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Intestinal dysfunction in Parkinson’s disease: Lessons learned from translational studies and experimental models

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3 **Title page**

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5 **Intestinal dysfunction in Parkinson's disease: Lessons learned from**
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7 **translational studies and experimental models**

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Abstract

Background Symptoms of digestive dysfunction in patients with Parkinson's disease (PD) occur at all stages of the disease, often preceding the onset of central motor symptoms. On the basis of these PD-preceding symptoms it has been proposed that PD could initiate in the gut, and that the presence of alpha-synuclein aggregates, or Lewy Bodies (LBs) in the enteric nervous system might represent one of the earliest signs of the disease. Following this hypothesis, much research has been focused on the digestive tract to unravel the mechanisms underlying the onset and progression of PD, with particular attention to the role of alterations in enteric neurotransmission in the pathophysiology of intestinal motility disturbances. There is also evidence suggesting that the development of central nigrostriatal neurodegeneration is associated with the occurrence of gut inflammation, characterized by increments of tissue pro-inflammatory markers and oxidative stress, which might support conditions of bowel neuromotor abnormalities.

Purpose The present review intends to provide an integrated and critical appraisal of the available knowledge on the alterations of enteric neuromuscular pathways regulating gut motor activity both in humans and pre-clinical models of PD. Moreover, we will discuss the possible involvement of neuro-immune mechanisms in the pathophysiology of aberrant gastro intestinal gut transit and neuromuscular activity in the small and large bowel.

Keywords

Parkinson's disease, colonic motility, small bowel alterations, pre-clinical model, patients

1. Introduction

Parkinson's disease (PD) is the second most common neurodegenerative disorder in the general population, with a prevalence of 1% at the age of 55 years, which increases with aging, resulting in an accumulating burden for healthcare. Nigrostriatal dopaminergic loss of neurons, the pathological hallmark of PD, triggers complex functional alterations within the basal ganglia circuitry, which then cause typical motor symptoms (tremor, rigidity, bradykinesia) (1). However, PD is associated also with other peripheral symptoms, that include functional gastrointestinal (GI) abnormalities, consisting mainly of delayed gastric emptying, constipation and anorectal dysfunction (2-4).

GI motility dysfunction often precedes the onset of motor symptoms in patients with PD by many years (5). Furthermore, Lewy bodies (LBs), the typical α -synuclein (α -syn) positive inclusions found in PD brain, have been detected also in neurons of the myenteric plexus and dorsal motor nucleus of the vagus (DMV) of PD patients, suggesting that the underlying pathological process involves also the autonomic nervous system (6). Interestingly, Del Tredici and colleagues postulated that the gut could represent the starting site of PD, and that the presence of LBs in the enteric nervous system (ENS) might be one of the earliest signs of the disease (7). In this context, research efforts are currently focused on understanding the pathophysiological mechanisms underlying gut neuromuscular dysfunctions associated with PD. The present review intends to provide an appraisal of the available knowledge about enteric functional, neurochemical and molecular alterations associated with PD. Special attention has been paid to the pre-clinical PD models would translate to human PD pathophysiology and what role the enteric cell network plays in the pathophysiology of bowel disorders associated with PD.

2. Intestinal enteric abnormalities in PD patients

Infrequent bowel movements and constipation represent the main intestinal disturbances in patients with PD (8). Such disorders may occur both in the early and advanced stages of the disease, worsening the patients' quality of life (9). There is an increasing recognition of the involvement of cells of the enteric neuromuscular compartment in this process (i.e. intrinsic primary afferent neurons, enteric glia, interstitial cells of Cajal), that contribute actively to the regulation of small bowel functions. A number of clinical investigations have attempted to unravel the pathophysiological mechanisms underlying bowel alterations occurring in PD patients. Data on neurochemical, molecular and functional enteric changes in PD patients are discussed in the following sections and summarized in Table 1.

2.1 Small bowel

Little is known about the pathophysiology of small bowel dysmotility associated with PD. Even though the presence of α -syn accumulation has been detected in duodenal and ileal biopsies from PD patients at different stages of the disease, the same biopsies did not reveal any myenteric ganglion loss or alterations of nitric oxide (NO), vasoactive intestinal peptide (VIP), dopamine and catecholamine neuronal density (10-12). Increasingly, the role of enteric luminal microbiota in GI dysfunction associated with brain neurodegenerative and inflammatory diseases (i.e. Alzheimer's disease, PD and multiple sclerosis) is recognized (13, 14). At present, there is some clinical evidence supporting the contribution of enteric bacteria to the alterations of intestinal motility associated with PD. In particular, the occurrence of both small intestine bacterial overgrowth (SIBO) and malabsorption syndrome, characterized by increased bacterial density in the small bowel, with frequent impairment of GI motility, have been observed in PD patients (15, 16). However, the actual involvement of intestinal microbial flora in the pathogenesis of GI abnormalities in PD remains to be elucidated and,

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3 most importantly, there is a substantial lack of data on the possible role of the brain-gut-
4
5 microbiota axis in the pathophysiology of PD. Moreover, it remains unclear whether bacterial
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7 overgrowth acts as a possible pathophysiological mechanism, or whether it rather occurs as a
8
9 mere consequence of the impaired small bowel motility. These are open issues, which
10
11 represent areas of interest for future investigations.
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14 15 16 *2.2 Large bowel*

17
18 Chronic constipation is the most widely recognized functional gut disorder associated with
19
20 PD. It occurs with a prevalence ranging from 70 to 80% and precedes the onset of movement
21
22 disorders in 87% of PD patients (5), with negative impact on the patient's quality of life and
23
24 increased health care costs. PD patients with constipation are characterized by infrequent
25
26 bowel movements, impairment of propulsive colonic motility and prolonged colonic transit
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28 time as well as reduced rectal contractions and abnormalities in motor activity of the anal
29
30 sphincter (17).
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33
34 The pathogenic mechanisms underlying the development of colonic constipation associated
35
36 with PD are presently unclear, as most, if not all, studies fail to demonstrate changes in the
37
38 neurochemical and/or innervation pattern in intestinal tissue of PD patients. An initial study of
39
40 human colonic morphological and molecular changes associated with PD was provided by
41
42 Singaram et al. (18). These authors performed a detailed quantification of submucosal and
43
44 myenteric dopaminergic neurons by labeling both TH and dopamine. They also assessed the
45
46 density of VIP neurons, substance P and neuropeptide Y. Their results showed that, in the
47
48 submucosal plexus, the number of both TH and dopamine containing neurons did not differ
49
50 from that observed in control tissues, while in the myenteric plexus the number of
51
52 immunopositive neurons for dopamine were reduced, without significant changes in TH
53
54 immunostaining. In both plexuses, there were no significant variations seen in the density of
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3 VIP-ergic neurons, SP and neuropeptide Y. The myenteric decrease in the density of
4
5 dopaminergic neurons was associated with a reduced dopamine content in the muscularis
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7 externa, while no differences were detected in the mucosal layer, suggesting an impairment of
8
9 dopaminergic neurotransmission in the PD human colon (18). Lebouvier et al. (19) confirmed
10
11 the lack of significant changes in the density of submucosal dopaminergic neurons in colonic
12
13 biopsies from PD patients. Subsequent investigations by Annerino et al. (11), consistent with
14
15 the findings reported by Lebouvier et al. (19), did not observe any significant change in the
16
17 proportion of myenteric nitrenergic, VIP-ergic or dopaminergic neurons. Likewise, Corbille et
18
19 al. (20) did not find abnormalities in the density of dopaminergic neurons as well as the
20
21 expression levels of dopaminergic and noradrenergic markers in colonic biopsies from PD
22
23 patients. Taken together, most of current evidence suggests that colonic dopamine, nitrenergic
24
25 and peptidergic pathways are not significantly affected in PD patients. It is interesting to note
26
27 that neurochemical evidence from non-PD patients with chronic constipation showed distinct
28
29 profiles of neurotransmitter levels in colonic neuromuscular compartment as compared to PD
30
31 patients, characterized by a decreased VIP and substance P expression and enhanced nitrenergic
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33 innervation (21).
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39 When considering the distribution and localization of LBs and Lewy neurites in colonic
40
41 tissues from PD patients, the presence of α -syn inclusions in colonic neurons has been
42
43 recently documented at all stages of the disease, suggesting that their accumulation could be
44
45 viewed as a biomarker of PD (22). However, clear relationships between colonic α -syn
46
47 accumulation and bowel neuromuscular dysfunctions during PD have not been
48
49 mechanistically established and should be investigated in the near future.
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52 Over the last years, growing interest has been dedicated to the role played by central and
53
54 peripheral neuroinflammation in the pathophysiology of neurodegenerative diseases (23). Of
55
56 note, recent studies, showing a significant increase in pro-inflammatory cytokine levels and
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3 enteric glial activation in colonic biopsies from PD patients, suggest that also enteric
4
5 inflammation could play a key role in the pathogenesis of bowel dysfunctions associated with
6
7 PD (22, 24, 25). In a recent paper, Keshavarzian et al. (26) analyzed the fecal and mucosal
8
9 colonic microbiota compositions and showed that PD patients are characterized by a condition
10
11 of dysbiosis and inflammation. In particular, PD patients displayed decreased abundance of
12
13 “anti-inflammatory” butyrate-producing bacteria, concomitant with increased levels of “pro-
14
15 inflammatory” Proteobacteria of the genus *Ralstonia*, that could contribute to promote bowel
16
17 inflammatory/immune responses during PD (26, 27). In addition, Clairembault et al.(28) have
18
19 observed morphological alterations of the intestinal epithelial barrier, characterized by
20
21 reduced levels of occludin in colonic tissues of PD patients (28). Taken together, these
22
23 observations suggest that alterations of the enteric microbiome, along with changes in
24
25 intestinal permeability, could contribute to the occurrence of intestinal inflammation observed
26
27 in PD patients.
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31 Overall, current knowledge points out the presence of aggregated α -syn in enteric neurons,
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33 intestinal dysbiosis and the occurrence of inflammatory activity in colonic tissues of PD
34
35 patients, even though these alterations have not yet been linked to the pathogenesis of colonic
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37 motor dysfunctions. It remains also to be conclusively demonstrated whether, and to what
38
39 extent, impairment of enteric neurotransmission, as well as the immune/inflammatory
40
41 responses could contribute to colonic dysmotility in PD.
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46 47 **3. Bowel alterations in pre-clinical models of PD**

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49 In an attempt to better understand the pathophysiological mechanisms underlying bowel
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51 dysmotility occurring in patients with PD, efforts were made to study gut dysfunctions in pre-
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53 clinical models of PD. Current models allow two different approaches: 1) peripheral induction
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55 of PD-like pathological alterations by systemic administration of neurotoxins; 2) induction of
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3 nigrostriatal denervation by central injection of neurotoxins. The main features of PD models
4
5 employed in studies on the evaluation of GI dysfunctions are summarized in Table 2. In the
6
7 following sections, data available on functional, neurochemical and molecular intestinal
8
9 alterations in pre-clinical models of PD are discussed.
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11 12 13 14 *3.1 Small bowel*

15 16 *3.1.1 Functional alterations*

17
18 Current evidence, concerning motor dysfunctions occurring in the small intestinal tract in the
19
20 presence of PD, is fragmentary and often conflicting (Table 3). In the peripheral model of
21
22 dopaminergic degeneration induced by 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP,
23
24 which blocks mitochondrial complex I activity into dopaminergic neurons of the substantia
25
26 nigra pars compacta) in mice, Anderson et al. (29) did not detect any significant alteration in
27
28 small intestinal transit. In PD induced by central injection of toxins, only one study, showing
29
30 alterations of intestinal transit, is available. In particular, a delayed transit in the distal region
31
32 of small bowel has been observed by means of geometric center analysis in rats injected with
33
34 6-OHDA, suggesting that central dopaminergic denervation is associated with impaired small
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36 bowel motility (30). However, given the paucity of data, extensive investigations are needed
37
38 to better understand the impact of central dopaminergic neurodegeneration on the onset of
39
40 intestinal dysmotility.
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47 *3.1.2 Neurochemical and molecular alterations*

48 49 *3.1.2.1 Inhibitory pathways*

50
51 Most of the available knowledge about the alterations of enteric neurotransmission in small
52
53 bowel during PD concerns changes of the dopaminergic network in both peripheral and
54
55 central models of dopaminergic neurodegeneration (Table 3, Figure 1). In particular, mice
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3 treated with MPTP displayed a significant decrease in the number of dopamine transporter
4 (DAT) and tyrosine hydroxylase (TH)-positive neurons in both myenteric and submucosal
5 plexus of the duodenum and ileum (29). Likewise, molecular analysis showed a reduced
6 protein expression of TH and DAT in the duodenum of MPTP-treated mice (31). These
7 findings are in keeping with a subsequent report by Natale et al. (32), who observed a
8 significant reduction of dopamine content in the duodenum of MPTP-treated mice, suggesting
9 that this peripheral model is characterized by an impairment of small bowel dopaminergic
10 neurotransmission.
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21 When considering PD elicited by central toxins, only one study examined the alterations of
22 dopaminergic markers in intestinal tissues from rats with 6-hydroxydopamine (6-OHDA)-
23 induced PD. This work showed a significant increase in TH and DAT expression (31), in
24 contrast to data obtained from the MPTP model, where a decrease in enteric dopaminergic
25 markers was evident. These differential rearrangements of the enteric dopaminergic network
26 could depend on intrinsic differences of PD models in different animal species, which might
27 also reflect different pathophysiological stages of the disease.
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There is scarce evidence on the neurochemical modifications of enteric nitrergic and VIP-
ergic inhibitory nerve pathways in the presence of PD. Of note, no changes in the number of
both enteric nitrergic and VIP-ergic neurons were detected upon central denervation induced
by peripheral toxins (29, 32-34). In the setting of 6-OHDA-induced nigrostriatal neuro-
degeneration, only two studies have examined the possible alterations of nitrergic and VIP-
ergic pathways in the small intestine. Colucci et al. (35) observed a decrease in the density of
neuronal nitric oxide synthase (nNOS) positive neurons along with an increased percentage of
VIP-immunoreactive neurons in the myenteric plexus of distal ileum, while Toti and Travagli
(36) showed an increase in the density of nNOS neurons in the duodenum. These results
suggest that changes in the expression of nitrergic neuronal markers could vary depending on

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3 the intestinal region considered. However, additional investigations are needed to better
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5 understand the involvement of nitrergic pathways in intestinal dysfunctions associated with
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7 PD.
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10 11 12 *3.1.2.2 Excitatory pathways*

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14 Information on the alterations of small bowel cholinergic excitatory neurotransmission in
15
16 experimental models of PD are quite limited and heterogeneous (Table 3). For instance, no
17
18 changes in the number of myenteric cholinergic neurons were observed in the duodenum and
19
20 ileum of mice treated with MPTP (29, 32). In the 6-OHDA model of central dopaminergic
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22 degeneration, Colucci et al. (35) did not detect any variation in the number of cholinergic
23
24 enteric neurons, while Toti and Travagli (36) reported a decrease of this neuronal population,
25
26 likely as a result of the different bowel regions and timing. Indeed, Toti and Travagli (36)
27
28 performed their experiments on the duodenum 5 weeks after the induction of central
29
30 neurodegeneration, while Colucci et al. (35) performed their assays in the distal ileum from
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32 rats sacrificed 4 weeks after 6-OHDA injection. Thus, it appears that in this model the
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34 alterations of enteric cholinergic neurons are more likely to occur in the proximal sections of
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36 small bowel.
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43 *3.1.3 Discussion*

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45 Taken together, current data suggest that in the presence of central dopaminergic
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47 neurodegeneration the small intestinal transit is impaired, likely as a consequence of
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49 alterations in the chemical coding of enteric inhibitory neurons. However, there is a lack of
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51 consistent data concerning the involvement of cholinergic, nitrergic and VIP-ergic pathways
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53 in small bowel dysfunctions during PD. Likewise, it remains still unclear whether enteric
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55 dopamine plays a role in intestinal alterations associated with central dopaminergic
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3 neurodegeneration, since a characterization on the physiological role of dopamine in the
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5 control of small bowel motility is still lacking. Furthermore, there is no detailed information
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7 about the pathophysiological mechanisms underlying the development of intestinal
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9 dysmotility in PD. Based on this picture, the effects of central neurodegeneration on small
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11 bowel motor abnormalities remain a field largely opened to future investigations.
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14 15 16 3.3 Large bowel

17 18 3.3.1 Functional alterations

19
20 A number of studies have evaluated the abnormalities of colonic motility associated with
21
22 central dopaminergic neurodegeneration (Table 3). Two reports described a delay of *in vivo*
23
24 colonic transit and constipation in rotenone- and MPTP-induced neurodegeneration in mice
25
26 (32, 33). Functional *in vitro* experiments demonstrated an increase in contractile activity and
27
28 an impaired relaxation in the proximal colon from MPTP-treated mice, suggesting an
29
30 alteration in the inhibitory control of colonic motility (29, 33). Likewise, in the central 6-
31
32 OHDA model, a significant decrease in stool frequency and delay in colonic transit were
33
34 detected (37, 38). In addition, an *ex-vivo* analysis of peristalsis displayed an altered pattern of
35
36 colonic longitudinal muscle contraction and a reduced peak pressure, suggesting that central
37
38 dopaminergic neurodegeneration is associated with an impairment of colonic motility
39
40 resulting in a reduced efficiency of the peristaltic reflex (35). These findings have been
41
42 corroborated by recent *in vitro* functional studies, showing an impairment of colonic
43
44 spontaneous contraction and a decrease in electrically evoked cholinergic contractions of both
45
46 longitudinal and circular colonic smooth muscle from 6-OHDA rats (38, 39). However, in the
47
48 same model, Pellegrini et al. (40) described also an enhanced *in vitro* longitudinal colonic
49
50 tachykininergic contractile activity. This apparent discrepancy with a reduced peristaltic
51
52 activity can be explained by the fact that the patterns of *in vivo* colonic transit do not
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necessarily reflect the contractile responses elicited by specific stimulation of the excitatory tachykininergic pathway, but rather can be viewed as a result of an integrated combination of smooth muscle contractions regulated by both excitatory and inhibitory neurotransmitter pathways.

3.2 Neurochemical and molecular alterations

3.2.1 Inhibitory pathways

Changes in both inhibitory and excitatory enteric pathways regulating colonic motility in experimental PD have been evaluated by morphological investigations (Table 3). In the PD model elicited by rotenone (an isoflavone insecticide), no changes in nitrergic, VIP-ergic and dopaminergic enteric neuron number or innervation quality were detected (33). By contrast, in the setting of PD induced by intranigral injection of 6-OHDA, alterations of inhibitory enteric pathways were observed. In particular, increments of VIP-ergic and dopaminergic neurons in concomitance with a decrease in nitrergic neurons were detected in the proximal colon (35, 37, 41). Colucci et al. (35) showed that dopamine D₂ receptors were mostly expressed in enteric cholinergic and dopaminergic neurons, and that their immunoreactivity was markedly reduced in myenteric neurons of both proximal and distal colon. In addition, in two previous studies (39, 42) an increase in dopamine content was observed in the colonic muscularis externa of 6-OHDA rats, which could contribute to colonic dysmotility in PD. Zhang et al. (39) observed also an increase in adrenergic β_3 receptor expression and a decreased expression of serotonergic 5-HT₄ receptors, suggesting that neurochemical and molecular changes affecting the enteric pathways might contribute to colonic dysmotility and constipation displayed by 6-OHDA rats.

Additional support to the concept that alterations of inhibitory dopaminergic and nitrergic pathways can contribute to the occurrence of colonic dysfunctions associated with

1
2
3 experimental PD has been provided by molecular studies. When considering the
4
5 dopaminergic network, a study by Tian et al. (31) showed that, in the MPTP mouse model,
6
7 the colonic TH protein levels were decreased, while in the 6-OHDA rat model an increase in
8
9 the protein levels of TH and DAT was found. The latter observations are consistent with those
10
11 reported by Zhu et al. (37), who showed an increase in TH protein levels in the proximal
12
13 colon of 6-OHDA rats, suggesting that differential changes in the colonic dopaminergic
14
15 system can occur depending on the different experimental models of PD. Interestingly, central
16
17 dopaminergic denervation is associated also with an impairment of colonic nitrenergic
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19 pathways, as documented by a significant decrease in nNOS protein levels in the proximal
20
21 colon of PD rats (37).
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27 *3.2.1 Excitatory pathways*

28
29 At present, there is inconsistent data on putative changes occurring in the colonic excitatory
30
31 cholinergic system during PD (Table 3, Figure 2). Only one study has shown alterations of
32
33 cholinergic enteric neurons in the PD model induced by peripheral toxins. In particular, in the
34
35 rotenone model, ChAT immunoreactivity was significantly decreased in the myenteric plexus
36
37 after 1 and 6 weeks from toxin infusion (43). When considering PD induced by 6-OHDA, the
38
39 evidence showing changes in the cholinergic excitatory pathway are fragmentary and
40
41 conflicting. Two studies did not show alterations in the density of ChAT-positive neurons, as
42
43 well as in ChAT protein and mRNA expression levels in the proximal colon from 6-OHDA
44
45 rats (35, 37). Conversely, in a recent paper by Fornai et al. (38) a decrease in ChAT immune
46
47 positivity in the distal colon of 6-OHDA rats was detected. Furthermore, a significant
48
49 decrease in acetylcholine release from colonic preparations was observed, suggesting an
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51 impairment of colonic cholinergic neurotransmission in the presence of central dopaminergic
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53 neurodegeneration. In the same study, additional molecular analysis showed an increased
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3 expression of muscarinic M₂ and M₃ receptors in the colonic neuromuscular layer and isolated
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5 colonic smooth muscle cells from 6-OHDA rats, thus indicating an up-regulation of muscular
6
7 muscarinic receptors as a compensatory response to the impairment of cholinergic
8
9 neurotransmission (38). Consistently with these findings, *in vitro* functional experiments
10
11 displayed an impaired colonic motility in PD rats (38). More recently, Pellegrini et al. (40)
12
13 observed significant changes in the tachykininergic pathway of the colonic neuromuscular
14
15 layer from 6-OHDA rats. In particular, nigrostriatal degeneration was followed by an increase
16
17 in endogenous substance P expression in myenteric ganglionic neurons, along with an
18
19 enhancement of NK₁ receptors in longitudinal muscle cells, thus indicating that the enteric
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21 excitatory tachykininergic pathway can be affected also by central dopaminergic
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23 neurodegeneration.
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29 3.1.3 Discussion

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31 An overall appraisal of current data suggest that nigrostriatal neurodegeneration is followed
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33 by an impairment of distal colonic motility, leading to a reduced efficiency of peristaltic
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35 reflex. Such alterations seem to result from a rearrangement in the chemical coding of enteric
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37 inhibitory and excitatory neurons, since a loss of neurons in the colonic myenteric plexus has
38
39 not been detected (38). In particular, the impairment of cholinergic neurotransmission appears
40
41 to play a significant role in the onset of colonic dysmotility during PD. However, the roles
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43 played by enteric nitrergic and dopaminergic neurotransmission in colonic dysmotility
44
45 associated with PD remains scarcely understood and deserve further investigations.
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51 4. Overall conclusions and future directions

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53 Human studies and evidence from pre-clinical models suggest that PD is associated with
54
55 significant functional, neurochemical and molecular alterations throughout the intestinal tract.
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3 Studies in animal models support the view that intestinal motor abnormalities are associated
4 with changes in the neurochemical coding of myenteric neurons, both in the small and large
5 bowel. Most of functional pre-clinical findings are consistent with bowel dysfunctions
6 observed in patients with PD, even though current morphological and molecular data on both
7 inhibitory and excitatory pathways regulating enteric motility are fragmentary and often
8 conflicting. In addition, a detailed characterization of abnormalities of bowel motor pathways
9 occurring in PD is lacking. It remains also unclear whether the occurrence of rearrangements
10 in the enteric neuronal network and intestinal dysfunctions are direct consequences of the
11 central neurodegeneration or could represent the starting point of the disease. In this respect, a
12 crucial role seems to be played by the neuro-immune brain-gut axis, particularly for DMV,
13 that provides parasympathetic innervation to the majority of GI tract and is regarded as one of
14 the two CNS sites displaying the earliest pathological involvement in PD (44). Indeed, the
15 functional and neurochemical changes affecting the ENS, following central dopaminergic
16 denervation, have been shown to directly result from alterations occurring in the DMV, which
17 is modulated by brainstem dopaminergic circuitries and represents a prominent target of
18 neurodegenerative PD-related processes (45, 46). In support of this view, a reduced
19 expression of ChAT in the DMV of rats with 6-OHDA-induced PD and a reduced digestive
20 motor activity have been reported. In addition, when animals were subjected to vagotomy, the
21 digestive dysmotility improved, thus suggesting that gut dysfunctions in PD could depend on
22 the impairment of vagal brain-gut axis (47).

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47 The vagus nerve is referred also as the “cholinergic anti-inflammatory pathway”, since it
48 seems to exert tonic anti-inflammatory actions, while vagotomy confers an increased
49 susceptibility to the development of inflammatory motility disturbances (48). In the setting of
50 PD, two recent pioneering studies, performed both in colonic biopsies from PD patients and
51 colonic tissues from 6-OHDA-induced PD in rats, have shown an increase in pro-
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3 inflammatory cytokine levels and enteric glial activation (24, 40). In addition, the pre-clinical
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5 study demonstrated also an increase in colonic tissue oxidation, an increment of neutrophil
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7 and mast cell density, and a polarization of peritoneal macrophages towards a pro-
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9 inflammatory phenotype (40). These results suggest the occurrence of an inflammatory
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11 condition in the gut during PD, which could result from an impairment in the vagal brain-gut
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13 axis, and might contribute to generate or worsen enteric dysfunctions related to central
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15 neurodegeneration (24). Since the extrinsic vagal impairment is expected to affect the overall
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17 bowel functions, the development of tissue inflammatory response should affect the whole
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19 intestinal tract. On the other hand, as in the CNS, changes in the ENS of PD patients or
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21 animal models might be restricted to functionally specific and limited bowel regions or
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23 ganglia, and this outcome might increase the likelihood of under- or over-estimating the
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25 actual gut alterations. However, current state of the art does not allow to accept or rule out any
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27 of the above hypotheses, and therefore this issue remains open to future investigations aimed
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29 at evaluating the molecular, morphological and functional changes occurring in different
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31 intestinal regions, at different time points, in different experimental models and PD patients.
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36 In conclusion, current knowledge encourages research efforts dedicated to clarify the role
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38 played by neuro-immune brain-gut axis in the pathogenesis of gut motor dysfunctions in PD.
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40 In particular, it appears of primary relevance to investigate whether gut neuro-inflammation is
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42 a consequence of the impairment of vagal cholinergic anti-inflammatory pathway, and
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44 whether such a condition could be responsible for the alterations of gut motor pathways
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46 occurring in the presence of central dopaminergic neurodegeneration.
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26 CP, MF and LA wrote the first draft of the manuscript. CP prepared the figures. EB, RC, VB,
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28 NB, WdJ and CB revised the manuscript
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Table 1. Functional, neurochemical and molecular alterations associated with bowel disorders in PD patients

<i>Region</i>	<i>Disorder</i>	<i>Observations</i>	<i>Ref.</i>
Small bowel	<i>Dysmotility</i>	✓ α -syn accumulation in mucosal submucosal nerve fibres and myenteric ganglia of duodenum and ileum	[10]
		✓ No alterations of nitrergic, VIPergic, dopaminergic and adrenergic neuron density in myenteric neurons	[11] [12]
		✓ Small intestine bacteria overgrowth	[16]
Large bowel	<i>Constipation</i> <i>Anorectal dysfunction</i>	✓ Reduction of dopamine levels in submucosal and myenteric plexus	[11]
		✓ No alterations of neuronal density in submucosal and myenteric plexus	[18] [19]
		✓ No alterations of nitrergic, VIPergic, dopaminergic and adrenergic neuron density in myenteric neurons	[24] [25]
		✓ Aggregated α -syn in cholinergic and substance P-containing neurons in submucosal and mucosal biopsies	[49] [50]
		✓ Increase in cytokine levels (TNF, IL-1 β , IL-6, IFN- γ) and enteric glial activation (GFAP and Sox-10)	

Abbreviations: α -syn: α -synuclein; GFAP: glial fibrillary acidic protein; IL-1 β : interleukin 1 beta; IL-6: interleukin-6; IFN- γ : interferon gamma; Sox-10: sex determining region Y (SRY) box containing gene 10; TNF: tumor necrosis factor; VIP: vasoactive intestinal peptide.

Table 2. Main features of experimental models employed in the evaluation of bowel alterations associated with central dopaminergic neurodegeneration

<i>Experimental models</i>	<i>Functional, neurochemical and molecular characteristics</i>		<i>Ref.</i>
<i>Neurodegeneration induced by peripheral toxin administration</i>	MPTP	<ul style="list-style-type: none"> ✓ Blockade of mitochondrial complex I in SNpc ✓ Loss of dopaminergic neurons in the SNpc ✓ Intraperitoneal administration not associated with formation of LB-like cytoplasmic inclusions ✓ Administration via osmotic minipumps followed by formation of LB-like cytoplasmic inclusions containing ubiquitin and α-syn 	[51]
	Rotenone	<ul style="list-style-type: none"> ✓ Different routes of administration (oral, intraperitoneal, intravenous and systemic by osmotic pump) ✓ Degeneration of nigrostriatal dopaminergic neurons ✓ Blockade of mitochondrial electron transport chain ✓ Increase in oxidative stress ✓ Inhibition of proteasome activity ✓ LB-like cytoplasmic inclusions containing ubiquitin and α-syn 	[51] [52] [53]
<i>Neurodegeneration induced by intranigral toxin administration</i>	6-OHDA	<ul style="list-style-type: none"> ✓ Bilateral or unilateral intranigral administration ✓ Massive anterograde degeneration of the nigrostriatal pathway (nigral cell loss and striatal dopamine depletion) ✓ Increase in hydrogen peroxide formation ✓ Inhibition of mitochondrial complex I in SNpc 	[54]

Abbreviations: α -syn: α -synuclein; LB: Lewy bodies; MPTP: 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; 6-

OHDA: 6-hydroxydopamine; SNpc: substantia nigra pars compacta

Table 3. Summary of functional, neurochemical and molecular evidence in the small and large bowel associated with central dopaminergic neurodegeneration

<i>Experimental models (Species)</i>	<i>Functional evidence</i>	<i>Neurochemical evidence</i>	<i>Molecular evidence</i>	<i>Ref.</i>
Small bowel				
<i>MPTP (mouse)</i>	Reduced myoelectric activity Prolonged irregular contractions No alterations of overall transit	↓ TH ↓ DAT No alterations of VIP No alterations of NET	↓ TH protein expression ↓ DAT protein expression ↓ DA concentration	[29] [31] [32]
<i>Unilateral intranigral injection of 6-OHDA (rat)</i>	Reduced transit	↓ nNOS (distal ileum) ↓ nNOS (duodenum) ↑ VIP No alterations of ChAT (distal ileum) ChAT (duodenum) ↓ No alterations of HuC/D expression	↑ TH protein expression ↑ DAT protein expression	[31] [35] [36]
Large bowel				
<i>MPTP (mouse)</i>	Delay of transit Impaired inhibitory control of motility	n.a.	↓ TH protein expression	[29] [31] [32]
<i>Rotenone (mouse)</i>	Delay of transit	No alterations of VIP No alterations of NOS No alterations of TH	n.a.	[33]
<i>Unilateral intranigral injection of 6-OHDA (rat)</i>	Decrease in stool frequency Delayed colonic transit Reduced efficiency of peristaltic reflex Alterations of longitudinal muscle contraction Decrease in electrically evoked cholinergic contractions	↑ VIP ↑ TH ↓ nNOS ↓ D ₂ dopaminergic receptors No alteration of ChAT (proximal colon) ↓ ChAT (distal colon)	↑ TH protein expression ↑ DAT protein expression ↓ nNOS protein expression ↓ nNOS mRNA expression ↓ ChAT immunopositivity ↓ Acetylcholine release from enteric neurons ↑ M2 and M3 muscarinic receptor expression	[29] [35] [37] [38] [41]

Abbreviations: ChAT: choline acetyltransferase; DAT: dopamine transporter; DA: dopamine; NET: noradrenaline transporter; nNOS: neuronal nitric oxide synthase; TH: tyrosine hydroxylase; VIP: vasoactive intestinal peptide; n.a: data not available.

Figure legends

Figure 1. Schematic illustration of small bowel abnormalities in experimental models of Parkinson's disease (PD). Left panel: central dopaminergic denervation, induced by intranigral toxins injection, is associated with an increase in choline acetyl transferase (ChAT) and nitric oxide (NO) expression in myenteric neurons of duodenum. In the distal ileum, the expression of tyrosine hydroxylase (TH), dopamine transporter (DAT) and vasoactive intestinal peptide (VIP) is increased in both myenteric and submucosal plexus, while the expression of neuronal nitric oxide synthase (nNOS) is decreased in myenteric neurons. The overall small intestinal transit is reduced. Right panel: central dopaminergic denervation induced by peripheral toxins injection is associated with a decrease in DAT and TH expression in both myenteric and submucosal plexus of duodenum and ileum. No changes in small intestinal transit have been reported.

Figure 2. Representative picture showing changes in colonic cholinergic neurotransmission in the presence of central dopaminergic denervation elicited by 6-OHDA. The expression of choline acetyltransferase (ChAT) in myenteric plexus is decreased, as well as acetylcholine release from colonic myenteric nerves, in comparison with normal animals. The expression of muscarinic M₂ and M₃ receptors in the colonic smooth muscle layer is enhanced. The colonic motor activity is impaired.

Figure 1. Small bowel alterations in central and peripheral models of PD

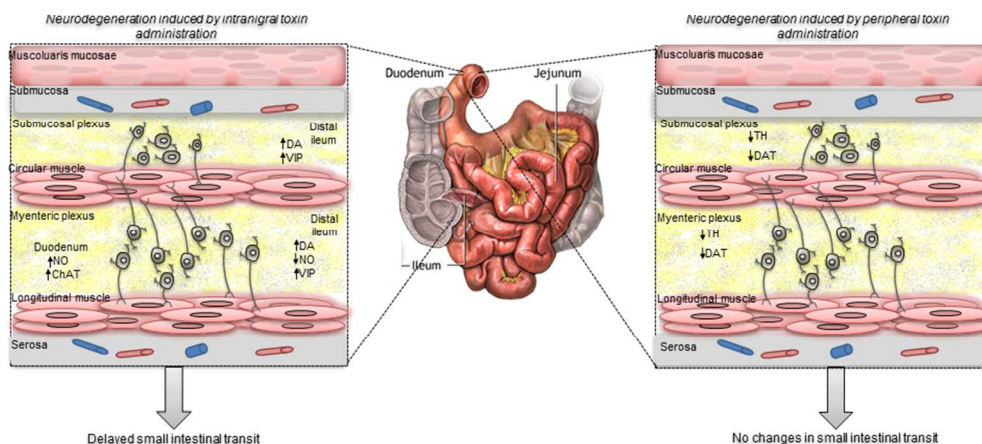


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Figure 2. Alterations of colonic cholinergic neurotransmission in the presence of central dopaminergic denervation induced by intranigral injection of 6-OHDA

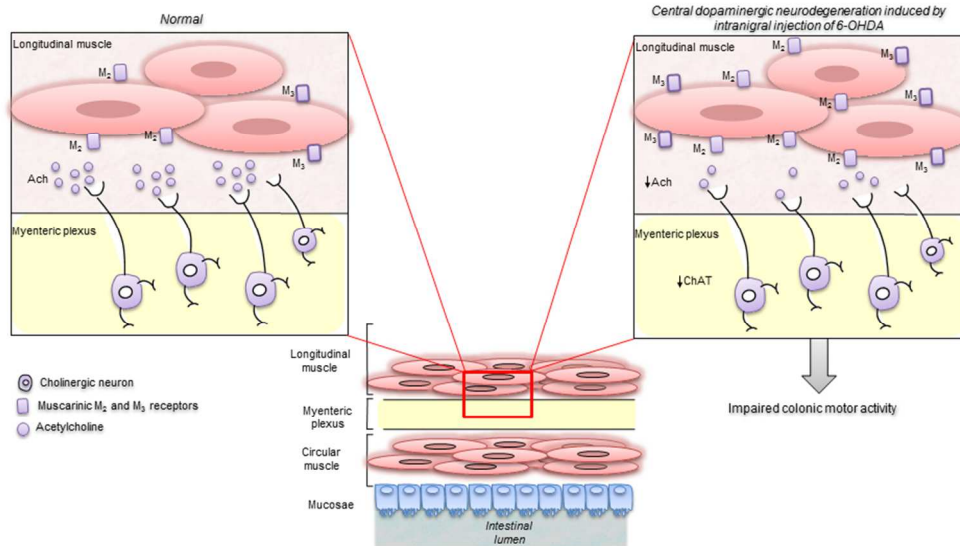


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Key points

- Infrequent bowel movements and overt constipation represent the main intestinal disturbances in patients with PD.
- PD patients are characterized by aggregated α -syn in enteric neurons, intestinal dysbiosis and the occurrence of inflammatory activity in colonic tissues.
- The implementation of preclinical PD models is allowing a characterization of the role played by the enteric cell network in the pathophysiology of bowel disorders associated with PD.
- Studies in animal models support the view that intestinal motor abnormalities are associated with changes in the neurochemical coding of myenteric neurons, both in the small and large bowel.

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