylated $\alpha_{S1\text{-CN}}$ content (*p* = 7 classes); $\alpha_{S2\text{-CN}}_q$ is the fixed effect of the *q*th class of $\alpha_{S2\text{-CN}}$ content (*q* = 7 classes); $\beta\text{-CN}_r$ is the fixed effect of the *r*th class of $\beta\text{-CN}$ content (*r* = 7 classes); $\kappa\text{-CN}_s$ is the fixed effect of the *s*th class of $\kappa\text{-CN}$ content (*s* = 7 classes); $\beta\text{-LG}_t$ is the fixed effect of the *t*th class of $\beta\text{-LG}$ content (*t* = 7 classes); $\alpha\text{-LA}_u$ is the fixed effect of the *u*th class of $\alpha\text{-LA}$ content (*u* = 7 classes); and $e_{ijklmnopqrstuvwxy}$ is the residual random error $\sim N(0, \sigma_e^2)$. Herd and residuals were assumed to be normally distributed with a mean of zero and variances of σ_h^2 and σ_e^2 , respectively. Although the effect of genetic variants is beyond the scope of this study, the protein genotypes (GT) of each fraction ($\beta\text{-CN}$, $\kappa\text{-CN}$, and $\beta\text{-LG}$) were included in the model to account for potential specific effects of the genetic variants and to avoid possible confusion with the quantitative effect of the protein fraction concentration. We neither report nor discuss the effect of genetic variants.

A second, qualitative model (**M-%Cas**) was used to study the effects of milk protein fractions expressed as proportions of total casein. In this model, the effect of the total casein content of milk (7 classes) replaced the effect of dMY as the fixed effect to distinguish between the quantitative effect of total casein and the qualitative effect of its composition. As casein is directly involved in the cheese-making process, we decided to express all the protein fractions as percentage of total casein instead of total milk protein, thereby treating the whey protein fractions as supplemental material rather than

constituents of cheese, and avoided negative multicollinearity between caseins and whey proteins.

Each of the 7 classes of protein fractions (as for total casein content and dMY) was determined on the basis of the distribution of the variables. Each individual class explained 0.5 SD of the variable; the fourth was centered on the mean value, and the first and seventh represented the tails of the distribution. Polynomial contrasts ($P < 0.05$) were obtained to evaluate the trends (linear, quadratic, and cubic relationships) of cheese-making traits for each protein fraction effect.

RESULTS

Descriptive Statistics

Descriptive statistics of milk gross composition, protein fractions [expressed as % (wt/wt) of milk and as % of total casein content], and cheese-making traits are presented in Table 1. Most of these traits were almost normally distributed, showing kurtosis and skewness values close to zero, with the exception of fat, fat recovery in the curd, lactose, and α_{S2} -CN (% of casein), which exhibited a tendency to a leptokurtic distribution (data not shown). Individual cows produced an average of 24.4 kg of milk/d with a composition of 4.22% fat, 3.71% protein, and 4.85% lactose. Casein content was 2.89% and had a ratio to protein of 0.78 (casein number), whereas the ratio to fat had a mean value of 0.68.

As expected, the predominant casein fraction was β -CN, which, together with α_{S1} -CN, represented about three-quarters of total casein content. The remainder was shared by α_{S2} -CN and κ -CN, with a small presence of α_{S1} -CN-phosphorylated form (**Ph**). Analysis of the whey protein fractions revealed there to be approximately 3 times the amount of β -LG as α -LA, although the variability of these 2 fractions was similar when expressed as coefficient of variation (**CV**). The variability (in SD) of each protein was positively correlated with their quantities also in the case of caseins.

The mean value of %CY_{curd} was 15.0% and was almost equally distributed between TS and water in the curd (%CY_{solids} = 7.22%; %CY_{water} = 7.80%). The observed trait was slightly lower than the Th-%CY_{curd}, resulting in an Ef-%CY_{curd} equal to 0.99. The average composition of fresh curd was 19.5% protein, 26.2% fat, and 48.4% TS (data not shown), resulting from average recoveries in the curd of 78.1% for protein, 89.9% for fat, and 52.0% for TS. Protein recovery in the curd had a lower standard deviation than the other REC, but the CV value was higher than the casein index (2.4% for REC_{protein} vs. 1.6% for the casein index) with respect to both casein losses in the whey and retention of whey proteins in the curd.

Daily cheese production of individual cows was on average 3.63 kg/d for dCY_{curd} and 1.74 for dCY_{solids}. These traits had a higher CV (about 33%) than %CY in direct relation to the high variability in daily milk production.

Cheese-Making

Tables 2, 3, and 4 and Supplemental Table S2 (<https://doi.org/10.3168/jds.2018-14503>) show the results of the ANOVA (M-%milk and M-%cas) for the %CY, REC, Ef-%CY, and dCY groups of traits, respectively, and of the contrasts (linear, quadratic, and cubic) for total casein content and each protein fraction. The effects related to individual cows (DIM and order of parity), farm (individual herd and dairy system), and cheese-making vat were included in both the M-%milk and M-%cas models to correct the cheese-making traits for these sources of variation. However, we did not observe any relevant significant differences between the 2 models (Tables 2, 3, and 4 and Supplemental Table S2; <https://doi.org/10.3168/jds.2018-14503>). Effects of protein genotypes (β -CN, κ -CN, and β -LG) were also included in the models so that we could analyze the effects of the amounts of individual protein fractions in milk and their proportions in casein independently of their potential genetic variants.

Moving on to total casein content (model M-%cas), the increment in this component had a favorable effect on both Ef-%CY_{curd} and Ef-%CY_{solids} due to its linear positive relationship with %CY and REC traits (Figure 1). As expected, the effects of each fraction expressed as percentage in milk were more relevant for all the %CY traits than the effects observed in M-%cas, as the former traits are closely related to total protein quantity in each processed milk sample. In all the REC traits, the proportions of protein fractions to total casein (model M-%cas) were more important than those in %CY, although the amounts of proteins in milk were also found to have a greater influence on REC traits (M-%milk).

Among casein fractions, we found α_{S1} -CN to be very important for the 3 %CY traits when considered as percentage of milk (M-%milk; Figure 2a). This protein, together with β -CN the most important quantitatively (Table 1), produced a linear increase in the 3 cheese yields, with %CY_{curd} increasing by 3.0 percentage points (20% of the average value) from the first to the last class of α_{S1} -CN least squares means. Although the recoveries of individual and overall components [Figure 3a and Supplemental Figure S1a (<https://doi.org/10.3168/jds.2018-14503>), respectively] were also linearly influenced by the increase in this casein fraction in milk, α_{S1} -CN increased cheese-making efficiency only with respect to

Table 1. Descriptive statistics of milk yield, protein fraction (% in milk and % on casein), cheese-making traits, and daily cheese production¹

| Trait | Mean | SD | P5 | P95 |
|-------------------------------------|------|------|------|------|
| Daily milk yield, kg/d | 24.4 | 7.9 | 12.3 | 37.9 |
| Fat, % | 4.22 | 0.73 | 3.14 | 5.42 |
| Protein, % | 3.71 | 0.44 | 3.03 | 4.43 |
| Casein, % | 2.89 | 0.33 | 2.38 | 3.44 |
| Casein index, % | 0.78 | 0.01 | 0.76 | 0.80 |
| Lactose, % | 4.85 | 0.20 | 4.50 | 5.13 |
| SCS, unit | 2.98 | 1.86 | 0.21 | 6.20 |
| Milk protein fraction, % | | | | |
| In milk | | | | |
| α_{S1} -CN | 0.95 | 0.13 | 0.76 | 1.18 |
| α_{S1} -CN-Ph ² | 0.05 | 0.02 | 0.02 | 0.10 |
| α_{S2} -CN | 0.34 | 0.06 | 0.25 | 0.44 |
| β -CN | 1.19 | 0.15 | 0.95 | 1.44 |
| κ -CN | 0.35 | 0.07 | 0.23 | 0.46 |
| β -LG | 0.33 | 0.07 | 0.22 | 0.45 |
| α -LA | 0.09 | 0.02 | 0.06 | 0.12 |
| On casein | | | | |
| α_{S1} -CN | 33.0 | 2.1 | 29.6 | 36.7 |
| α_{S1} -CN-Ph ² | 1.86 | 0.79 | 0.60 | 3.26 |
| α_{S2} -CN | 11.8 | 1.5 | 9.7 | 14.4 |
| β -CN | 41.2 | 3.0 | 37.0 | 46.7 |
| κ -CN | 12.2 | 1.8 | 8.8 | 14.6 |
| β -LG | 11.2 | 2.0 | 8.2 | 14.5 |
| α -LA | 3.02 | 0.65 | 1.95 | 4.09 |
| Cheese yield (CY), % | | | | |
| %CY _{curd} | 15.0 | 1.89 | 12.0 | 18.3 |
| %CY _{solids} | 7.22 | 0.93 | 5.77 | 8.80 |
| %CY _{water} | 7.80 | 1.28 | 5.85 | 9.95 |
| Theoretical %CY (Th-%CY) | | | | |
| Th-%CY _{curd} | 15.2 | 1.8 | 12.3 | 18.4 |
| Th-%CY _{solids} | 7.30 | 0.88 | 5.89 | 8.85 |
| Efficiency of %CY (Ef-%CY) | | | | |
| Ef-%CY _{curd} | 0.99 | 0.09 | 0.85 | 1.14 |
| Ef-%CY _{solids} | 0.99 | 0.06 | 0.90 | 1.07 |
| Nutrient recovery (REC), % | | | | |
| REC _{protein} | 78.1 | 2.41 | 74.0 | 81.9 |
| REC _{fat} | 89.9 | 3.58 | 82.6 | 94.5 |
| REC _{solids} | 52.0 | 3.58 | 46.1 | 58.1 |
| REC _{energy} | 67.3 | 3.32 | 61.8 | 72.5 |
| Daily cheese production (dCY), kg/d | | | | |
| dCY _{curd} | 3.63 | 1.17 | 1.80 | 5.73 |
| dCY _{solids} | 1.74 | 0.57 | 0.87 | 2.72 |
| dCY _{water} | 1.88 | 0.63 | 0.92 | 3.04 |

¹P5 = 5th percentile; P95 = 95th percentile.

²The α_{S1} -CN fraction with 9 instead of 8 phosphorylated serine residues.

Ef-%CY_{solids} (M-%milk; Table 3; Figure 4a). The effect of its phosphorylated form was positive but not very important (Figures 2b, 3b, 4b, and 5b). Moreover, the amount of α_{S1} -CN-Ph in total casein content reduced milk protein recovery in the curd (Figure 3b).

Unlike α_{S1} -CN, the effect of the α_{S2} -CN fraction was mostly significant when expressed as percentage of total casein and lowered cheese-making efficiency in terms of solids yield (Ef-%CY_{solids}; Figure 4c). The effect of α_{S2} -CN on all %CY traits was linearly negative in the M-%cas model, whereas a quadratic pattern for %CY_{curd}, %CY_{solids}, and REC_{solids} was found in the M-%milk model. The REC_{protein} and energy recovery in the

curd were also negatively influenced by the increase in the content of this protein fraction in milk and in casein [Figure 2c and Supplemental Figure S1c (<https://doi.org/10.3168/jds.2018-14503>)], respectively].

When expressed as percentage in milk, β -CN was the protein fraction with the greatest effect on %CY_{curd} (Figure 2d) and had strong positive importance for all 3 observed %CY traits, causing, in particular, an increase in water retention in the curd (Figure 2d). Although β -CN had a negative influence on REC_{fat} (Figure 3d), the strong effect of this protein fraction on all the %CY traits was due to its positive relationship with REC_{protein} and REC_{solids} [Figure 3d and Supple-

mental Figure S1d (<https://doi.org/10.3168/jds.2018-14503>), respectively]. In qualitative terms, we did not find any strong relationships between β -CN and the observed cheese-making traits in the M-%cas model, although this fraction influenced the efficiency of %CY with significant quadratic (with intermediate optimum value) and cubic (decreasing) contrasts for Ef-%CY_{curd} and Ef-%CY_{solids}, respectively (Figure 4d).

The κ -CN fraction expressed as percentage in milk was important for all the %CY traits, exhibiting linear and positive relationships (Figure 2e); this and α_{S1} -CN were the only fractions positively influencing Ef-%CY_{solids}. The results explaining the relationship between curd recoveries and the κ -CN fraction were more heterogeneous. Although this protein fraction had a very clear and favorable influence on the recovery of

Table 2. Analysis of variance (model M-%milk and model M-%cas¹; *F*-value and significance) for cheese yield (%CY; curd, solids, and retained water) and results of contrasts (linear, quadratic, and cubic; *F*-value and significance) for total casein content and milk protein fractions

| Effect | %CY _{curd} | | %CY _{solids} | | %CY _{water} | |
|-----------------------------------|---------------------|----------|-----------------------|----------|----------------------|----------|
| | M-%milk | M-%cas | M-%milk | M-%cas | M-%milk | M-%cas |
| Dairy system | 1.5 | 1.8 | 4.7** | 4.2** | 0.6 | 0.5 |
| Herd ² | 35 | 34 | 19 | 19 | 47 | 45 |
| Vat | 1.9* | 1.6 | 2.5** | 2.2** | 1.7* | 1.7* |
| DIM, d | 2.3* | 1.9 | 0.6 | 0.4 | 3.5** | 2.9* |
| Parity | 1.7 | 1.5 | 2.3 | 2.2 | 3.2* | 1.7 |
| Protein fraction genotype | | | | | | |
| β -CN | 0.4 | 0.2 | 1.2 | 1.0 | 0.7 | 0.3 |
| κ -CN | 1.6 | 1.5 | 0.3 | 0.4 | 3.6* | 2.4 |
| β -LG | 10.7*** | 6.7** | 2.4 | 1.0 | 6.8** | 4.6* |
| Daily milk yield, kg/d | 0.9 | — | 0.4 | — | 1.4 | — |
| Casein | | | | | | |
| Linear | — | 791.7*** | — | 441.2*** | — | 410.6*** |
| Quadratic | — | 0.4 | — | 9.0** | — | 2.3 |
| Cubic | — | 3.2 | — | 1.9 | — | 2.7 |
| α_{S1} -CN | | | | | | |
| Linear | 174.1*** | 3.9* | 123.6*** | 1.6 | 62.1*** | 0.1 |
| Quadratic | 0.0 | 0.0 | 1.5 | 0.7 | 0.9 | 0.2 |
| Cubic | 1.4 | 0.6 | 3.8 | 0.0 | 0.0 | 0.7 |
| α_{S1} -CN-Ph ³ | | | | | | |
| Linear | 33.7*** | 3.8 | 18.8*** | 2.7 | 16.5*** | 0.5 |
| Quadratic | 1.6 | 3.1 | 0.1 | 0.7 | 3.1 | 2.5 |
| Cubic | 0.8 | 0.2 | 1.9 | 0.6 | 0.0 | 0.6 |
| α_{S2} -CN | | | | | | |
| Linear | 0.6 | 16.0*** | 2.7 | 13.1*** | 0.4 | 10.6** |
| Quadratic | 5.2* | 3.2 | 9.1** | 5.9* | 0.0 | 0.1 |
| Cubic | 0.4 | 4.8* | 0.2 | 1.7 | 0.5 | 5.2* |
| β -CN | | | | | | |
| Linear | 124.6*** | 0.3 | 29.1*** | 0.9 | 98.8*** | 0.0 |
| Quadratic | 1.4 | 0.0 | 3.2 | 1.9 | 0.0 | 3.9* |
| Cubic | 0.5 | 1.4 | 0.0 | 0.2 | 2.0 | 0.1 |
| κ -CN | | | | | | |
| Linear | 34.2*** | 0.0 | 34.8*** | 2.2 | 16.0*** | 1.4 |
| Quadratic | 0.2 | 0.0 | 0.3 | 0.2 | 0.1 | 0.0 |
| Cubic | 8.0 | 0.9 | 3.3 | 1.3 | 7.3 | 0.0 |
| β -LG | | | | | | |
| Linear | 37.0*** | 43.4*** | 14.2*** | 18.5*** | 25.3*** | 23.7*** |
| Quadratic | 1.9 | 1.4 | 0.4 | 0.2 | 4.1 | 3.2 |
| Cubic | 0.4 | 0.0 | 0.0 | 0.1 | 0.5 | 0.1 |
| α -LA | | | | | | |
| Linear | 4.2* | 4.3* | 1.1 | 0.6 | 4.2* | 5.5* |
| Quadratic | 1.4 | 1.5 | 2.4 | 1.4 | 0.0 | 0.2 |
| Cubic | 0.1 | 1.0 | 0.4 | 0.5 | 3.7 | 1.2 |
| Root mean squared error | 0.98 | 1.01 | 0.61 | 0.61 | 0.72 | 0.74 |

¹Proteins expressed as content in milk (M-%milk) or as percentage of total casein content (M-%cas).

²The variance of herd within dairy system is expressed as ratio with total variance (herd plus residual).

³The α_{S1} -CN fraction with 9 instead of 8 phosphorylated serine residues.

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

fat in the curd in both the M-%milk and the M-%cas models, it negatively influenced REC_{protein} , especially when expressed as percentage of total casein (Figure 3e).

β -Lactoglobulin had a strong negative influence on all the cheese-making traits in both the M-%milk and M-%cas models, particularly for $Ef\text{-}\%CY_{\text{curd}}$ and $Ef\text{-}\%CY_{\text{solids}}$ (Figure 4f). Contrary to what we observed for β -LG, α -LA was the protein fraction with the least influence on the $\%CY$ in both statistical models. Despite the significance of the effect of α -LA on $\%CY_{\text{curd}}$ and CY_{water} in the M-%milk model, the variability of these traits in relation to this effect was mostly negligible. Moreover, the only notable effect of α -LA was that it

fat in the curd in both the M-%milk and the M-%cas models, it negatively influenced REC_{protein} , especially when expressed as percentage of total casein (Figure 3e).

Table 3. Analysis of variance (model M-%milk and model M-%cas¹; F -value and significance) for milk protein and fat recovery (REC) in the curd and for efficiency of cheese yield ($Ef\text{-}\%CY$), together with results of contrasts (linear, quadratic, and cubic; F -value and significance) for total casein content and milk protein fractions

| Effect | REC_{protein} | | REC_{fat} | | $Ef\text{-}\%CY_{\text{curd}}$ | | $Ef\text{-}\%CY_{\text{solids}}$ | |
|--|------------------------|----------|--------------------|---------|--------------------------------|---------|----------------------------------|---------|
| | M-%milk | M-%cas | M-%milk | M-%cas | M-%milk | M-%cas | M-%milk | M-%cas |
| Dairy system | 0.5 | 1.4 | 1.9 | 2.2 | 0.3 | 0.5 | 4.7** | 5.2*** |
| Herd ² | 41 | 41 | 31 | 30 | 42 | 42 | 28 | 28 |
| Vat | 1.3 | 1.2 | 1.2 | 1.7 | 2.1* | 2.0* | 1.5 | 1.7 |
| DIM, d | 3.0** | 3.7** | 10.5*** | 12.8*** | 6.8*** | 6.1*** | 2.9* | 3.0* |
| Parity | 21.6*** | 16.8*** | 0.7 | 0.8 | 2.1 | 1.0 | 0.9 | 1.1 |
| Protein fractions | | | | | | | | |
| genotype | | | | | | | | |
| β -CN | 10.0*** | 8.4*** | 1.4 | 0.9 | 1.8 | 2.0 | 1.0 | 0.6 |
| κ -CN | 3.7* | 5.5** | 2.1 | 1.1 | 6.5** | 5.1** | 0.5 | 0.0 |
| β -LG | 232.1*** | 215.3*** | 2.8 | 4.5* | 2.2 | 2.0 | 0.7 | 0.6 |
| Daily milk yield, kg/d | 3.9*** | — | 1.2 | — | 1.5 | — | 0.3 | — |
| Casein | | | | | | | | |
| Linear | — | 55.0*** | — | 27.0*** | — | 28.7*** | — | 15.8*** |
| Quadratic | — | 0.4 | — | 2.8 | — | 14.9*** | — | 1.9 |
| Cubic | — | 0.0 | — | 0.2 | — | 0.5 | — | 0.5 |
| α_{S1} -CN | | | | | | | | |
| Linear | 102.0*** | 0.4 | 15.8*** | 1.7 | 0.8 | 0.1 | 5.9* | 0.7 |
| Quadratic | 0.0 | 0.4 | 0.1 | 0.0 | 0.0 | 1.4 | 2.3 | 0.0 |
| Cubic | 0.0 | 0.0 | 0.1 | 0.0 | 1.2 | 0.2 | 0.3 | 0.5 |
| α_{S1} -CN-Ph ³ | | | | | | | | |
| Linear | 0.0 | 8.5** | 5.3* | 2.5 | 3.0 | 0.1 | 3.8 | 0.8 |
| Quadratic | 0.5 | 1.0 | 0.1 | 0.3 | 0.9 | 0.2 | 0.1 | 0.0 |
| Cubic | 0.1 | 2.1 | 0.3 | 0.9 | 0.4 | 0.7 | 0.1 | 0.0 |
| α_{S2} -CN | | | | | | | | |
| Linear | 36.0*** | 61.3*** | 0.3 | 0.7 | 1.3 | 0.4 | 2.0 | 4.6* |
| Quadratic | 0.3 | 0.8 | 0.0 | 0.2 | 3.5 | 1.7 | 1.0 | 0.2 |
| Cubic | 6.0* | 5.2* | 0.1 | 2.2 | 0.2 | 0.9 | 0.1 | 0.4 |
| β -CN | | | | | | | | |
| Linear | 119.6*** | 0.2 | 4.7* | 0.8 | 19.5*** | 0.4 | 0.0 | 2.4 |
| Quadratic | 0.1 | 0.0 | 1.1 | 0.5 | 7.0** | 4.8* | 1.9 | 0.1 |
| Cubic | 1.5 | 0.1 | 0.0 | 4.0* | 1.7 | 4.2* | 0.0 | 3.9* |
| κ -CN | | | | | | | | |
| Linear | 9.1** | 44.9*** | 35.6*** | 15.3*** | 0.6 | 2.3 | 10.0** | 0.5 |
| Quadratic | 0.0 | 1.5 | 2.0 | 0.5 | 2.3 | 0.4 | 5.3* | 1.3 |
| Cubic | 0.6 | 0.5 | 3.5 | 0.3 | 1.6 | 0.7 | 0.9 | 0.2 |
| β -LG | | | | | | | | |
| Linear | 171.9*** | 182.4*** | 26.7*** | 25.7*** | 20.0*** | 19.3*** | 23.7*** | 28.0*** |
| Quadratic | 2.1 | 0.2 | 0.1 | 5.3* | 0.8 | 1.3 | 0.7 | 9.0** |
| Cubic | 2.7 | 2.4 | 0.0 | 0.1 | 1.1 | 0.0 | 0.2 | 0.3 |
| α -LA | | | | | | | | |
| Linear | 2.7 | 1.8 | 7.1** | 6.3* | 3.2 | 4.6* | 2.3 | 1.4 |
| Quadratic | 3.5 | 5.7* | 0.1 | 0.0 | 0.9 | 0.5 | 0.0 | 0.6 |
| Cubic | 0.2 | 1.3 | 0.4 | 0.1 | 3.9* | 0.6 | 1.4 | 2.0 |
| Root mean squared error | 1.34 | 1.33 | 2.67 | 2.65 | 0.07 | 0.07 | 0.05 | 0.05 |

¹Proteins expressed as content in milk (M-%milk) or as percentage of total casein content (M-%cas).

²The variance of herd within dairy system and season is expressed as ratio with total variance (herd plus residual).

³The α_{S1} -CN fraction with 9 instead of 8 phosphorylated serine residues.

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Table 4. Analysis of variance (model M-%milk and model M-%cas¹; *F*-value and significance) for daily cheese production (dCY) and results of contrasts (linear, quadratic, and cubic; *F*-value and significance) for total casein content and milk protein fractions

| Effect | dCY _{curd} | | dCY _{solids} | |
|-------------------------------------|---------------------|---------|-----------------------|---------|
| | M-%milk | M-%cas | M-%milk | M-%cas |
| Dairy system | 16.2*** | 16.1*** | 19.7*** | 18.8*** |
| Herd ² | 39 | 40 | 35 | 37 |
| Vat | 1.2 | 1.2 | 1.4 | 1.6 |
| DIM, d | 22.8*** | 20.2*** | 18.4*** | 17.4*** |
| Parity | 19.4*** | 21.2*** | 23.5*** | 25.4*** |
| Protein fractions genotype | | | | |
| β-CN | 1.9 | 3.3* | 0.6 | 1.7 |
| κ-CN | 1.2 | 0.8 | 0.7 | 0.4 |
| β-LG | 1.8 | 1.4 | 1.4 | 0.8 |
| Casein | | | | |
| Linear | — | 4.5* | — | 1.1 |
| Quadratic | — | 4.9* | — | 1.5 |
| Cubic | — | 0.5 | — | 0.0 |
| α _{S1} -CN | | | | |
| Linear | 21.8*** | 6.5* | 23.9*** | 7.2** |
| Quadratic | 0.2 | 4.8* | 0.0 | 4.2* |
| Cubic | 0.0 | 0.1 | 0.2 | 0.0 |
| α _{S1} -CN-Ph ³ | | | | |
| Linear | 4.1* | 2.2 | 2.6 | 1.9 |
| Quadratic | 0.7 | 0.8 | 0.0 | 0.3 |
| Cubic | 0.1 | 1.1 | 0.0 | 1.8 |
| α _{S2} -CN | | | | |
| Linear | 24.3*** | 9.0** | 26.1*** | 8.9** |
| Quadratic | 1.5 | 0.1 | 0.2 | 0.0 |
| Cubic | 0.9 | 1.3 | 0.9 | 0.6 |
| β-CN | | | | |
| Linear | 0.0 | 2.3 | 2.0 | 1.5 |
| Quadratic | 3.0 | 0.5 | 1.3 | 0.0 |
| Cubic | 0.0 | 0.0 | 0.8 | 0.1 |
| κ-CN | | | | |
| Linear | 0.0 | 1.1 | 0.2 | 2.5 |
| Quadratic | 1.2 | 0.1 | 2.2 | 0.6 |
| Cubic | 2.7 | 0.5 | 2.0 | 0.6 |
| β-LG | | | | |
| Linear | 19.1*** | 20.2*** | 13.9*** | 13.7*** |
| Quadratic | 0.2 | 0.1 | 0.1 | 0.0 |
| Cubic | 2.1 | 0.0 | 1.5 | 0.3 |
| α-LA | | | | |
| Linear | 15.0*** | 22.8*** | 12.9*** | 22.1*** |
| Quadratic | 1.1 | 0.8 | 1.9 | 0.7 |
| Cubic | 0.2 | 0.1 | 0.0 | 0.7 |
| Root mean squared error | 0.72 | 0.72 | 0.36 | 0.36 |

¹Proteins expressed as content in milk (M-%milk) or as percentage of total casein content (M-%cas).

²The variance of herd within dairy system and season is expressed as ratio with total variance (herd plus residual).

³The α_{S1}-CN fraction with 9 instead of 8 phosphorylated serine residues.

P* < 0.05; *P* < 0.01; ****P* < 0.001.

had a favorable effect on REC_{fat}, whether expressed as percentage in milk or in relation to total casein (Figures 3g).

When individual dCY traits were assessed, we found α_{S1}-CN to have positive effects and α_{S2}-CN to have negative effects (Figure 5a and c), results that reflect, in part, what we observed for %CY. β-Lactoglobulin also had a negative effect, whereas α-LA, although of negligible importance for all the %CY traits, had a positive linear effect on dCY traits, whether expressed

as percentage in milk or with respect to casein (Figure 5g).

DISCUSSION

Effect of Total Casein Content

Researchers who have studied the relationship between casein or protein (or their fractions) and cheese yields at the individual cow level (Marziali and Ng-

Kwai-Hang, 1986; Wedholm et al., 2006) have not attended to the effects of proteins on the efficiency of cheese production and individual milk component recoveries in the curd. Furthermore, results are affected

by laboratory conditions and procedures that do not always simulate the cheese-making process in field conditions—for example, the use of a centrifuge to separate the curd from the whey (Hallén et al., 2010). It is widely

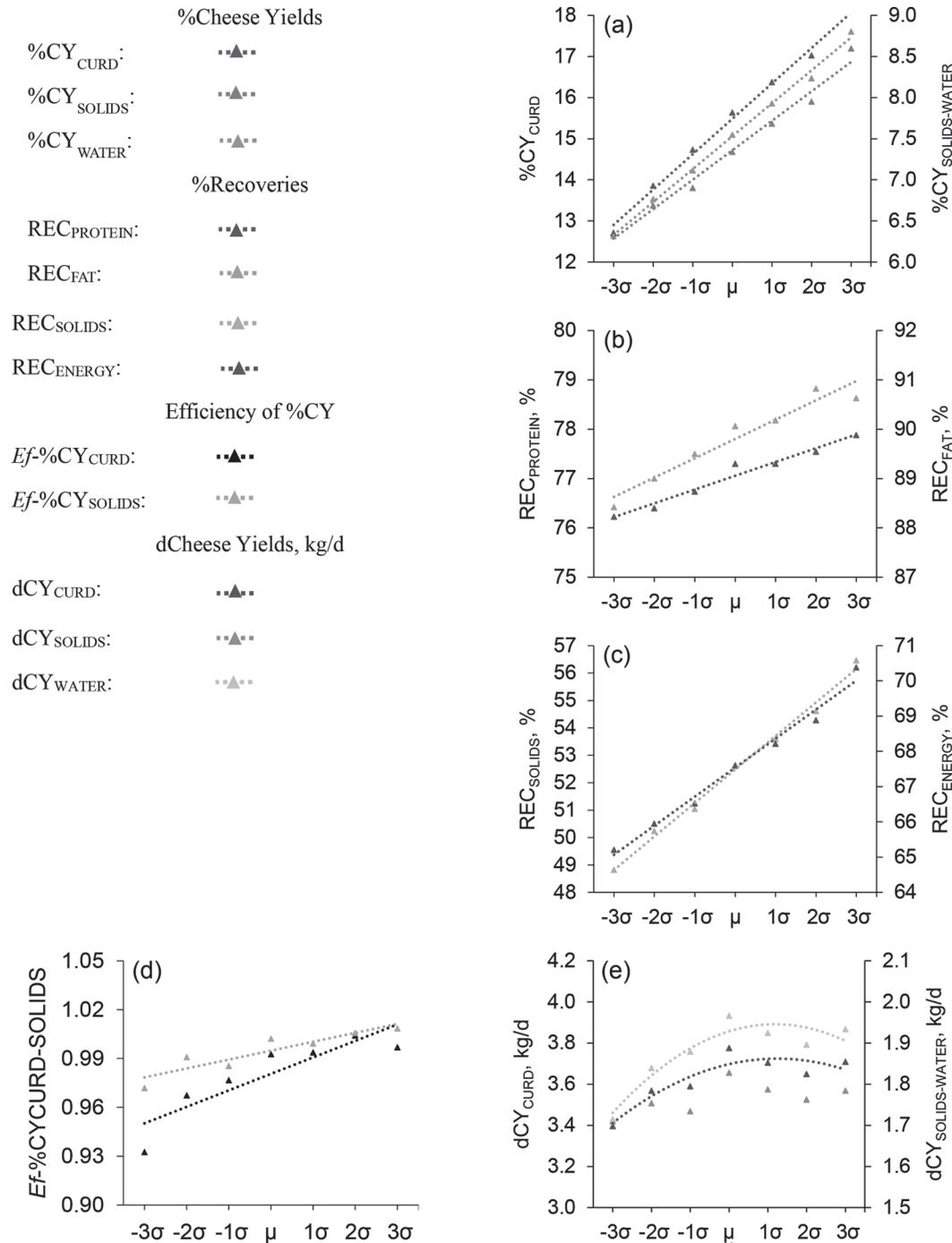


Figure 1. Effect of casein content on (a) cheese yields ($\%CY$; $\%CY_{CURD}$, $\%CY_{SOLIDS}$, and $\%CY_{WATER}$), (b) milk components recovery (REC) in the curd ($REC_{PROTEIN}$ and REC_{FAT}), (c) milk REC in the curd (REC_{SOLIDS} and REC_{ENERGY}), (d) efficiency of cheese yield ($Ef-\%CY$; $Ef-\%CY_{CURD}$ and $Ef-\%CY_{SOLIDS}$), and (e) daily cheese yields (dCY ; dCY_{CURD} , dCY_{SOLIDS} , and dCY_{WATER}). Results of the polynomial contrasts have been reported as the response curve (linear, quadratic, or cubic) of the variables across classes of milk casein. For each protein fraction, the class 4 represents the central class (-0.25 to $+0.25$ SD with respect to the mean). The amplitude of the other classes is 0.5 SD. Color version available online.

recognized that the effects of casein on cheese-making can be influenced by processing conditions and by the category of cheese being produced. The microstructure

of the coagulum, and afterward the structure of the formed curd, markedly affect rheological and syneretic properties, and subsequently the $\%CY_{\text{curd}}$, due to the

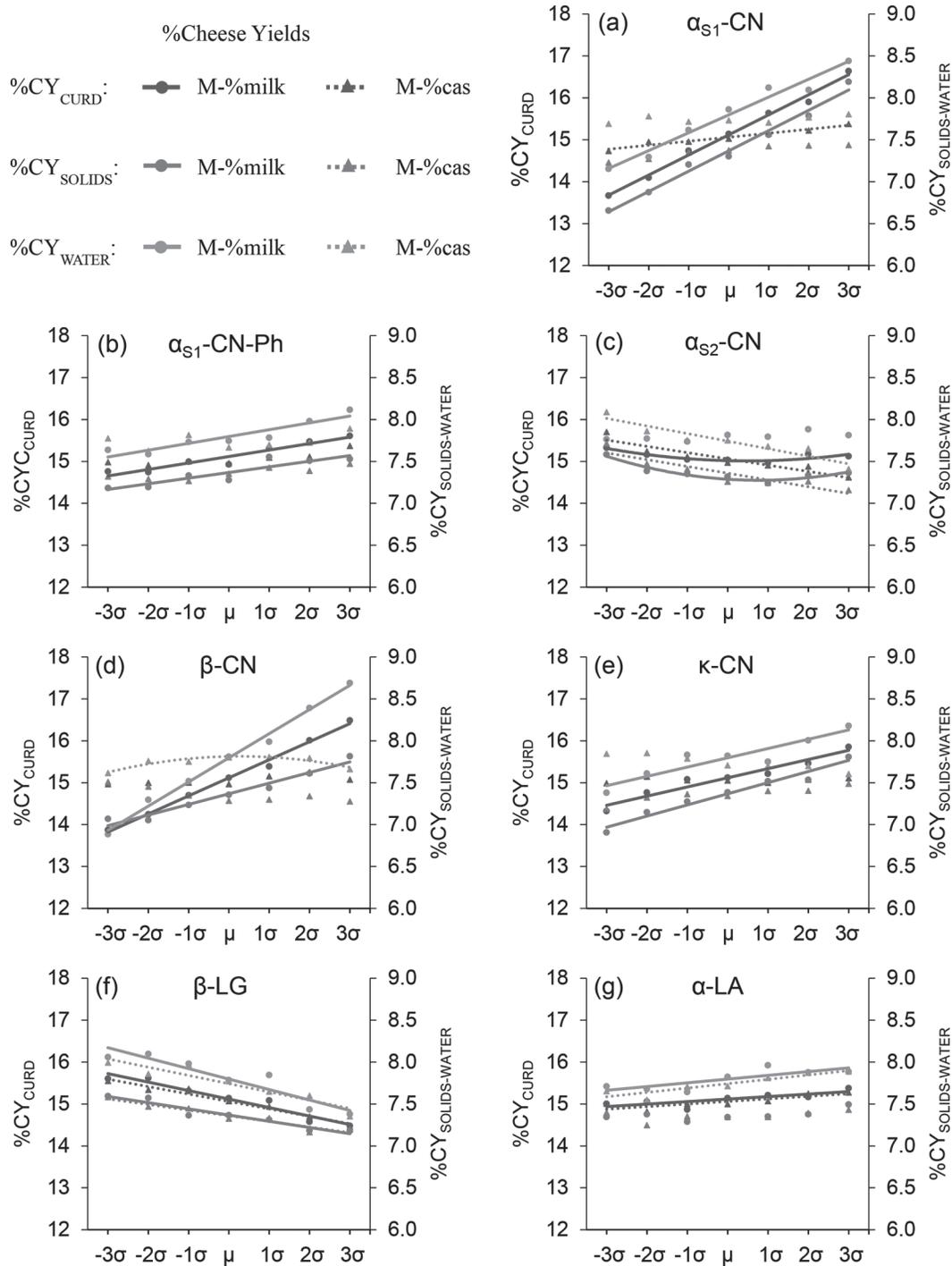


Figure 2. Effect of milk protein fractions content on cheese yields ($\%CY$; $\%CY_{\text{curd}}$, $\%CY_{\text{solids}}$ and $\%CY_{\text{water}}$): (a) α_{S1} -CN; (b) α_{S1} -CN-Ph (the α_{S1} -CN fraction with 9 instead of 8 phosphorylated serine residues); (c) α_{S2} -CN; (d) β -CN; (e) κ -CN; (f) β -LG; and (g) α -LA. Results of the polynomial contrasts have been reported as the response curve (linear, quadratic, or cubic) of the variables across classes of milk proteins: solid lines represent data from the M-%milk model, and dotted lines represent data from the M-%cas model [proteins expressed as content in milk (M-%milk) or as percentage of total casein content (M-%cas)]. For each protein fraction, the class 4 represents the central class (-0.25 to $+0.25$ SD with respect to the mean). The amplitude of the other classes is 0.5 SD. Color version available online.

different recoveries of milk components (protein and fat) in the curd (Guinee et al., 1995) and the amount of water retained. In the present study, we examined

the pattern of %CY, REC, Ef-%CY, and dCY traits in relation to total casein content and all the protein fractions expressed as percentage in milk and as percentage

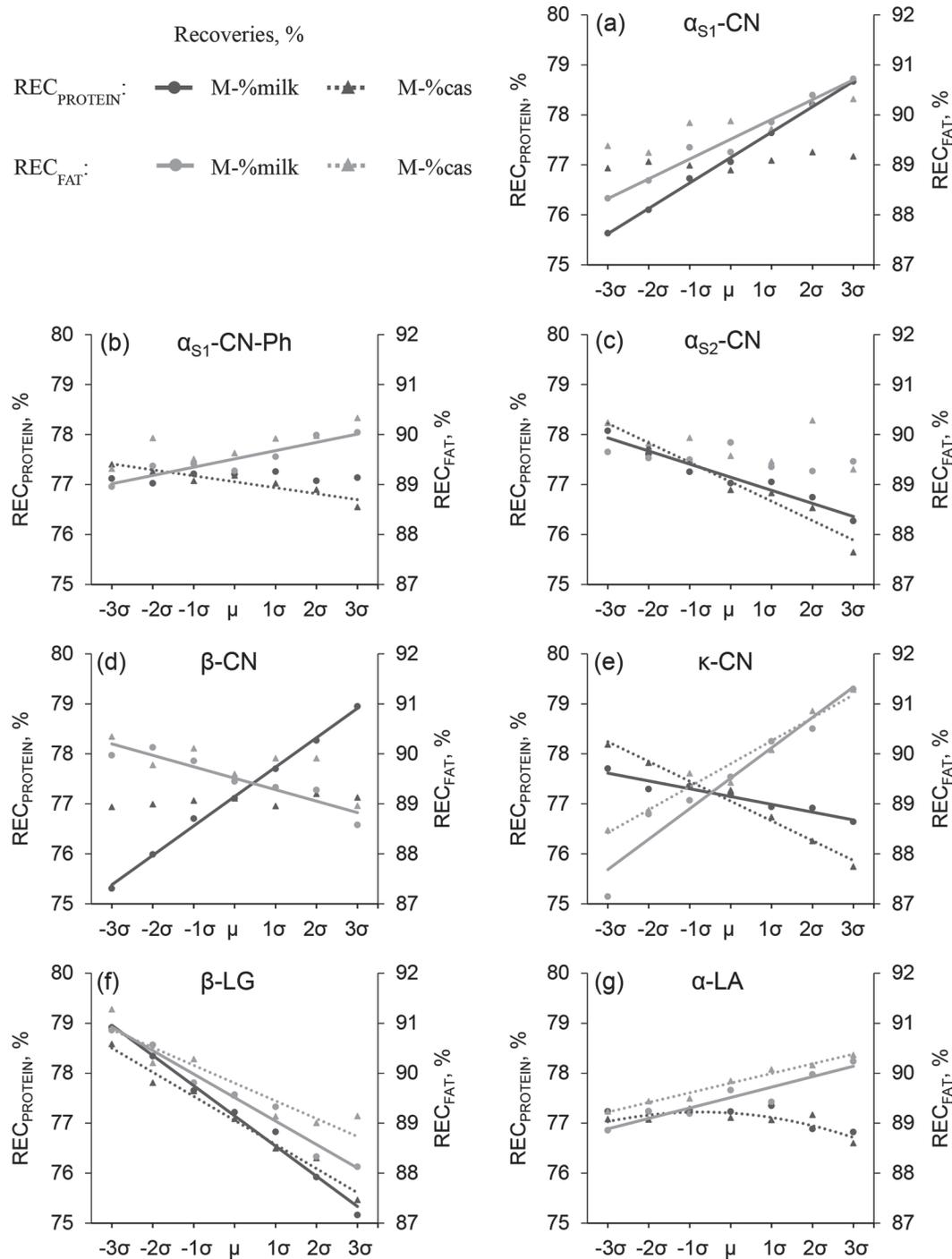


Figure 3. Effect of milk protein and fat recovery (REC) in the curd (REC_{protein} and REC_{fat}). (a) α_{S1} -CN; (b) α_{S1} -CN-Ph (the α_{S1} -CN fraction with 9 instead of 8 phosphorylated serine residues); (c) α_{S2} -CN; (d) β -CN; (e) κ -CN; (f) β -LG; and (g) α -LA. Results of the polynomial contrasts have been reported as the response curve (linear, quadratic, or cubic) of the variables across classes of milk proteins: solid lines represent data from the M-%milk model, and dotted lines represent data from the M-%cas model [proteins expressed as content in milk (M-%milk) or as percentage of total casein content (M-%cas)]. For each protein fraction, the class 4 represents the central class (-0.25 to +0.25 SD with respect to the mean). The amplitude of the other classes is 0.5 SD. Color version available online.

of total casein. As expected, casein and its fractions (expressed as % of milk) had a positive linear effect on almost all the observed %CY traits, and we found that

an increment in milk casein of 0.1 percentage points contributed to a linear increase of 0.61% in %CY_{curd}, 0.27% in %CY_{solids}, and 0.33% in %CY_{water} (M-%cas;

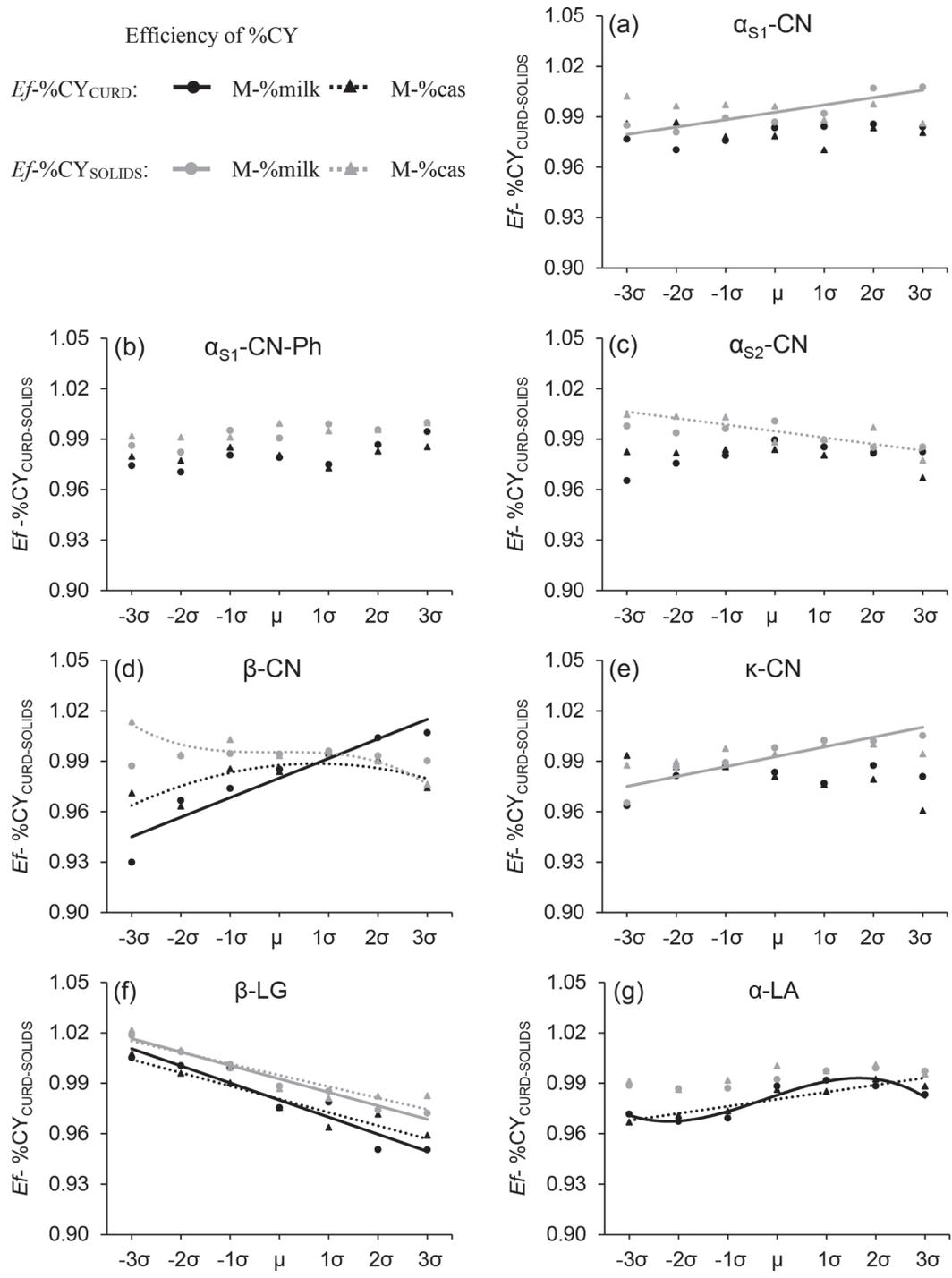


Figure 4. Effect of milk protein fractions content on efficiency of cheese yield ($Ef\text{-}\%CY$; $Ef\text{-}\%CY_{curd}$ and $Ef\text{-}\%CY_{solids}$). (a) $\alpha_{S1}\text{-CN}$; (b) $\alpha_{S1}\text{-CN-Ph}$ (the $\alpha_{S1}\text{-CN}$ fraction with 9 instead of 8 phosphorylated serine residues); (c) $\alpha_{S2}\text{-CN}$; (d) $\beta\text{-CN}$; (e) $\kappa\text{-CN}$; (f) $\beta\text{-LG}$; and (g) $\alpha\text{-LA}$. Results of the polynomial contrasts have been reported as the response curve (linear, quadratic, or cubic) of the variables across classes of milk proteins: solid lines represent data from the M-%milk model, and dotted lines represent data from the M-%cas model [proteins expressed as content in milk (M-%milk) or as percentage of total casein content (M-%cas)]. For each protein fraction, the class 4 represents the central class (-0.25 to +0.25 SD with respect to the mean). The amplitude of the other classes is 0.5 SD.

Figure 1). Also expected was the effect of proteins on individual dCY, which can be considered a measure of the efficiency of cows' cheese production at the herd

level. These traits are closely related to a combination of daily milk production, amounts of components in milk, recoveries of milk nutrients in the curd, and

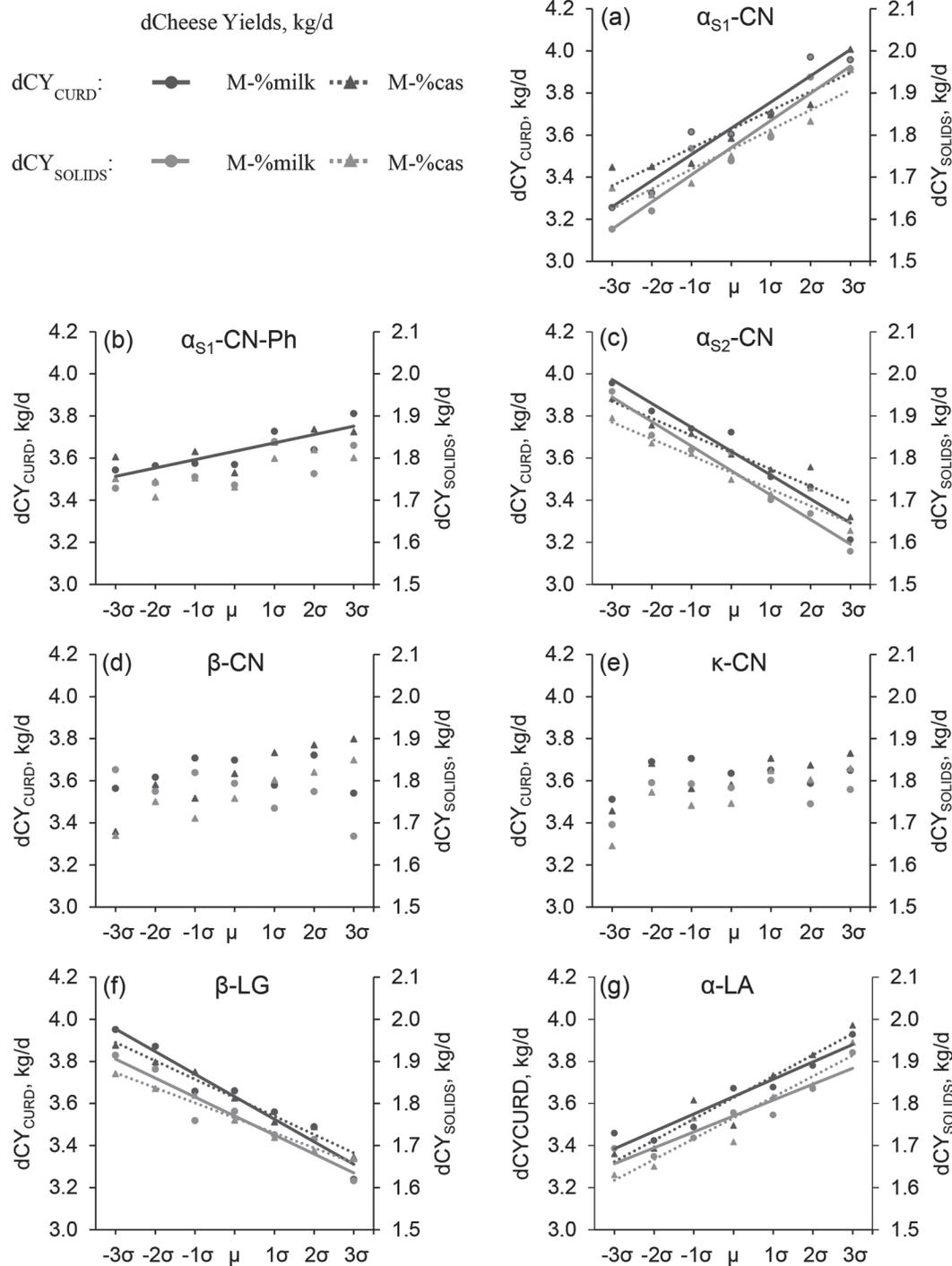


Figure 5. Effect of milk protein fractions content on daily cheese yields (dCY; dCY_{CURD} and dCY_{SOLIDS}). (a) α_{S1} -CN; (b) α_{S1} -CN-Ph (the α_{S1} -CN fraction with 9 instead of 8 phosphorylated serine residues); (c) α_{S2} -CN; (d) β -CN; (e) κ -CN; (f) β -LG; and (g) α -LA. Results of the polynomial contrasts have been reported as the response curve (linear, quadratic, or cubic) of the variables across classes of milk proteins: solid lines represent data from the M-%milk model, and dotted lines represent data from the M-%cas model [proteins expressed as content in milk (M-%milk) or as percentage of total casein content (M-%cas)]. For each protein fraction, the class 4 represents the central class (-0.25 to $+0.25$ SD with respect to the mean). The amplitude of the other classes is 0.5 SD. Color version available online.

%CY. A high content of casein in milk can make the cheese-making process highly efficient because of its effect on milk constituent recovery in the curd. For each increment of 0.1% of casein in milk, we observed an increase of 0.27% in REC_{fat} and 0.20% in $REC_{protein}$. This strong effect meant that total casein had a robust linear effect on the efficiency of %CY, especially when expressed as weight of cheese ($Ef\text{-}\%CY_{curd}$), due also to the greater amount of water retained in the curd.

Effects of α_{S1} - and α_{S2} -CN

It is widely accepted that the concentrations and the proportions of each casein fraction play an essential role in the structure and rheological properties of the coagulum (Guinee, 2003), although less is known about the contribution of the content of each protein fraction to cheese yielding capacity and the efficiency of the cheese-making process. At the individual cow level, little attention has been paid to α_{S1} -CN, because of low variability at its locus, and to α_{S2} -CN, which exhibits almost monomorphic characteristics (Farrell et al., 2004).

An important function of α_{S1} -CN is its capacity to reduce the amounts of either β - or κ -CN, and in this regard many researchers have attributed to α_{S1} -CN the role of “molecular detergent” for the other casein fractions (Farrell et al., 2006). In our data, we found that α_{S1} -CN correlated negatively only with β -CN (-0.63) when expressed as relative content in total casein, whereas no relationship was found with κ -CN when expressed as relative content in total casein or as percentage in milk (data not shown).

Although α_{S1} -CN was present in lower amounts than β -CN in our study, we observed a very positive linear effect of the percentage of α_{S1} -CN in milk, which showed the greatest increment in $\%CY_{curd}$ (Figure 2a) compared with the other protein fractions ($M\text{-}\%milk$), which resulted from it having the highest yield of DM ($\%CY_{solids}$) and, in particular, high total milk protein recovery in the curd (Figure 3a). Our results are, in part, explained by results obtained by N. Amalfitano (University of Padova), C. Cipolat-Gotet (University of Parma), A. Cecchinato (University of Padova), M. Malacarne (University of Parma), A. Summer (University of Parma), and G. Bittante (University of Padova) (unpublished data): these authors reported greater curd firming and maximum curd firmness for milk samples with higher content of α_{S1} -CN in milk. We observed that an increase in the amount of α_{S1} -CN in milk had a positive effect on curd firming and syneresis, which are probably responsible for greater nutrient retention in the curd. The good efficiency of α_{S1} -CN and its phosphorylated form was also confirmed by the ratio

between the linear regression coefficient for $\%CY_{curd}$ (or $\%CY_{solids}$) and the standard deviation of those fractions with higher values than expected (data not shown).

When compared with other caseins, the results for α_{S2} -CN indicate this fraction as having a small influence on the efficiency of the cheese-making process. Its relative content in total casein negatively influenced milk protein recovery in the curd, probably as a result of its negative association with maximum curd firmness (Joudou et al., 2008; N. Amalfitano, C. Cipolat-Gotet, A. Cecchinato, M. Malacarne, A. Summer, and G. Bittante, unpublished data). Although the fluctuation in the α_{S2} -CN content in milk and in casein did not have a relevant influence on %CY, we observed a more marked negative effect on dCY (Figure 5c), although its relation with daily milk production was not relevant. With respect to $Ef\text{-}\%CY$, the effect of α_{S2} -CN was comparable with that of α_{S1} -CN and presented even worse efficiency, especially for DM in the curd, with its higher proportion of total casein in milk (Figure 4c).

Effect of β -CN

More studies have been carried out on β -CN and its relationship with coagulation and curd firming processes, including its use in technological treatments to enrich the natural composition of milk (St-Gelais and Haché, 2005), than on α -proteins. Several authors (Dunnewind et al., 1996; De Roos et al., 2000) have found a large reduction in the association between κ -CN and chymosin following addition of a small amount of β -CN to the milk solution. It seems that the role of chymosin in one or more binding sites located on para- κ -CN is shielded by β -CN, or it may even be that this protein fraction is a competitor of chymosin during enzymatic coagulation of these binding sites. Okigbo et al. (1985) failed to demonstrate any relationship between the proportions of β -CN and κ -CN in total casein content and showed that inferior coagulating characteristics were associated only with a higher content of γ - and other degraded caseins. However, these results were probably a consequence of the low number ($n = 9$) of individual cows sampled. Several authors observed a reduction in curd firming time and an increase in maximum curd firmness when analyzing milk samples with a higher proportion of β -CN in total casein (Joudou et al., 2008; N. Amalfitano, C. Cipolat-Gotet, A. Cecchinato, M. Malacarne, A. Summer, and G. Bittante, unpublished data).

With respect to cheese yield, Marziali and Ng-Kwai-Hang (1986) found that β -CN content in milk had a positive relationship with actual cheese yield, although it was less important than that of α -CN. Moreover, when individual milk samples from 110 cows of Swedish

Red and Swedish Holstein breeds were processed, β -CN content exhibited a strong correlation with the amount of casein retained in the curd (Hallén et al., 2010). In our study, $\%CY_{\text{curd}}$ increased in relation to greater REC_{protein} only for α_{S1} -CN and β -CN, whereas in the case of κ -CN the increase in $\%CY$ was related more to the greater recovery of milk fat in the curd (Figure 3e). The scenario observed for β -CN resulted in higher efficiency values for $\%CY_{\text{curd}}$, although this protein induced a loss in terms of $Ef-\%CY_{\text{solids}}$ when expressed as a proportion of total casein (Figure 4d). These results confirm what has been found in the processing of goats' milk (Damian et al., 2008), where a decrease in $\%CY$ was related to an increase in β -CN as a proportion of total casein content in milk.

Effect of κ -CN

In enzymatic coagulation, κ -CN is hydrolyzed to produce para- κ -CN (κ -CN fragment: 1–105), which is linked to the micelle core, and macropeptides (κ -CN fragment: 106 \pm 169), which constitute about 30% κ -CN and represent 4 to 5% of milk total casein. During the coagulation process, when about 85% of the total κ -CN is hydrolyzed, micelles lose their colloidal stability and begin to aggregate progressively into a gel network (Guinee, 2003).

Given the high importance of κ -CN function in the coagulation process, the genetics of its variants have also been widely investigated. The superiority of the B variant with regard to coagulation time and the curd firming process is well known (Bittante et al., 2012). However, Bonfatti et al. (2011a) discovered that this superiority is due to the high κ -CN content in milk (and relative proportion in casein) when this variant is present and found that when the statistical model included the effect of the content of this protein fraction, the coagulation properties were not affected by its genetic variants. Moreover, Bonfatti et al. (2011a) found that $\%CY_{\text{curd}}$ in field conditions is not affected by genetic variants when milk samples with similar total CN composition are processed. Verdier-Metz et al. (2001) reported greater $\%CY$ in milks presenting high frequencies of the κ -CN B variant with higher casein:protein ratios in the production of Saint Nectaire. However, milk with a high κ -CN content has small casein micelles, which form rennet curd with improved rheological properties and a high capacity to entrap milk constituents (especially fat droplets) into the paracasein matrix (Niki et al., 1994; Walsh et al., 1998). Using a laboratory cheese-making procedure to process individual milk samples, Macheboeuf et al. (1993) also observed a positive effect of κ -CN content in milk on $\%CY_{\text{water}}$, although water retention in the curd could

have been influenced by the use of a centrifuge for whey expulsion (Cipolat-Gotet et al., 2016). Our study confirms the positive linear effect of κ -CN on all $\%CY$ traits when expressed as percentage of milk, albeit less important than the effects of α_{S1} -CN and β -CN, and subsequently a weaker effect on this protein's cheese yield efficiency, in particular for $Ef-\%CY_{\text{curd}}$ (Figure 4e; M-%milk and M-%cas). This was probably due to the higher losses of whey proteins resulting from the negative effect of κ -CN, REC_{protein} , and syneresis (N. Amalfitano, C. Cipolat-Gotet, A. Cecchinato, M. Malacarne, A. Summer, and G. Bittante, unpublished data), especially when there is a greater proportion of κ -CN in the casein. Moreover, in enzymatic coagulation, the higher content of κ -CN in milk can increase the amount of glycomacropeptide (fragments 106–169) lost in the whey as a consequence of κ -CN hydrolysis by chymosin. The positive effect of κ -CN on the recovery of milk components (Donnelly et al., 1984; Dalgleish et al., 1989) in the casein matrix was mostly related to fat whether this protein was expressed as a percentage of milk or of total casein content (Figure 3e). These results agree with Malacarne et al.'s (2006) study of milk samples from Italian Brown cows in the production of Parmigiano Reggiano.

Effect of Whey Proteins

In the present study, we analyzed the 2 main whey proteins in bovine milk, β -LG and α -LA, both of mammary derivation, that contribute approximately 50% and 20% to total whey proteins, respectively. Although they are not directly involved in the enzymatic coagulation process, whey proteins are dissolved in the cheese moisture (about 4% in total) and contribute very little to $\%CY_{\text{curd}}$. Through the use of milk treatments, such as concentration by ultrafiltration, whey proteins can be incorporated into the curd, making the cheese-producing process more efficient in terms of REC_{protein} and $\%CY_{\text{curd}}$. Moreover, as these proteins precipitate under intense heating or salting, an increase in REC_{protein} can also be observed when whey proteins are subjected to heat treatment, such as in the case of ricotta (Guinee, 2003).

The finding that whey protein had little influence in the gelation phase (N. Amalfitano, C. Cipolat-Gotet, A. Cecchinato, M. Malacarne, A. Summer, and G. Bittante, unpublished data) compared with caseins, for which we found a relation between coagulation and efficiency of the cheese-making processes, was not in line with our results for $\%CY$, REC , and $Ef-\%CY$ traits, especially in the case of α -LA. Whey proteins are not involved directly in the renneting process, but they greatly influence recovery in the curd, as a part of

these fractions ends up in the curd (as part of the whey retained in the curd) and the whey. We observed that β -LG had a highly negative influence on the cheese-making process, especially with regard to fat and protein recoveries (Figure 3f), where it exhibited a worse pattern than the other protein fractions. As a consequence of the strong relationship between recoveries in the curd and cheese-making efficiency, this fraction was the least efficient protein in terms of %CY (Figure 4f). In contrast, α -LA had a fairly positive effect on the recovery of fat in the curd when it increased as percentage of casein. The increase in α -LA compared with the other fractions may be considered positive in terms of Ef-%CY_{curd} (Figure 4g), and it had a positive effect on daily cheese production. In contrast, β -LG, together with α _{S2}-CN, were the only fractions that negatively influenced daily cheese production at the individual level cow. Dadousis et al. (2018) reported a positive genetic correlation between α -LA and dMY, and the highest value of dMY variance explained by this fraction compared with the other proteins. Through the use of 2-dimensional electrophoresis, Hsieh and Pan (2012) observed that a fraction of β -LG was found in the curd together with α _{S1}-, α _{S2}-, β -, and κ -CN after incubation of milk with chymosin. In contrast to our results, Hallén et al. (2010) observed that milk nutrient losses in the curd after a simulated pressing were positively associated with α -LA content in milk, but, as mentioned before, the use of a centrifuge to separate the curd from the whey could influence the results in terms of milk nutrient recoveries and Ef-%CY traits.

CONCLUSIONS

We report a highly significant effect of protein fractions on the efficiency of the cheese-making process. This study permitted us to assess the effect of each single protein fraction quantity in milk and its proportion to total casein on the cheese-making process. The quantities and interactions of protein fractions appear to be important in milk-to-cheese processes, where the effects of the amounts of proteins in the milk were more significant. Among the different proteins in the milk, Ef-%CY_{curd} was positively associated mostly with β -CN, due in particular to greater water retention, whereas the amounts of α _{S1}-CN and κ -CN influenced the efficiency of DM yield as a consequence of their relationship with the recovery rates of protein and fat in the curd. In contrast, β -LG was highly negatively related to all the traits associated with the cheese-making process. In terms of daily cheese yield, whey proteins were also of high importance as they exhibited positive relationships with α -LA and negative relationships with β -LG. Although the characteristics of coagulation and

the curd firming process can influence the relationships between milk protein (and its fractions) and cheese-making efficiency, it was very important to carry out a study using a laboratory cheese-making procedure to process individual milk samples so that information could be collected for use in detailed monitoring of milk quality at the herd level, breeding selection programs (also linking details of the proteins' genetic variants), and designing appropriate milk payment systems. Given that HPLC analyses to assess protein fractions are expensive, complex, and time consuming, it would be useful from a practical point of view to find faster, easier-to-use, and less costly alternative methods (e.g., infrared spectroscopy). Assessment of the effectiveness of infrared technology and the predictive ability of calibration equations for milk protein fractions as well as the study of the genetic and genomic variations in Fourier-transform infrared-based predictions at the population level remain matters for further studies.

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