

University of Parma Research Repository

Country of origin label monitoring of musky and common octopuses (Eledone spp. and Octopus vulgaris) by means of a portable near-infrared spectroscopic device

This is the peer reviewd version of the followng article:

Original

Country of origin label monitoring of musky and common octopuses (Eledone spp. and Octopus vulgaris) by means of a portable near-infrared spectroscopic device / Varra, M. O.; Ghidini, S.; Fabrile, M. P.; Ianieri, A.; Zanardi, E.. - In: FOOD CONTROL. - ISSN 0956-7135. - 138:(2022), p. 109052.109052. [10.1016/j.foodcont.2022.109052]

Availability: This version is available at: 11381/2923189 since: 2024-12-10T08:51:46Z

*Publisher:* Elsevier Ltd

Published DOI:10.1016/j.foodcont.2022.109052

Terms of use:

Anyone can freely access the full text of works made available as "Open Access". Works made available

Publisher copyright

note finali coverpage

1	Country of origin label monitoring of musky and common octopuses	
2	(Eledone spp. and Octopus vulgaris) by means of a portable near-	
3	infrared spectroscopic device	
4		
5	Maria Olga Varrà, Sergio Ghidini, Maria Pia Fabrile, Adriana Ianieri, Emanuela	
6	Zanardi*	
7		
8	Department of Food and Drug, University of Parma, Vi <u>Strad</u> a del Taglio, 10, 43126	
9	Parma, Italy	
10		
11		
12		
13		
14	*CORRESPONDING AUTHOR:	
15	E-mail address: emanuela.zanardi@unipr.it (E. Zanardi)	Codice campo modificato
16		
17		
18		
19		
20		
20 21		
20 21 22		
20 21 22 23		
20 21 22 23	1	

## 24 ABSTRACT

The recognized economic and nutritional value of cephalopods has recently led to a widespread 25 capture fishery and distribution worldwide, thus increasing the possibility of fraudulent substitution 26 27 of the products and posing into question the truthfulness of the geographic indications reported on 28 the label.-Modern analytical techniques using miniaturized and portable near infrared (NIR) spectroscopy instruments are particularly suited for assessing the authenticity of fishery products 29 30 since meeting the requirements of rapidity, eco-friendliness, cost-effectiveness, and easiness of application. The objective of the present study was to verify the suitability of use of a portable and 31 ultra-compact NIR spectrometer combined with machine learning to characterize the geographic 32 origin of two widely consumed octopus species. Replicate NIR spectra in the (908.1-1676.2 nm). NIR 33 34 region of 118 musky and 29 common octopus specimens (Eledone spp. and Octopus vulgaris) from Portuguese Atlantic or Spanish Mediterranean fishing areas were recorded, pre-processed and 35 elaborated via the following classification algorithms: orthogonal partial least square discriminant 36 analysis (OPLS-DA), logistic regression (LR), random forest (RF), support vector machine (SVM), 37 38 and multilayer perceptron-artificial neural network (MLP-ANN). When 7-fold cross validation was performed on 75% of data, the results showed that linear tools (OPLS-DA and LR) were the most 39 powerful and stable techniques in recognizing the origin of both octopus species, with (mean 40 41 sensitivity, specificity, accuracy, and precision values above 98%)-and the lowest associated standard deviations. During the external validation phase (using 25% of the remaining spectral data) OPLS-42 DA, SVM, and MLP-ANN performed better for common octopuses-(with no classification errors), 43 while LR and MLP-ANN for musky octopuses (with only 2 and 3 Mediterranean samples 44 45 misclassified, respectively).\_\_The achieved outcomes suggest the combination of portable NIR spectroscopy and machine learning as a promising plan of action to be adopted for the creation of 46 an integrated analytical platform with capabilities for automated data recording, processing, and 47 reporting, which may be helpful for on-site and in-line monitoring of fishery products. 48

# 49 Abbreviations

50	first derivative, 1stDer; second derivative, 2ndDer; area under the receiver operating characteristic
51	curves, AUROC; logistic regression, LR; multilayer perceptron artificial neural network, MLP-ANN;
52	multiplicative scatter correction, MSC; near infrared, NIR; orthogonal partial least square-
53	discriminant analysis, OPLS-DA; principal component analysis, PCA; principal component, PC;
54	radial basis function, RBF; random forest, RF; root mean square error from cross-validation,
55	RMSECV; standard normal variate, SNV; support vector machine, SVM;

56

57 Keywords: rapid methods; machine learning; chemometrics; food authenticity; geographical origin;
58 cephalopods.

#### 59 1. Introduction

Cephalopod mollusk species belonging to the Octopodidae and Eledonidae families (collectively 60 known as octopuses) represent an important fishery resource, with high economic, social, cultural, 61 and nutritional values, especially for Asiatic and Mediterranean countries. Common octopus 62 (Octopus vulgaris) and horned and musky octopus (Eledone spp.) are the main octopus species 63 produced and exploited for human consumption by Mediterranean and Central Eastern Atlantic 64 countries. According to the most recent data, Italy, Spain, Portugal, Greece, and France together 65 account for 7% and 95% of world and European production, respectively (European Market 66 67 Observatory for Fisheries and Aquaculture Products (EUMOFA), 2020), being characterized by large- as well as small-scale artisanal octopus fisheries which are of extreme importance for local 68 69 economy (Pita et al., 2021). Notwithstanding this, octopus catches by European Union countries have steadily declined over the past decade as a combined result of the effects of overfishing practices, 70 new fisheries management policies focusing on sustainable practices, and climate change (Tinacci et 71 al., 2020). On the other hand, to compensate the increased demand, large volumes of frozen octopus 72 73 are now imported from third countries (Morocco and Mauritania mainly) (EUMOFA, 2020) and retailed by the local markets predominantly as thawed products. This market configuration increases 74 the chances of species and fisheries suffering unfair competition, as well as illegal, unreported, and 75 76 unregulated fishing activities being pursued, thus bringing about commercial frauds regarding the falsification of the geographic origin and traceability problems which, in turn, have important 77 economic and sustainability repercussions (Fox et al., 2018). Indeed, fraudulent mislabeling of 78 cephalopods and cephalopod-based products, occurring at any level of the supply chain, was reported 79 more than two and three times frequently compared to that of crustaceans and fish (Guardone et al., 80 2017). Despite being economically motivated, mislabelling concerning the geographic origin of 81 cephalopods may represent a safety risk for consumers due to the potential exposure to different 82 contaminants and pollutants. Indeed, cephalopods originating from areas at risk of harmful algal 83 bloom have been reported to accumulate several marine biotoxins, such as tetrodotoxin, saxitoxins, 84

85 palitoxins, and domoic acid, thus possibly acting as toxin vectors to humans (Lopes et al., 2013; Whitelaw et al., 2019; Karlson et al., 2021). Public health consequences may also arise from the 86 consumption of cephalopods from specific polluted fishing areas due to the presence of very high 87 88 concentrations of heavy metals and persistent organic pollutants (Gomes et. al., 2013; Rjeibi et al., 2014; Roldán-Wong et al., 2018). Similarly, an actual food safety concern is related to the human 89 exposure to microplastics through the consumption of seafood whose contamination can vary a lot 90 among countries over the world (EFSA CONTAM Panel, 2016). Microplastics were in fact reported 91 to be a vector for chemical contaminants (heavy metals, organochlorine pesticides, drug residues, 92 polycyclic aromatic hydrocarbons, polychlorinated biphenyls, and polybrominated diphenyl ethers) 93 (Brennecke et al., 2016; Camacho et al., 2019; De-la-Torre, 2020). 94

The prevention of food fraud should be based on a management system approach relying on two main 95 96 supports: a proper vulnerability assessment system and the design and implementation of mitigation measures (Fox et al. 2018). That one is the framework within which the development of appropriate 97 98 analytical methods, testing compliance of foods with their label descriptions, is set as a fundamental 99 part of the management of food fraud incidents (Stadler et al., 2016). Recent attempts in literature 100 and practice have suggested a few analytical laboratory methods as appropriate means to identify different types of frauds affecting cephalopod products. The issue of species mislabeling was 101 addressed by using DNA barcoding (Guardone et al., 2017; Tatulli et al., 2020), while the replacement 102 103 of fresh with frozen/thawed products was successfully identified through proteomics (Guglielmetti et 104 al. 2018) or histological evaluation of tissues (Tinacci et al., 2020). Likewise, illicit water addition 105 was uncovered by measuring electric conductivity and dielectric properties of samples (Mendes et 106 al., 2018), as well as through the development of ad hoc fast 3D scanning methods (Han et al., 2020). Facing with the task of identifying the geographical origin of cephalopods is more challenging 107 108 because of many pre-catch (e.g., seasonality, sizes) and post-catch (e.g., storage conditions) factors 109 overlapping with the issue of interest and the lack of target measurable parameters providing the 110 certainty of geographical authenticity (Esslinger et al., 2014; Varrà et al., 2021a). Nevertheless,

considering the territorial nature, the small activity area, and the very small capability of metabolization (Oliveira et al. 2018; Arechavala-Lopez et al., 2019) cephalopod mollusks and, in particular, octopuses, appear to be good indicators of the seawater areas they inhabit. Based on this consideration, the use of comprehensive approaches relying on the combination of modern instrumental and advanced statistical methods, such as those based on near infrared (NIR) spectroscopy and machine learning, may represent a direct solution for the identification of the sources of origin of cephalopod stocks.

118 With the advances in instrumentatal technology, the latest years are witnessing a shift from the laboratory usage of stationary benchtop NIR equipments to miniature and portable devices for quality, 119 safety, and authenticity testing of food of animal origin, including fish and seafood (Grassi et al., 120 2018; Cruz-Tirado et al., 2021; Dos Santos et al., 2020; Silva et al., 2020; Dos Santos Pereira et al., 121 122 2021; González-Mohino et al., 2020; Müller-Maatsch et al., 2021a; Pennisi et al., 2021; Yakes et al., 123 2021; Yu et al., 2020; Currò et al., 2022). Nevertheless, only one study proved the suitability of using 124 portable NIR technology coupled with machine learning to monitor traceability of cuttlefish 125 cephalopods (Currò et al., 2021). Portable instruments, besides being rugged, user-friendly, compact, 126 ultra-light, and cheaper compared to traditional stationary instruments, allow direct analysis without sample processing and consumption, thus facilitating on-site or in-line analysis and globally 127 minimizing times and costs associated with the analytical flow (Beć et al. 2021; McVey et al. 2021; 128 129 Müller-Maatsch et al., 2021b). Moreover, methods based on the use of portable NIR devices meet the 130 requirements and goals of 'White Analytical Chemistry', showing a sinergy between analytical, 131 ecological, and practical attributes (Nowak et al., 2021). Indeed, the greenness of the approach 132 (absence of waste generation and toxic solvents, low power consumption) is perfectly balanced both with analytical efficiency (accuracy, precision, sensitivity) and practical/economic efficiency (Nowak 133 134 et al., 2021). Although these benefits and, the extensive research done, and the substantial progress in the development of more efficient computer algorithms, no reliable methods haves been yet 135 136 deployed in routine monitoring or accepted as an official standard since there is still the need to

137 identify the proper machine learning algorithms able to speed up and simplify the analytical workflow and accurately ascertain the sought food authenticity features (Ríos-Reina et al., 2021). 138 139 Based on the above considerations, the goal of the present study is to propose a rapid, cheap, and eco-140 friendly analytical methodology to identify possible fraudulent mislabeling concerning the country of origin of different octopus species, which can be potentially useful for regulators, industry, and 141 stakeholders, for the inspection and certification of cephalopods authenticity. To this end, a handheld, 142 portable, and wireless NIR spectrometer was used to analyze two species of octopus originating from 143 144 two different fishing areas (Spanish Mediterranean and Portuguese Atlantic), and the resulting 145 spectral fingerprints were patterned by different traditional and modern machine learning tools in order to identify a fit-for-purpose methodology for their origin recognition. 146

## 147 2. Materials and Methods

## 148 2.1. Sample collection and handling

149 Three different batches of musky octopuses (Eledone spp., Cephalopoda: Octopodidae) of medium 150 size (200-300 g total body weight, bw) and fished by means of otter trawls were collected during the 151 autumn season from each of the two sampling sites chosen, corresponding to the FAO fishing areas 152 37.1.1 (i.e., Balearic waters of the Western Mediterranean Sea) and 27.9.a (i.e., Eastern Portuguese 153 waters of the North-East Atlantic Ocean). Musky octopus from Mediterranean Sea and Atlantic 154 Ocean accounted for 61 and 57 specimens, respectively. Similarly, three different batches of common octopuses (Octopus vulgaris, Cephalopoda: Octopodidae) of medium size (1300-1500 g bw) and 155 caught by means of otter trawls in summer were retrieved from the FAO fishing area 37.1.1, 156 157 accounting for a total of 29 specimens. Other 10 common octopus samples of the same size (1000-1500 g bw) and fishing season were collected from the FAO fishing areas 27.9.a. The sampling plan 158 is graphically summarized in Figure 1. 159

All the samples were transported and delivered as fresh products and stored at freezing temperature (-21  $\pm$  2°C) once arrived at the laboratory. Before analysis, the samples were defrosted in a refrigerator ( $4 \pm 2$  °C per at least 18 hours) and the temperature gradually brought to room-conditions ( $20 \pm 2$  °C) for 60–90 min. The octopuses were then manually skinned and dissected. Since the mantle was the flesh portion selected for the experiments, it was separated from the arms and left whole for subsequent spectral analysis.

### 166 2.2. MicroNIR setup and measurement

The octopus mantles were analyzed by using the ultracompact, portable and wireless NIR device
MicroNIR OnSite-W (Viavi Solutions, Santa Rosa, CA) equipped with the spectral acquisition
software MicroNIR Pro<sup>TM</sup> (v.3.1, Viavi Solutions, Santa Rosa, CA, USA).

Diffuse reflectance NIR spectra were recorded in the 908.1-1676.2 nm region as 100 co-added scans 170 and with an integration time of 10 ms. The spectral apparent resolution was 6.25 nm, hence each 171 spectrum consisted of 125 reflectance points. NIR spectra were recorded by perpendicularly 172 173 interfacing the acquisition window of NIR spectrometer with the surface of the sample. Spectral 174 scanning was performed on four different points of the ventral and dorsal side of the mantle in order 175 to collect heterogenicity of composition and thickness. The four replicate spectra were all individually 176 used for statistical processing, thus resulting in two data matrices consisting of 472 spectra of musky 177 octopus (118 samples  $\times$  4 spectral replicates) and 156 spectra of common octopus (39 specimens  $\times$  4 178 spectral replicates). Before NIR analysis and every 15 minutes during the analysis execution, the MicroNIR device was calibrated by recording a total absorbance (dark) reference spectrum (by 179 180 leaving the lamps on and the acquisition window of the spectrometer empty) and a total reflectance reference spectrum (by using the external white diffuse reflectance standard disc Spectralon® 99%, 181 182 LabSphere, North Sutton, NH, USA) to correct the background signal for the proper response over operation time and any little temperature changes. 183

184 *2.3. Statistics and data modeling pipeline* 

The spectral data were exported, transformed to apparent absorbance values, mean-centered, and preprocessed by using standard normal variate (SNV), multiplicative scatter correction (MSC), first derivative (1<sup>st</sup> Der), and second derivative (2<sup>nd</sup> Der), alone or as combined spectral filters, to identify
the best solution enhancing the signal-to-noise ratio and spectral resolution.

189 To achieve significant and robust results, each classification model was trained on 75% and validated 190 on 25% of the common or musky octopus spectral pre-processed dataset. The low number of samples in the common octopus dataset hindered the possibility of averaging the four spectral replicates 191 recorded for each sample. Therefore, it was decided to retain the four individual spectral replicates of 192 193 each sample and to perform the 75:25 splits by randomly allocating them together into either the 194 internal calibration sets (i.e., training sets) or the external validation sets (i.e., validation sets), as follows: i) 354 spectral data into the training set of musky octopuses (118 samples  $\times$  3 spectral 195 replicates, i.e. 75% of total spectral replicates); ii) 118 spectral data into the validation set of musky 196 octopuses (118 samples × 1 spectral replicate, i.e. 25% of total spectral replicates); iii) 156 spectral 197 198 data into the training set of common octopuses (39 samples  $\times$  3 spectral replicates, i.e. 75% of total 199 spectral replicates); iv) 39 spectral data into the training set of common octopuses (39 samples  $\times 1$ 200 spectral replicate, i.e. 25% of total spectral replicates. Thereby, a balanced repartition of the two 201 representative classes to be discriminated (i.e., the two geographical provenances of the samples), as 202 well as the maximum independence among samples in the two sets were assured. In order to enhance statistical confidence with small sample sizes, training data were also 7-fold cross-validated (internal 203 calibration phase), thus the models were trained on 6/7th and internally evaluated on 1/7th of the data. 204 205 Considering that for small-sized datasets the whole performances of the machine learning models are 206 influenced by the exact repartition of samples into the training and the test-validation sets, the 75:25 splits were repeated 4 times for all the models. Hence, four different training and testing validation 207 208 sets were generated for each octopus species. The final statistical outcomes resulting from training 209 and validation stages were reported as means and standard deviations (Michelucci & Venturini, 210 2021).

After data repartition, the spectra of the training sets were used to create two principal component analysis (PCA) models (one for each of the octopus species considered). PCA was used for the sake 213 of screening data structure, to reveal any potential hidden correlations among samples and variables, 214 and to detect potential outliers, otherwise detrimental for the accuracy and stability of the subsequent 215 machine learning classification models. Nevertheless, since used as a preliminary investigation tool, 216 PCA was computed only once, i.e., by using data included in only one of the four training sets created. The following supervised classification tools were then tested: orthogonal partial least square 217 discriminant analysis (OPLS-DA), logistic regression (LR), random forest (RF), support vector 218 machine (SVM), and multilayer perceptron artificial neural network (MLP-ANN). Calculations of 219 220 PCA and OPLS-DA were performed by using the statistical software packages SIMCA (v. 16.0.2, Sartorius Stedim Data Analytics AB, Umea, Sweden), while calculations of LR, RF, SVM, and MLP-221 ANN were done by using IBM SPSS Modeler software (v. 18.2, SPSS Inc., Chicago, IL, USA). 222

## 223 2.3.1 Training parameters of the machine learning models

In this work, the number of predictive and orthogonal new latent variables of the calibration OPLS-DA models was estimated by cross-validation. The fitting and prediction abilities of the models were evaluated by analyzing the following parameters:  $R^2X$  (cumulative variability of the spectral data modelled by all the extracted latent variables),  $R^2Y$  (cumulative variation associated to class labels explained by all the extracted latent variables),  $Q^2X$  (cumulative variability associated to class labels predicted by all the extracted latent variables), RMSECV (root-mean-squared error of cross validation) and RMSEP (root-mean-squared error of prediction).

- The overall significance of the logistic regression equation to classify octopus samples was indeed estimated by the likelihood ratio Chi-square test (using -2 times the log of the likelihood as reference value) and taking into consideration the Cox-Snell pseudo-R<sup>2</sup> regression value.
- As for RF models, their structures were created using the Gini Impurity Index as a tree branching and variable selection criterion. According to the default settings suggested by the software, the number of trees to be generated was beforehand set to a maximum of 100. To avoid over splitting, the maximum depth of the tree structure was set to 10 levels while the minimum number of samples to

238	be included into each child node was set to 5. Finally, the number of split variables for each tree node
239	was set at 11 (square root of the total number of variables).
240	For the training of the SVM models, the Radial Basis Function (RBF) was used as kernel function.
241	The balance between the model complexity and training error was established by setting the
242	regularization parameter C (box-constraint or penalty factor) to 90, the additional kernel function $\gamma$
243	parameter to 0.1, and the regression precision parameter $\varepsilon$ to 0.1, by following the default settings
244	suggested by the software.

Finally, the architecture of the MLP-ANN was built automatically and included an input layer (containing the NIR spectral data), one single hidden layer with one hidden neuron (transforming the weighted sum of the inputs by a hyperbolic tangent activation function to generate the outputs), and an output layer (using SoftMax activation function to estimate the probability of samples belonging to each classification group).

The relative contribution of each NIR wavelength to the predictive models was measured through
 different functions, based on the machine learning algorithm employed: while model-dependent
 methods consisting on the evaluation of the of Variable influence on projection (VIP) index, t-test

253 statistics, and mean squared error were applied for OPLS-DA, LR, and RF, respectively, model-

254 independent method based on the calculation of the area under the receiver operating characteristic

- 255 (ROC) curve values (AUROC) were applied both for SVM and MPL-ANN.
- 256 2.3.2. Evaluation and comparison of the classification models performances

The estimation of the goodness of each classification model was performed on multiple fronts. Considering that each classification model is characterized by its own statistic outputs, standardized metrics providing a direct comparison of the performances of the models were chosen.

Firstly, the mean accuracy, specificity, sensitivity, and precision parameters (Fawcett, 2006) were calculated from the confusion matrices reporting percentages of common and musky octopus samples of the training sets correctly classified in the proper class during the cross-validation process. Prediction capabilities of the models were then graphically inspected through the area under the receiver operating characteristic (ROC) curve values (AUROC)<u>AUROC values</u> for each of the two classes (Mediterranean, Atlantic) of the validation sets, which is an optimal compromise to summarize sensitivity and specificity. ROC curves <u>for validation data</u> were created by plotting the true positive rate (TPR or sensitivity) versus the false positive rate (FPR or 1-specificity) at all predicted probability cut-off values (Fawcett, 2006).

## 269 3. Results and Discussion

#### 270 *3.1. NIR spectral characteristics and correction*

Pre-processing of NIR spectra is quite a mandatory step in common practice to minimize the systematic variation in the spectra deriving from light scattering. Non-linearity and multiplicative effects deriving from this variation appear in the form of baseline shifts and drifts which are not directly related to the chemical properties, but rather to the structural features and physical status of the sample (Rinnan et al., 2009).

As it can be observed form Figure 2, light scattering effects were found in the raw absorbance spectra 276 277 of both musky and common octopus samples recorded by MicroNIR. Therefore, --different pre-278 processing techniques were applied to the raw spectra by finding a compromise among drifts/shifts 279 minimization, an acceptable peak separation degree, and the addition of unwanted noise. As a result, MSC and 2<sup>nd</sup> Der were discarded, while transformation by SNV followed by 1<sup>st</sup> Der (Norris-Williams, 280 281 quadratic polynomial order, 15 points gap) was selected the best suited combination of spectral filters 282 since allowed to re-align NIR spectra and partially suppress broad bands, without over processing and potential information loss (Figure 2). Despite intrinsic differences related to species, the average 283 284 SNV plus 1<sup>st</sup> Der spectra of musky and common octopuses from the two geographical provenances were not characterized by rough visual differences in the absorbance pattern. The predominant bands 285 were found in the 950-1000 nm, 1100-1200 nm, and in the 1300-1450 nm NIR regions (Figure 2). 286 287 Nevertheless, considering that the original maximum peaks in the raw spectra correspond to the zerocrossing segment of the 1<sup>st</sup> Der spectra, the predominant individual features observables within the 288

289 above mentioned NIR regions for common octopus dataset were at 1194 and 1440 nm, where -CH<sub>2</sub> 290 and -CH<sub>3</sub> bonds of aliphatic hydrocarbons were reported to absorb (Workman & Weyer, 2012). In 291 the case of musky octopus spectral dataset, the first feature moved to a lower wavelength (1186 nm), 292 while the second one was located at the same wavelength (1440 nm) (Figure 2). Due to difficulties in 293 sampling procedures and in the interpretation of the NIR spectra, only a few research works have focused on the correlation between NIR absorbance spectra and the geographical origin of fish and 294 295 seafood. Mostly, NIR wavelengths related to lipid absorption and, sometimes, proteins, were already 296 identified as useful for the classification of fish by origin (Ghidini et al., 2019; Currò et al., 2021; Varrà et al., 2021b). Nevertheless, the assumption behind the possibility of fingerprinting the origin 297 of cephalopods by using lipid and protein spectral features rely on the well-known link existing 298 299 between the characteristics of the specific marine environment (water saline composition and average 300 temperature, sediments, currents, seasonal temperature changes, population, and availability of fish preys) and the parallel variation of the chemical constituents of the fish tissues (Saito et al., 1997 In 801 302 this context, octopus species, since particularly sensitive to any variation of the aquatic ecosystem, 303 can provide useful information about the seawaters of origin, including the environment pollution 804 status (Sillero-Ríos et al., 2018), Indeed, especially -due to previous studies demonstrated that both 305 variations of the fatty acid and elemental profiles vary a lot among O. valgaris populations sampled in different areas and, therefore, they can be successfully used as markers of geographical origin 306 807 (Arechavala-Lopez et al., 2019; Semedo et al., 2012). On the other hand, also pollutants or toxic 808 elements on the fishing area may be similarly reflected into octopus tissues, leading to questioning food safety. This is one of the reasons why the authentication of seafood according to the geographic 809 310 origin represent an important prerequisite of food safety (Freitas et al., 2020).

The possible differences in composition pointed out in the NIR spectra might explain the results achieved upon applying PCA. Specifically, the PCA models for musky and common octopuses were characterized by 8 and 7 principal components (PCs), covering 99.3 and 99% of the total variance, respectively. From the score scatter plots of the first three PCs (Figure 3), it can be sated that none of the octopus samples was suspected of being an outlier, since not crossing the 95% confidence limits for Hotelling's  $T^2$  defined by plot ellipse. At the same time, the PC1 collected the inter-origin variability only among common octopuses, but not among musky octopuses, which did not separate efficiently each other.

Nevertheless, all the above aspects suggested a promising route for the application of supervised classification methods which could efficiently address the challenge of identifying the origin of octopuses by using NIR spectroscopy.

## 322 3.2. Analysis and comparison of machine learning models performances

323 In the field of food quality, safety and authenticity, there is a definite trend towards the automation and the use of smart fingerprint technologies, able to collect a huge amount of data to characterize 324 325 foods and food systems in a comprehensive way, such as those based on miniaturized and portable spectroscopic sensors (Mevey et al., 2021). This trend has been accompanied by a substantial progress 326 327 in the development of more efficient computer algorithms and solutions, aimed to speed up and 328 simplify the analytical workflow without sacrificing the reliability of the results. At the same time, 329 coupling fingerprinting techniques with advanced computer assisted data analysis offers the advantages of extending the domain of food applications thanks to the possibility of monitoring 830 quality, safety, authenticity though one single analysis and, thus, preventing food fraud and food-331 332 borne illness.

In this work, five different powerful machine learning tools were tested against NIR spectroscopic data of musky and common octopus specimens of Mediterranean or Atlantic fishing origin<u>included</u> <u>into the training sets</u>, with the aim to develop a new tool for the prevention of potential frauds related to the falsification of the origin. The information embedded into the <u>SNV + 1<sup>st</sup> Der</u> pre-processed spectra was thus modelled by selecting OPLS-DA, LR, SVM, RF, and ANNs as supervised classification tools, which were initially tested on the training data by applying a cross-validation process (see *Section 2.3*). The cross-validation results of the five tested machine learning tools are illustrated in Figure 4, while a summary of the modelling statistics is reported in *Supplementary Materials* (Tables S1, S2, S3, Figure S1). Each model was trained by considering all the spectral variables included into the dataset, corresponding to 125 NIR absorbance values (908.1–1676.2 nm spectra). The only exception was represented by RF models which automatically perform a feature selection to avoid overfitting. These models were built by using a total of 53 and 60 input variables for musky and common octopus classifications, respectively.

347 Contrary to what achieved when applying PCA (see Section 3.1) slightly better results were obtained 348 when modelling musky octopuses compared to common octopuses, thus potentially confirming that the higher number of the analyzed samples still included in their NIR spectra a fraction of discriminant 349 information related to the origin which was captured and described by the more powerful supervised 350 351 algorithms. The performance metrics in cross validation were all above 96% except for RF models, 352 which were characterized by the highest error rates (with approx. 6 and 16% of musky and common 353 octopus wrongly recognized). The conventional linear algorithms were the most performant ones. 354 Specifically, LR was found to be the best solution to recognize the origin labelling of common 355 octopuses (average values of accuracy, specificity, sensitivity, and precision over 4 repetitions of  $99.79 \pm 0.43$ ,  $99.86 \pm 0.29$ ,  $99.86 \pm 0.29$ , and  $99.60 \pm 0.81\%$ , respectively), while OPLS-DA showed 356 the highest accuracy, specificity, sensitivity, and precision metrics for musky octopuses (average 357 358 values over 4 repetitions of  $99.58 \pm 0.67$ ,  $99.59 \pm 0.66$ ,  $99.59 \pm 0.66$ , and  $99.57 \pm 0.68\%$ , respectively) (Figure 4). In particular, the highest sensitivity (related to the true positive rates) and specificity 859 360 (indicating the true negative rate) values shown by OPLS-DA and LR have both important positive 361 consequences on the overall goodness of the discriminant methodology. In fact, sensitivity values indicate the degree of confidence in identifying the real authentic samples, thus having a direct impact 362 on the economic side. On the other hand, specificity values indicate the degree of confidence in 363 364 identifying the real non-authentic samples, with significant repercussions on the legal front.

365 As for computational performances of both MLP-ANN and SVM, also these tools showed very good 366 predictions in cross-validation. SVM is one of the most frequently used machine leaning technique 367 in food chemistry and authentication studies and, compared to linear classification methods, offers 368 the advantages of adaptability towards the non-linear distribution of the data typical of NIR spectroscopy (Jiménez-Carvelo et al., 2019). The superiority of SVM methods over traditional linear 369 370 classifiers combined with NIR spectral data in verifying different food authenticity claims has been demonstrated in several works (Cardoso & Poppi, 2021; Benes et al., 2020; Parastar et al., 2020; 371 372 Sampaio et al., 2020; Bisutti et al., 2019). The suitability of using SVM to authenticate cephalopods (Sepia officinalis) has been also confirmed in a recent work, where its application yielded 83-100% 373 balanced accuracy, 67-100% sensitivity, and 88-100% specificity for the classification of the 374 samples according to 5 different geographical origins (Currò et al., 2021). It should be noted, 375 376 however, that in the present work SVM, as well as RF, were characterized by very large standard 377 deviations associated with all the metrics (Figure 4). This result might alert on the dependency of 378 these techniques on the specific repartition of samples into training and validation sets, thus 379 suggesting a potential instability and lack of robustness towards future prediction of unknown 380 samples.

In conclusion, it can be stated that the data support the hypothesis that the simplest and the most 381 interpretable classifiers (i.e., OPLS-DA and LR) also guarantee the best results in cross validation. 382 383 The reason underlying this finding could be due to the existence of a direct linear rather than indirect correlation between NIR spectral patterns and the geographical origin of octopuses achieved by 384 385 applying optimal spectral pre-processing operations. Therefore, although the complex nature of the 386 samples, this correlation can be easily extrapolated by traditional linear techniques (Zareef et al., 387 2020). Evidence for this theory is however limited to the results obtained throughout the present research and it can be easily supposed that, with increasing sample size and non-linear variability (in 388 389 terms of different fishing seasons, batches, sizes, storage times and temperatures), the more complex 390 and flexible algorithms such as SVM, RF, and MLP-ANN might be the most performing ones. 391 Additionally, it is worth to say that the training of SVM and RF usually involves the identification of 392 the best numerical values to be assigned to building parameters, so as to increase accuracy and 393 performance of the final models. This operation is a very complex task requiring time, expertise, and 394 efforts and, therefore, it does not sit well with the purpose of having a speedy, cost-effective, easy and fully exploitable procedure. In this work, the values for these building parameters (maximum 395 number and depth of trees, minimum size and number of variables included into each child nodes for 396 RF, as well as the C,  $\gamma$ , and  $\varepsilon$  parameters for SVM) were chosen by following the default settings 397 398 recommended by the software and no complex operations such as manual search, use of genetic algorithms or grid search were performed (Phan et al., 2017). 399

400 3.2.1 Comparison of predictive NIR wavelengths

401 Given the different mathematical nature of the models being presented, also the relative strength of 402 each NIR wavelength in guiding the classification of octopus samples based on the geographic 403 provenance is expected be different. For each predictive model computed, a different ranking of NIR 404 wavelengths in terms of their importance (i.e., their relative contribution to the predictive models) 405 was obtained since dependent on the mathematical function employed (see Section 2.3.1). Information about the first ten most influential NIR bands extracted as strong predictors of origin by 406 407 individual cross-validated models obtained by training sets is provided in Table 1. Considering that the training phase was performed four consecutive times by changing sample datasets (Section 2.3), 408 409 the kind and order of importance of the wavelengths were sometimes found to be different based on the dataset considered for modelling. For the sake of conciseness, those extracted by the most accurate 410 411 of the four fitted models were reported.

NIR wavelength absorbance at 1453.2 nm was the most common important variable for musky octopuses, since it was extracted by four out five machine learning algorithms (OPLS-DA, RF, SVM, and MLP-ANN). Similarly, NIR band at 1632.8 had a strong influence on musky octopus samples discrimination for OPLS-DA, LR, and SVM. On the contrary, RF stood out the most from the other models because six out ten wavelengths (1081.5, 1075.3, 1465.6, 1137.3, 1341.7, 1093.9 nm) were

417 exclusive predictors. As for common octopuses, absorbance peaks at 970 and 976.2 nm were shared as important predictors respectively by OPLS-DA, LR, RF, and SVM and by OPLS-DA, RF, SVM, 418 and MLP-ANN, while that located at 963.8 by OPLS-DA, SVM, MLP-ANN (Table 1). 419 420 From the above results, it seems that NIR bands around the 1137.3 and 1341.7 nm, which are potentially related to aliphatic and aromatic hydrocarbons (Workman & Weyer, 2012), were strongly 421 involved in differentiating Atlantic from Mediterranean musky octopuses, thus corroborating what 422 emerged from the visual inspection of the pre-processed spectra discussed in Section 3.1. However, 423 whereas OH-group absorption bands (1453 nm, 1075.3-1093.9 nm, and 963.8-976.2 nm) were 424 425 clearly influent, the additional contribution of water to musky and octopus discrimination by origin should not be overlooked (Workman & Weyer, 2012). Considering that sample processing before 426 NIR recording was standardized and freezing/defrosting as well as acquisition procedures performed 427 428 at the same time/temperature conditions, it could be inferred that the water content varied across 429 specimens to such an extent that inter-origin difference was higher than inter-individual one in both 430 octopus species. Nevertheless, another hypothesis can be derived from the principles of 431 aquaphotomics, according to which, since one single biological compound is solvated by many water molecules, the NIR response to individual biological compounds was amplified by water absorption 432 which, indirectly, contributed to the achievement of high accuracy in prediction (Muncan & 433 434 Tsenkova, 2019).

# 435 3.3. Independent evaluation of the predictivity of machine learning models

The ability of the <u>fitted\_trained</u> models to generalize beyond the training data and confidently assign the correct labels of geographical origin to future candidate octopus samples was estimated by using the <u>samples-25% of data (i.e., one out four spectral replicates for each sample) belonging the</u> <u>validation sets</u>) and previously excluded from the calibration phase (see *Section 2.3*). The resulting label assignments for <u>118 musky</u> and <u>39 spectral data of musky and common, respectively, included</u> <u>into-of</u> the validation sets are reported in the confusion matrices plotted in Figure 5. In agreement to what observed for PCA, but in contrast to results of cross-validation, the absolute best outcomes in 443 external validation were achieved when recognizing the origin of common octopuses: three out five models (OPLS-DA, SVM, and MLP-ANN) predicted both the Mediterranean and Atlantic origin of 444 445 common octopus validation these samples with 100% accuracy (Figure 5). More in detail, OPLS-DA, 446 SVM, and MLP-ANN classifiers were all characterized by mean AUROC values of 1, with OPLS-DA and MLP-ANN also showing the lowest associated standard deviation values for the prediction 447 of Mediterranean and Atlantic common octopus samples, respectively. On the contrary, despite the 448 optimistic results in cross-validation, the analysis of the LR confusion matrix reported in Figure 5 449 450 revealed the poorest performances, since one Atlantic (10%) and nine Mediterranean common 451 octopus validation samples (31%) were misclassified. The associated AUROC values were in fact 452  $0.638 \pm 0.002$  and  $0.631 \pm 0.019$  and, thus, quite close to the randomly guess rate of class membership 453 recognition of 0.5. In this instance, it is important to reiterate that the better best classification of 454 common octopuses compared to musky octopuses is likely to be the consequence of the smaller-sized 455 group which determined the inclusion into the models of a smaller amount of variation available for 456 self-learning which, in turn, hindered finding the best predictive correlation between NIR 457 wavelengths and octopus origins. Hence, definite conclusions on this aspect could not be drawn, but it can be assumed that, although the noisy and collinear nature of the NIR spectra, none of the trained 458 models underwent overfitting, as can be seen from the similarity between training and validation 459 460 results.

As for musky octopuses, no models were able to recognize the provenance with 100% accuracy 461 462 (Figure 5). The maximum correct classification rates in validation were shown by SVM and LR 463 predicting the Mediterranean samples (98 and 97%, respectively) and by LR and MLP-ANN 464 predicting the Atlantic ones (100%). Although SVM and MLP-ANN classifiers had high mean AUROC values, the lowest associated standard deviations were found for LR classifiers (AUROC= 465  $0.989 \pm 0.009$  for Mediterranean samples; AUROC=  $0.992 \pm 0.009$  for Atlantic samples). Shifting 466 467 the focus from machine learning classifiers to single classes (i.e., geographical origins), it can be also 468 noticed that, whatever the octopus species considered, samples from Atlantic Ocean were better 469 recognized than samples from Mediterranean sea. From this finding it could be inferred that the small 470 extension of the water surface and the close proximity to the coast of the Eastern Portuguese waters 471 of the North-East Atlantic Ocean (FAO fishing area 27.9.a) are reflected in a more uniform 472 environment which, in turn, might be responsible for the composition of octopus originating from this area to be more stable and preserved compared to that of octopuses from Western Mediterranean 473 waters (FAO fishing areas 37.1.1). In fact, if on the one hand Western Mediterranean Sea is a semi-474 475 enclosed area characterized by stable temperature and salinity, the North-East Atlantic Ocean is 476 characterized by a continuous input of organic matter from the Portuguese coast. This contributes to 477 the permanent availability of prey and constant accessibility to food, which is the main factor influencing the fatty acid composition of octopuses (Massutí et al., 2004). Indeed, in the present work, 478 the same lipid composition was hypothesized to be determinant in differentiating samples according 479 480 to their country of origin (Section 3.1). As an example, concentrations of monounsaturated and n-6 481 polyunsaturated fatty acids were found to be constantly lower in Eastern Atlantic populations of O. 482 vulgaris than Western Mediterranean ones, while concentrations of total fatty acids and n-3-6 483 polyunsaturated fatty acids higher in Atlantic populations (Arechavala-Lopez et al., 2019; Torrinha 484 et al., 2014).

## 485 4. Conclusions

486 The results achieved from this study indicate that portable NIR sensors a potential integrated analytical platform combining portable and miniature NIR spectroscopy and machine learning might 487 be a suitable solution to can identify with great accuracy the geographical origin of two widely 488 489 consumed octopus species widely consumed in Europe, coming from Mediterranean or Atlantic fishing areas, thus helping fraud prevention and having a direct impact on the quality and safety of 490 the products. Regardless of the classification method employed, equally good results were achieved 491 492 when fitting the models. Nevertheless, musky octopuses (*Eledone* spp.) were better modelled 493 compared to common octopuses (O. vulgaris) when using traditional linear algorithms, (OPLS-DA 494 and LR), thus suggesting the presence of a direct linear relationship between NIR spectra and the

495	provenances of octopuses which can be easily extracted with increasing sample sizes. Following the
496	validation of the fitted models, OPLS-DA, SVM, and MLP-ANN allowed to achieve the maximum
497	label recognition rates of common octopuses, while LR and MLP ANN performed better for musky
498	octopuses. Additionally, rRegardless of the octopus species considered, the origin label estimates
499	were better for the Atlantic sample population compared to the Mediterranean one, probably because
500	of the specific characteristics of the fishing waters, which contributed to make Atlantic population
501	more homogenous from a compositional point of view.
502	In conclusion, the obtained data might be transferred to the fish chain environment and, provided
503	their constant validation, find concrete application for the protection of the reputation of national and
504	regional traditional fisheries. This application may materialize in the direct interface of a portable
505	NIR system to the production flow and its customization based on the products to be handled and the
506	specific industrial facility processes, so as to online monitor quality, authenticity, and safety of
507	cephalopods and contribute to the development of quality certification schemes.
508	Given the promising outcomes, future research will be focused on the creation of multi-class
509	elassification models for the detection of commercial fraud, including additional fishing areas and,
510	possibly, also different species of octopus. In this context, the most important impact under a future
511	perspective would be the setting up of a tool to promote and protect the reputation of national and
512	regional traditional fisheries and which would also help in the development of quality certification
513	<del>schemes.</del>
514	If this is shown to be possible and satisfying results are achieved, then the next step might be the
515	exploitation of the methodology for complementary applications addressing food safety and
516	surveillance, such as the detection of contaminants, residues, and food additives. This way, a single
517	integrated methodology might be used to characterize in a comprehensive way the fishery products
518	found in the marketplace and ensure high quality and safety standards.
1	

519 Declaration of Competing Interest

520	The authors declare that they have no known competing financial interests or personal relationships	
521	that could have appeared to influence the work reported in this paper.	
522		
523	Acknowledgements	
524	The authors gratefully acknowledge Dr. Livio Artale (Proqualys S.r.l, Italy) for providing octopus	
525	samples.	
526		
527	Funding	
528	This work was supported by the University of Parma (Italy).	
529	CRediT author statement	
530	Maria Olga Varrà: Conceptualization; Investigation, Formal analysis, Methodology, Writing	
531	Original Draft; Sergio Ghidini: Conceptualization, Supervision, Methodology, Writing Review &	
532	Editing; Maria Pia Fabrile: Formal analysis, Methodology; Adriana Ianieri: Funding acquisition,	
533	Project administration, Writing Review & Editing; Emanuela Zanardi: Conceptualization,	
534	Supervision, Writing Review & Editing; Project administration.	
535		

- 536 Appendix A. Supplementary materials
- 537 The following are the Supplementary data to this article:

# 538 Table 1

539 Comparison among the ten most important predictive NIR wavelengths used by each machine

learning tool to categorize the octopus samples <u>of the training sets</u> by their origin.

Models Important predictors (wa		rs (wavelengths, nm)*
	Musky octopus	Common octopus
	1453.2; 1459.4; 1447; 1632.8; 1440.8;	982.4; 976.2; 988.6; 970; 963.8; 957.7; 951.5;
OPLS-DA	1564.7; 1558.5; 1626.6; 1570.9; 1434.6	994.8; 1193; 1199.2
	1552.3; 1614.3; 1577.1; 1632.8; 1564.7;	957.7; 951.5; 970; 1304.5; 1298.3; 982.4;
LR	1589.5; 1316.9; 1397.5; 1409.8; 1583.3	1316.9; 1248.8; 1236.4; 1224
	1453.2; 1081.5; 1459.4; 1323.1; 1075.3;	970; 1180.7; 1304.5; 976.2; 1174.5; 1298.3;
RF	1465.6; 1137.3; 1341.7; 1447; 1093.9	1205.4; 1434.6; 1440.8; 1193.0
	1632.8; 1626.6; 1620.5; 1614.3; 1608.1;	1583.3; 1248.8; 963.8; 1316.9; 994.8; 970;
SVM	1453.2; 1601.9; 1595.7; 1589.5; 1583.3	976.2; 1007.2; 982.4; 1180.7
	1453.2; 982.4; 1100.1; 1069.2; 1620.5;	1025.8; 1459.4; 976.2; 1019.6; 988.6; 963.8;
MLP-ANN	1106.3; 1205.4; 1007.2; 1001; 1186.8	1242.6; 1211.6; 1236.4; 1069.2;

541 \* For all the models, wavelengths are sorted in descending order of predictive importance. The reported wavelengths542 refer to the most accurate among the four trained models.

543

#### 544 References

- 545 Arechavala-Lopez, P., Capó, X., Oliver-Codorniú, M., Sillero-Rios, J., & Busquets-Cortés, C.
- 546 (2019). Fatty acids and elemental composition as biomarkers of Octopus vulgaris populations :
- 547 Does origin matter ? *Marine Pollution Bulletin*, *139*, 299–310.
- 548 https://doi.org/10.1016/j.marpolbul.2018.12.048
- 549 Beć, K. B., Grabska, J., & Huck, C. W. (2021). Principles and Applications of Miniaturized Near-
- 550 Infrared (NIR) Spectrometers. *Chemistry A European Journal*, 27(5), 1514–1532.
- 551 https://doi.org/10.1002/chem.202002838
- 552 Benes, E., Bajusz, D., Gere, A., Fodor, M., & Rácz, A. (2020). Comprehensive chemometric
- 553 classification of snack products based on their near infrared spectra. LWT Food Science and
- 554 Technology, 133, 110130. https://doi.org/10.1016/j.lwt.2020.110130
- 555 Bisutti, V., Merlanti, R., Serva, L., Lucatello, L., Mirisola, M., Balzan, S., Tenti, S., Fontana, F.,
- 556 Trevisan, G., Montanucci, L., Contiero, B., Segato, S., & Capolongo, F. (2019). Multivariate
- 557 and machine learning approaches for honey botanical origin authentication using near infrared
- spectroscopy. Journal of Near Infrared Spectroscopy, 27(1), 65–74.
- 559 https://doi.org/10.1177/0967033518824765
- 560 Brennecke, D., Duarte, B., Paiva, F., Caçador, I., & Canning-Clode, J. (2016). Microplastics as
- 561 vector for heavy metal contamination from the marine environment. *Estuarine, Coastal and*
- 562 Shelf Science, 178, 189-195. https://doi.org/10.1016/j.ecss.2015.12.003
- 563 Camacho, M., Herrera, A., Gómez, M., Acosta-Dacal, A., Martínez, I., Henríquez-Hernández, L.
- A., & Luzardo, O. P. (2019). Organic pollutants in marine plastic debris from Canary Islands
- beaches. *Science of the total environment*, 662, 22-31.
- 566 https://doi.org/10.1016/j.scitotenv.2018.12.422
- 567 Cardoso, V. G. K., & Poppi, R. J. (2021). Non-invasive identification of commercial green tea
- 568 blends using NIR spectroscopy and support vector machine. *Microchemical Journal*, 164,
- 569 106052. https://doi.org/10.1016/j.microc.2021.106052

570	Cruz-Tirado, J. P., Lucimar da Silva Medeiros, M., & Barbin, D. F. (2021). On-line monitoring of
571	egg freshness using a portable NIR spectrometer in tandem with machine learning. Journal of
572	Food Engineering, 306, 110643. https://doi.org/10.1016/j.jfoodeng.2021.110643
573	Currò, S., Balzan, S., Serva, L., Boffo, L., Ferlito, J. C., Novelli, E., & Fasolato, L. (2021). Fast and
574	Green Method to Control Frauds of Geographical Origin in Traded Cuttlefish Using a Portable
575	Infrared Reflective Instrument. Foods, 10, 1678. https://doi.org/10.3390/ foods10081678
576	Currò, S., Fasolato, L., Serva, L., Boffo, L., Ferlito, J. C., Novelli, E., & Balzan, S. (2022). Use of a
577	portable near-infrared tool for rapid on-site inspection of freezing and hydrogen peroxide
578	treatment of cuttlefish (Sepia officinalis). Food Control, 132, 108524.
579	https://doi.org/10.1016/j.foodcont.2021.108524
580	De-la-Torre, G. E. (2020). Microplastics: an emerging threat to food security and human
581	health. Journal of food science and technology, 57(5), 1601-1608.
582	https://doi.org/10.1007/s13197-019-04138-1
583	Dos Santos, D. A., Coqueiro, A., Gonçalves, T. R., Carvalho, J. C., Bezerra, J. S., Matsushita, M.,
584	de Oliveira, C. A. L., Março, P. H., Valderrama, P., & Ribeiro, R. P. (2020). Omega-3 and
585	Omega-6 Determination in Nile Tilapia's Fillet Based on MicroNIR Spectroscopy and
586	Multivariate Calibration. Journal of the Brazilian Chemical Society, 31(9), 1883–1890.
587	https://doi.org/10.21577/0103-5053.20200082
588	Dos Santos Pereira, E. V., de Sousa Fernandes, D. D., de Araújo, M. C. U., Diniz, P. H. G. D., &
589	Maciel, M. I. S. (2021). In-situ authentication of goat milk in terms of its adulteration with cow
590	milk using a low-cost portable NIR spectrophotometer. Microchemical Journal, 163.
591	https://doi.org/10.1016/j.microc.2020.105885
592	EFSA CONTAM Panel (EFSA Panel on Contaminants in the Food Chain) (2016). Presence of
593	microplastics and nanoplastics in food, with particular focus on seafood. Efsa Journal, 14(6),
504	204501 https://doi.org/10.2002/i.efaz.2016.4501

- 594 e04501. https://doi.org/10.2903/j.efsa.2016.4501
- 595 Esslinger, S., Riedl, J., & Fauhl-Hassek, C. (2014). Potential and limitations of non-targeted fi

596	ngerprinting for authentication of food in offi cial control. Food Research International, 60,
597	189-204. https://doi.org/10.1016/j.foodres.2013.10.015
598	European Market Observatory for Fisheries and Aquaculture Products (EUMOFA). (2020).
599	Octopus in the EU. Price structure in the supply chain. Focus on Italy, Spain and Greece.
600	Publications Office of the European Union. https://doi.org/10.2771/87203
601	Fawcett, T. (2006). An introduction to ROC analysis. Pattern Recognition Letters, 27(8), 861-874.
602	https://doi.org/10.1016/j.patrec.2005.10.010
603	Fox, M., Mitchell, M., Dean, M., Elliott, C., & Campbell, K. (2018). The seafood supply chain from
604	a fraudulent perspective. Food Security, 10, 939-963. https://doi.org/10.1007/s12571-018-
605	0826-z
606	Freitas, J., Vaz pires, P., Câmara, J.S., 2020. From aquaculture production to consumption :
607	Freshness, safety, traceability and authentication, the four pillars of quality. Aquaculture 518,
608	734857. https://doi.org/10.1016/j.aquaculture.2019.734857
609	Ghidini, S., Varrà, M. O., Dall'Asta, C., Badiani, A., Ianieri, A., & Zanardi, E. (2019). Rapid
610	authentication of European sea bass (Dicentrarchus labrax L.) according to production method,
611	farming system, and geographical origin by near infrared spectroscopy coupled with
612	chemometrics. Food Chemistry, 280, 321-327.
613	https://doi.org/10.1016/j.foodchem.2018.12.075
614	Gomes, F., Oliveira, M., Ramalhosa, M. J., Delerue-Matos, C., & Morais, S. (2013). Polycyclic
615	aromatic hydrocarbons in commercial squids from different geographical origins: levels and
616	risks for human consumption. Food and chemical toxicology, 59, 46-54.
617	https://doi.org/10.1016/j.fct.2013.05.034
618	González-Mohino, A., Pérez-Palacios, T., Antequera, T., Ruiz-Carrascal, J., Olegario, L. S., &
619	Grassi, S. (2020). Monitoring the processing of dry fermented sausages with a portable NIRS
620	device. Foods, 9(9), 1-12. https://doi.org/10.3390/foods9091294

621 Grassi, S., Casiraghi, E., & Alamprese, C. (2018). Handheld NIR device: A non-targeted approach

- 622 to assess authenticity of fish fillets and patties. *Food Chemistry*, 243, 382–388.
- 623 https://doi.org/10.1016/j.foodchem.2017.09.145
- 624 Guardone, L., Tinacci, L., Costanzo, F., Azzarelli, D., Amico, P. D., Tasselli, G., Magni, A., Guidi,
- 625 A., Nucera, D., & Armani, A. (2017). DNA barcoding as a tool for detecting mislabeling of
- fishery products imported from third countries : An official survey conducted at the Border
- 627 Inspection Post of Livorno-Pisa (Italy). *Food Control*, 80, 204–216.
- 628 https://doi.org/10.1016/j.foodcont.2017.03.056
- 629 Guglielmetti, C., Manfredi, M., Brusadore, S., Sciuto, S., Esposito, G., Giuseppe, P., Magnani, L.,
- Gili, S., Marengo, E., Luigi, P., & Mazza, M. (2018). Two-dimensional gel and shotgun
- 631 proteomics approaches to distinguish fresh and frozen-thawed curled octopus (*Eledone*
- 632 cirrhosa). Journal of Proteomics, 186, 1–7. https://doi.org/10.1016/j.jprot.2018.07.017
- 633 Han, C., Choi, H., Jo, S., Na, H., Kim, M. K., Kim, M., & Lee, J. (2020). Development of a 3D
- 634 scanning method to discriminate blocks of Octopus minor with surplus water gain. Food
- 635 *Chemistry*, 303, 125414. https://doi.org/10.1016/j.foodchem.2019.125414
- 536 Jiménez-Carvelo, A. M., González-Casado, A., Bagur-González, M. G., & Cuadros-Rodríguez, L.
- 637 (2019). Alternative data mining/machine learning methods for the analytical evaluation of food
- 638 quality and authenticity A review. *Food Research International*, *122*, 25–39.
- 639 https://doi.org/10.1016/j.foodres.2019.03.063
- 640 Karlson, B., Andersen, P., Arneborg, L., Cembella, A., Eikrem, W., John, U., West, J.J., Kerstin,
- 641 K., Kobos, J., Lehtinen, S., Lundholm, N., Mazur-Marzec, H., Naustvoll, L., Poelman, M.,
- 642 Provoost, P., De rijcke, M. & Suikkanen, S. (2021). Harmful algal blooms and their effects in
- 643 coastal seas of Northern Europe. *Harmful Algae*, 101989.
- 644 https://doi.org/10.1016/j.hal.2021.101989
- Lopes, V. M., Lopes, A. R., Costa, P., & Rosa, R. (2013). Cephalopods as vectors of harmful algal
- bloom toxins in marine food webs. *Marine drugs*, 11(9), 3381-3409.
- 647 https://doi.org/10.3390/md11093381

- 648 Massutí, E., Gordon, J. D. M., Moranta, J., Swan, S. C., Stefanescu, C., & Merrett, N. R. (2004).
- 649 Mediterranean and Atlantic deep-sea fish assemblages: Differences in biomass composition
- and size-related structure. *Scientia Marina*, 68(S3), 101–115.
- 651 https://doi.org/https://doi.org/10.3989/scimar.2004.68s3101
- 652 Mcvey, C., Elliott, C. T., Cannavan, A., Kelly, S. D., Petchkongkaew, A., & Haughey, S. A. (2021).
- 653 Portable spectroscopy for high throughput food authenticity screening: Advancements in
- 654 technology and integration into digital traceability systems. Trends in Food Science &
- 655 *Technology*, *118*, 777–790. https://doi.org/10.1016/j.tifs.2021.11.003
- 656 Mendes, R., Schimmer, O., Vieira, H., & Teixeira, B. (2018). Control of abusive water addition to
- 657 Octopus vulgaris with non-destructive methods. Journal of the Science of Food and
- 658 *Agriculture*, 98, 369–376. https://doi.org/10.1002/jsfa.8480
- Michelucci, U., & Venturini, F. (2021). Estimating Neural Network<sup>2</sup> s Performance with Bootstrap:
  A Tutorial. *Machine Learning and Knowledge Extraction*, *3*, 357–373.
- 661 https://doi.org/10.3390/make3020018
- 62 Müller-Maatsch, J., Alewijn, M., Wijtten, M., & Weesepoel, Y. (2021a). Detecting fraudulent
- 663 additions in skimmed milk powder using a portable, hyphenated, optical multi-sensor approach
- in combination with one-class classification. *Food Control*, 121, 107744.
- 665 https://doi.org/10.1016/j.foodcont.2020.107744
- 666 Müller-Maatsch, J., Bertani, F. R., Mencattini, A., Gerardino, A., Martinelli, E., Weesepoel, Y., &
- 567 Van Ruth, S. (2021b). The spectral treasure house of miniaturized instruments for food safety,
- quality and authenticity applications: A perspective. *Trends in Food Science and Technology*,
- 669 *110*, 841–848. https://doi.org/10.1016/j.tifs.2021.01.091
- 670 Muncan, J., & Tsenkova, R. (2019). Aquaphotomics-From Innovative Knowledge to Integrative
- 671 Platform in Science and Technology. *Molecules*, 24(15), 2742.
- 672 https://doi.org/10.3390/molecules24152742
- 673 Nowak, M., Wietecha-pos, R., & Pawliszyn, J. (2021). White Analytical Chemistry: an approach to

- 674 reconcile the principles of Green Analytical Chemistry and functionality. *Trends in Analytical*
- 675 *Chemistry*, *138*, 116223. https://doi.org/10.1016/j.trac.2021.116223
- 676 Oliveira, M., Gomes, F., Torrinha, Á., João, M., Delerue-matos, C., & Morais, S. (2018).
- 677 Commercial octopus species from different geographical origins : Levels of polycyclic
- aromatic hydrocarbons and potential health risks for consumers. *Food and Chemical*
- 679 *Toxicology*, *121*, 272–282. https://doi.org/10.1016/j.fct.2018.09.012
- 680 Parastar, H., van Kollenburg, G., Weesepoel, Y., van den Doel, A., Buydens, L., & Jansen, J.
- 681 (2020). Integration of handheld NIR and machine learning to "Measure & Monitor" chicken
- 682 meat authenticity. Food Control, 112, 107149. https://doi.org/10.1016/j.foodcont.2020.107149
- 683 Pennisi, F., Giraudo, A., Cavallini, N., Esposito, G., Merlo, G., Geobaldo, F., Acutis, P. L.,
- 684 Pezzolato, M., Savorani, F., & Bozzetta, E. (2021). Differentiation between fresh and thawed
- 685 cephalopods using NIR spectroscopy and multivariate data analysis. *Foods*, 10(3), 1–14.
- 686 https://doi.org/10.3390/foods10030528
- 687 Phan, A. V., Nguyen, M. Le, & Bui, L. T. (2017). Feature weighting and SVM parameters
- 688 optimization based on genetic algorithms for classification problems. Applied Intelligence,
- 689 46(2), 455–469. https://doi.org/10.1007/s10489-016-0843-6
- 690 Pita, C., Roumbedakis, K., Fonseca, T., Matos, F. L., Pereira, J., Villasante, S., Pita, P., Bellido, J.
- 691 M., Gonzalez, A. F., García-Tasende, M., Lefkaditou, E., Adamidou, A., Cuccu, D., Belcari,
- 692 P., Moreno, A., & Pierce, G. J. (2021). Fisheries for common octopus in Europe:
- 693 socioeconomic importance and management. *Fisheries Research*, 235, 105820.
- 694 https://doi.org/10.1016/j.fishres.2020.105820
- 695 Rinnan, Å., Berg, F. van den, & Engelsen, S. B. (2009). Review of the most common pre-
- 696 processing techniques for near-infrared spectra. In *TrAC Trends in Analytical Chemistry*, 28,
- 697 1201–1222. https://doi.org/10.1016/j.trac.2009.07.007
- 698 Ríos-Reina, R., Camiña, J. M., Callejòn, R. M., & Azcarate, S. M. (2021). Trends in Analytical
- 699 Chemistry Spectralprint techniques for wine and vinegar characterization, authentication and

quality control: Advances and projections. *Trends in Analytical Chemistry*, 134, 116121.

701 https://doi.org/10.1016/j.trac.2020.116121

- 702 Rjeibi, M., Metian, M., Hajji, T., Guyot, T., Chaouacha-Chékir, R. B., & Bustamante, P. (2014).
- 703 Interspecific and geographical variations of trace metal concentrations in cephalopods from
- Tunisian waters. *Environmental monitoring and assessment*, 186(6), 3767-3783.
- 705 https://doi.org/10.1007/s10661-014-3656-2
- 706 Roldán-Wong, N. T., Kidd, K. A., Ceballos-Vázquez, B. P., & Arellano-Martínez, M. (2018). Is
- 707 there a risk to humans from consuming octopus species from sites with high environmental
- levels of metals?. *Bulletin of environmental contamination and toxicology*, *101*(6), 796-802.
- 709 https://doi.org/10.1007/s00128-018-2447-9
- Saito, H., Ishihara, K., & Murase, T. (1997). The fatty acid composition in tuna (bonito, *Euthynnus pelamis*) caught at three different localities from tropics to temperate. *Journal of the Science of*
- 712 Food and Agriculture, 73(1), 53–59. https://doi.org/10.1002/(SICI)1097-

713 0010(199701)73:1<53::AID-JSFA707>3.0.CO;2-5

- 714 Sampaio, P. S., Castanho, A., Almeida, A. S., Oliveira, J., & Brites, C. (2020). Identification of rice
- flour types with near-infrared spectroscopy associated with PLS-DA and SVM methods.
- *European Food Research and Technology*, 246(3), 527–537. https://doi.org/10.1007/s00217-

717 019-03419-5

- 718 Semedo, M., Reis-Henriques, M. A., Rey-Salgueiro, L., Oliveira, M., Delerue-Matos, C., Morais,
- 719 S., & Ferreira, M. (2012). Science of the Total Environment Metal accumulation and oxidative
- 720 stress biomarkers in octopus (Octopus vulgaris) from Northwest Atlantic. Science of the Total
- 721 Environment, 433, 230–237. https://doi.org/10.1016/j.scitotenv.2012.06.058
- 722 Sillero-Ríos, J., Sureda, A., Capó, X., Oliver-Codorniú, M., & Arechavala-Lopez, P. (2018).
- 723 Biomarkers of physiological responses of Octopus vulgaris to different coastal environments in
- the western Mediterranean Sea. *Marine Pollution Bulletin*, *128*, 240–247.
- 725 https://doi.org/10.1016/j.marpolbul.2018.01.032

- 726 Silva, L. C. R., Folli, G. S., Santos, L. P., Barros, I. H. A. S., Oliveira, B. G., Borghi, F. T., Santos,
- 727 F. D. do., Filgueiras, P. R., & Romão, W. (2020). Quantification of beef, pork, and chicken in
- ground meat using a portable NIR spectrometer. *Vibrational Spectroscopy*, 111.
- 729 https://doi.org/10.1016/j.vibspec.2020.103158
- 730 Stadler, R. H., Tran, L. A., Cavin, C., Zbinden, P., & Konings, E. J. M. (2016). Analytical
- approaches to verify food integrity: Needs and challenges. *Journal of AOAC International*,
- 732 99(5), 1135–1144. https://doi.org/10.5740/jaoacint.16-0231
- 733 Tatulli, G., Cecere, P., Maggioni, D., Galimberti, A., & Pompa, P. P. (2020). A Rapid Colorimetric
- Assay for On-Site Authentication of Cephalopod Species. *Biosensors*, 10(190).
- 735 https://doi.org/10.3390/bios10120190
- 736 Tinacci, L., Armani, A., Scardino, G., Guidi, A., Nucera, D., Miragliotta, V., & Abramo, F. (2020).
- 737 Selection of Histological Parameters for the Development of an Analytical Method for
- 738 Discriminating Fresh and Frozen / Thawed Common Octopus (Octopus vulgaris) and
- 739 Preventing Frauds along the Seafood Chain. Food Analytical Methods, 13, 2111–2127.
- 740 https://doi.org/10.1007/s12161-020-01825-0
- 741 Torrinha, A., Cruz, R., Gomes, F., Casal, S., & Morais, S. (2014). Octopus Lipid and Vitamin E
- 742 Composition: Interspecies, Interorigin, and Nutritional Variability. Journal of Agricultural and
- 743 Food Chemistry, 62, 8508–8517. https://doi.org/10.1021/jf502502b
- 744 Varrà, M. O., Ghidini, S., Husáková, L., Ianieri, A., & Zanardi, E. (2021a). Advances in
- 745 Troubleshooting Fish and Seafood Authentication by Inorganic Elemental Composition.
- 746 Foods, 10, 270. https://doi.org/10.3390/foods10020270
- 747 Varrà, M. O., Ghidini, S., Ianieri, A., & Zanardi, E. (2021b). Near infrared spectral fingerprinting:
- A tool against origin-related fraud in the sector of processed anchovies. *Food Control*, 123.
- 749 https://doi.org/10.1016/j.foodcont.2020.107778
- 750 Whitelaw, B. L., Cooke, I. R., Finn, J., Zenger, K., & Strugnell, J. M. (2019). The evolution and
- 751 origin of tetrodotoxin acquisition in the blue-ringed octopus (genus Hapalochlaena). Aquatic

- 752 *Toxicology*, 206, 114-122. https://doi.org/10.1016/j.aquatox.2018.10.012
- 753 Workman, J. ., & Weyer, L. (2012). Practical Guide and Spectral Atlas for Interpretive Near-
- 754 Infrared Spectroscopy (2nd ed.). CRC Press (Taylor & Francis group).
- 755 Yakes, B. J., Ellsworth, Z., Karunathilaka, S. R., & Crump, E. (2021). Evaluation of Portable
- 756 Sensor and Spectroscopic Devices for Seafood Decomposition Determination. *Food Analytical*
- 757 *Methods*, 14, 2346–2356. https://doi.org/10.1007/s12161-021-02064-7
- 758 Yu, H. D., Zuo, S. M., Xia, G., Liu, X., Yun, Y. H., & Zhang, C. (2020). Rapid and Nondestructive
- 759 Freshness Determination of Tilapia Fillets by a Portable Near-Infrared Spectrometer Combined
- with Chemometrics Methods. *Food Analytical Methods*, *13*(10), 1918–1928.
- 761 https://doi.org/10.1007/s12161-020-01816-1
- 762 Zareef, M., Chen, Q., Hassan, M. M., Arslan, M., Hashim, M. M., Ahmad, W., Kutsanedzie, F. Y.
- 763 H., & Agyekum, A. A. (2020). An Overview on the Applications of Typical Non-linear
- 764 Algorithms Coupled With NIR Spectroscopy in Food Analysis. *Food Engineering Reviews*,
- 765 *12*(2), 173–190. https://doi.org/10.1007/s12393-020-09210-7

766

## 767 Figure captions

- 768 Figure 1. Sampling area extensions of musky and common octopuses.
- **Figure 2.** Effect of the application of different pre-processing filters (<u>Standard Normal Variate, SNV</u>;
- <sup>770</sup> first derivative, 1<sup>st</sup> Der) on the quality and usefulness of the 908.1–1676.2 nm spectra recorded by
- 771 MicroNIR (Atlantic samples: red lines; Mediterranean samples: red lines). The main spectral features
- 772 in SNV +  $1^{st}$  Der spectral patterns are highlighted by dotted marker boxes.
- **Figure 3.** 3-D score scatter plot from PCA applied to musky and common octopuses.
- **Figure 4** Main figures of merit (mean ± standard deviation) of the five different machine learning
- tools (OPLS-DA, LR, RF, SVM, and RF) obtained by cross-validation of the training samples for the
- characterization of the geographic origin of musky and common octopuses.
- **Figure 5.** Confusion matrices (mean classification rates) and corresponding <u>areas under the curve</u>
- 778 (AUC) values of receiver operating characteristic (AUROC) AUROC values (mean ± standard
- deviation) resulting from the origin prediction of <u>118</u> musky and <u>39</u> common octopus test-spectral
- samples (included into the external validation sets) by the five different classifiers (OPLS-DA, LR,
- 781 RF, SVM and MLP-ANN). Correct classification rates in confusions matrices are included into green
- 782 <u>boxes (Med: Mediterranean samples, Atl: Atlantic samples).</u>