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Paraoxonase-1 activities in individuals with different HDL circulating levels: Implication in reverse cholesterol transport and early vascular damage.

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**Title: Paraoxonase-1 activities in individuals with different HDL circulating levels:  
implication in reverse cholesterol transport and early vascular damage**

**Short title: Paraoxonase 1 activities and different HDL circulating levels**

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## Abstract

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3 BACKGROUND: Epidemiological data showing that high-density lipoprotein cholesterol (HDL-C)  
4 is inversely associated with cardiovascular disease (CVD), have led to the idea that the cholesterol  
5 contained in this lipoprotein may be protective. Against, recent evidence suggests that athero-  
6 protection from HDLs is most likely the result of other biological functions, unrelated to the  
7 cholesterol carried by these lipoproteins. HDL accessory proteins, such as paraoxonase 1 (PON1),  
8 have been suggested to endows HDL with antioxidant and anti-inflammatory proprieties and to  
9 strongly contribute to the athero-protective function of the lipoprotein.  
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13 AIM: To evaluate whether extreme fluctuation in HDL-C levels correlate with PON1 activity.  
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16 METHODS: Levels of PON1-related arylesterase and lactonase were assessed in subjects with  
17 primary hyperalphalipoproteinemia (HAL, HDL-C>90th percentile), hypoalphalipoproteinemia  
18 (HA, HDL-C<10th percentile) and Controls. Cholesterol Efflux Capacity (CEC) through several  
19 pathways and other metabolic parameters and markers of vascular disease were also determined.  
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23 RESULTS: Arylesterase and Lactonase were only slightly increased in HAL compared with HA  
24 subjects ( $p<0.05$ ), but not vs controls. After normalization (arylesterase/HDL-C and  
25 lactonase/HDL-C) of PON1 activity, the trend was reverted, with significantly higher PON1 activity  
26 in HA compared to controls and HAL ( $p<0.001$ ). Both enzymatic activities were positively  
27 associated only with aqueous diffusion CEC ( $r=0.318$ ,  $p<0.05$  and  $r=0.355$ ,  $p<0.05$ , respectively)  
28 and negatively with the presence of plaques ( $p<0.05$ ).  
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32 CONCLUSIONS: We showed that extreme high/low HDL-C levels are not associated with equal  
33 increase/decrease in PON1 activities. This enzyme appears to contribute to the HDL role in reverse  
34 cholesterol transport and anti-atherosclerosis processes. Further investigation is required to  
35 corroborate our findings.  
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**Keywords:** Paraoxonase 1, arylesterase and lactonase activity, hyperalphalipoproteinemia,

1 hypoalphalipoproteinemia, Cholesterol Efflux Capacity, markers of subclinical vascular disease  
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## Introduction

1 Epidemiological studies consistently support the concept of high-density lipoprotein cholesterol  
2 (HDL-C) as a strong inverse predictor of cardiovascular risk [1,2]. However, genetic  
3 polymorphisms influencing HDL-C levels do not consistently associate with CVD risk [3];  
4 moreover, interventional studies with HDL-C raising therapies generated inconsistent clinical  
5 outcomes [4,5]. Collectively, these findings suggest that the sole cholesterol content of HDL  
6 particles does not fully capture the HDL-related atheroprotective functions. Indeed, the  
7 functionality of HDL also stem from their capacity to exert anti-inflammatory and antioxidant  
8 activities. This well-recognized pleiotropic nature of HDL mostly relates to its major non-lipid  
9 constituents, i.e. apolipoprotein A-1 (Apo A1), apo A-2, apo E, and other accessory proteins, *in primis*  
10 Paraoxonase 1 (PON1) [6–11].

11 Paraoxonase-1 (PON1) is an esterase/lactonase enzyme almost exclusively associated with  
12 circulating HDL [6]. Despite intense efforts in understanding PON1 role, its catalytic mechanism  
13 and physiological significance is still a matter of debate. Endogen lipophilic lactones, such as those  
14 resulting from fatty acid oxidation (e.g. 5,6-dihydroxy-trienoic acid 1,5-lactone) or from the  
15 pathway of homocysteine catabolism (e.g. acyl-homoserine lactones) are the most likely  
16 physiological substrates of PON1 catalysis [12,13]. Through this activity, PON1 appears to  
17 contribute in protecting HDL, low density lipoprotein (LDL), macrophages, and endothelium from  
18 oxidative and inflammatory challenges, and to stimulate cholesterol efflux from macrophages  
19 [6,11,14,15]. PON1 can also exert arylesterase and paraoxonase activities on synthetic chemicals  
20 [12,16], the former being the most measured activity in epidemiological studies; indeed, it is  
21 minimally influenced by some prevalent PON1 genetic polymorphisms, thus disclosing low inter-  
22 individual variability [16–18]. The putative role of PON1 in promoting HDL biological functions  
23 might account for the epidemiological evidences linking low arylesterase or lactonase activity with  
24 high cardiovascular and neurodegenerative disease risk [19–24].

Hyperalphalipoproteinemia (HAL) is a heterogeneous syndrome characterized by very-high HDL-C (over the 90<sup>th</sup> percentile of the general population); it is caused by different diseases, organ malfunctions and genetic factors. It is not clear whether subjects bearing this condition also disclose low CV-risk [25,26]. In a recent report it has been shown that HAL patients have sub-clinical vascular damage and cholesterol efflux capacity comparable to normolipemic controls [27]. To test the hypothesis that in HAL the absence of putative CVD risk reduction may be at least partially ascribed to inadequate improvement in PON1 function, we compared arylesterase and lactonase plasma activity in three groups of individuals: a) subjects with primary HAL; b) subjects with hypoalphalipoproteinemia (HA: HDL-C <10th percentile); c) subjects with normal levels of HDL-C (Controls). In addition, taking into account the paucity of human studies on PON1-capacity to promote cholesterol efflux from macrophages, we explored the possible association between PON1 activities, HDL-promoted cholesterol efflux capacity (CEC), and sub-clinical atherosclerotic indexes.

## Patients and Methods

### Subjects

The study was conducted on three groups of individuals:

1. HAL group: Twenty subjects (18 females and 2 males; mean age: 52 years) with HDL-C  $\geq 85$  mg/dL for women and  $\geq 75$  mg/dL for men (90th percentile of the local reference population), in at least two consecutive serum samples. The previously reported genetic characterization of HAL subjects highlighted a high prevalence of a common polymorphism on CETP gene (SNP p.Val422Ile) [27];
2. HA group: Twenty subjects (11 females and 9 males; mean age: 49 years) with HDL-C  $< 35$  mg/dL for men and women (10th percentile of the local reference population) in at least two consecutive serum samples.
3. Control group: Twenty subjects (18 females and 2 males; mean age: 51 years) with HDL-C within normal range ( $> 35$  mg/dl AND  $< 75$  mg/dl in men OR  $< 85$  mg/dl in women).

Participants were enrolled among subjects attending the metabolic outpatient clinic of Sant'Anna University Hospital (Ferrara, Italy). All were in good health and none had liver or renal function test abnormalities. Hypothyroidism, pregnancy, alcohol consumption  $> 10$ g daily, active treatment with hormones or lipid-modifying drugs were considered exclusion criteria. Other details on inclusion/exclusion criteria have been reported previously [27].

Patients and controls were clinically evaluated by interview and physical examination; blood pressure, and anthropometric parameters were also measured. This study conforms to The Code of Ethics of the World Medical Association (Declaration of Helsinki) and was conducted accordingly to Good Clinical Practice guidelines. It was approved by the Local Ethics Committee; written informed consent was obtained from each patient and no personal information was available to Authors (blinding).

### Serum sampling and biochemical assays

Venous blood samples from patients were collected after overnight fasting and serum stored at -80°C until analyzed.

Total and unesterified cholesterol (TC and UC), HDL-C, triglycerides, and glucose were assayed by standard enzymatic-colorimetric methods; LDL-C was calculated according to the Friedewald formula. Apo A-1 and Apo B were determined by immunoturbidimetry.

Both arylesterase and lactonase assays were performed by UV-VIS spectrophotometric assays in a 96-well plate format by using a Tecan Infinite M200 microplate reader (Tecan group Ltd, Switzerland).

Arylesterase activity was assessed by using 1 mmol/L phenylacetate, in 9 mmol/L Tris-HCl (0.9 mmol/L CaCl<sub>2</sub>, pH 8) [22]. A molar extinction coefficient of  $1.3 \times 10^3 \text{ L}^{-1} \text{ mol}^{-1} \text{ cm}^{-1}$  was used for calculation of enzymatic activity, expressed in kilo unit per liter. One unit of arylesterase activity accounts for 1  $\mu\text{mol}$  of phenol produced in a minute under the conditions of the assay. Intra-assay CV was 3.8% whereas inter-assay CV was 9.7%.

PON1 lactonase activity was measured using gamma-thiobutyrolactone (TBL) as substrate and Ellman's procedure was used to spectrophotometrically monitor (412 nm) the accumulation of free sulfhydryl groups via coupling with 5,5-dithiobis(2-nitrobenzoic acid) (DTNB) [28]. The reaction was run in the working buffer (50 mmol/L Tris, 1 mmol/L CaCl<sub>2</sub>, 50 mmol/L NaCl, pH=8), 50 mmol/L DTNB and 10.5 mmol/L TBL. A molar extinction coefficient of  $13.6 \times 10^3 \text{ L}^{-1} \text{ mol}^{-1} \text{ cm}^{-1}$  was used for the calculation of enzyme activity, expressed in unit per liter. The intra-assay CV was 6.1 % whereas the inter-assay CV was 9.8 %.

### *Serum HDL cholesterol efflux capacity (CEC)*

HDL CEC results analyzed in the present work belong to a previous published study [26]. Five cholesterol efflux pathways were evaluated by using specific cell models: total CEC, aqueous diffusion (AD)-CEC, ATP-binding cassette A1 (ABCA1)-mediated CEC; ATP-binding cassette G1 (ABCG1)-mediated CEC and scavenger receptor class B (SR-BI)-mediated CEC [29]; the methods



were already been described [29]. Briefly, in all determinations cells were labeled with [1,2-<sup>3</sup>H]-cholesterol and exposed to HDL, isolated from serum by polyethilen glycole [30]. Serum HDL CEC was expressed as a percentage of the radioactivity released to the medium in 4 h (6 h for ABCG1-CEC) after over the total radioactivity incorporated by cells. Serum samples were determined in triplicate.

### *Vascular Assessment*

The determination of the following vascular parameters was performed in Controls and HAL subjects:

#### *Ultrasonography Assessment*

Vascular ultrasonography was performed by a single trained operator, blind to the patient general characteristics. Mean cIMT was measured in triplicate bilaterally at the far wall of the distal common carotid artery (10 mm before bifurcation) in a location not containing plaques. **Subclinical carotid atherosclerosis and sub-clinical femoral atherosclerosis were assessed.**

#### *Ankle-brachial index (ABI)*

The ABI of each lower extremity, measured during the clinical examination with a handheld Doppler stethoscope according a standardized protocol, was calculated by dividing the highest value of systolic blood pressure (SBP) of the posterior tibial or dorsalis pedis arteries by the highest SBP measured in both humeral arteries. A patient was considered to have PAD when the ABI was < 0.9 and arterial calcification with an ABI  $\geq$  1.4.

#### *Flow-mediated vasodilation of the brachial artery*

The measures were performed in a quiet room at a controlled temperature (24 °C), in resting subjects. Brachial artery was scanned in longitudinal section 2 cm above the antecubital fossa with B-mode ultrasonography images. The brachial artery diameters were calculated as an average of 3 consecutive measurements of the distance between the anterior wall and the posterior wall intima-lumen interface at end-diastole both before and 45 s after the deflation of a sphygmomanometer cuff, placed around the arm distal to the imaged artery segment, 60 mmHg above individual systolic

pressure for 5 min. Flow-mediated vasodilation (FMD) of the brachial artery was calculated as the percent change in arterial diameter compared with baseline resting diameter.

#### *Statistical analysis*

Sample size was determined as previously described [27]. Novel variables were generated by normalizing lactonase or arylesterase for HDL-C or Apo A1. These and the other continuous variables were first analyzed for normal distribution using Kolmogorov-Smirnov and Shapiro-Wilk tests. Variables not normally distributed were log transformed before entering statistical analyses. Group comparisons were performed using ANOVA (Sidak post-hoc for pairwise comparisons) for continuous variables and chi-square test for categorical variables. When it was possible, analysis of covariance (ANCOVA) was performed to test whether the differences revealed at univariate analysis were independent of potential confounding factors. Pearson's correlation coefficient was used to evaluate the possible association between PON1 activities and parameters of cholesterol efflux, vascular reactivity and damage. This test was followed by multiple regression analysis in order to check the independence of the observed simple associations.

The probability of having arterial plaques (Odds Ratio, O.R.; 95% confidence interval, C.I.—95%) in subjects with high arylesterase or high Apo A1 was calculated by multivariate logistic regression analysis. High/low arylesterase and ApoA1 serum levels were also combined into 3 groups in order to evaluate the risk of arterial plaques associated with the combination of these markers.

A two-tailed  $p < 0.05$  was considered statistically significant.

Statistical analysis was performed using SPSS 22.0 software (SPSS, Chicago, IL).

## Results

### 1. *Clinical characteristics, lipid profile and vascular parameters in the sample groups*

The main demographic, clinical and laboratory characteristics of the subjects enrolled in the study are summarized in **Supplemental Table 1**. HAL, HA, and Controls did not significantly differ by prevalence of obesity and smoking status. Of note, HAL and Controls, but not HA, were age- and gender-matched; HA differed from the other two groups as regards gender prevalence (Females: 5% vs. 90%), but not mean age. Conversely, HA group presented higher prevalence hypertension.

Regarding lipid profile, patients with HAL had the lowest level of triglyceride (ANOVA,  $p=0.026$ ), and the highest levels total cholesterol (ANOVA,  $p<0.001$ ) and LDL-C (ANOVA,  $p=0.018$ ), while neither Apo B nor non-HDL-C significantly differ between the groups. The ratio unesterified/total cholesterol, although showing the highest values in HAL subjects (ANOVA,  $p<0.001$ ), was within the reference range for all the three groups ( $<0.30$ ) [31]. ApoA-1 (HA  $99\pm 30$  mg/dL, ANOVA,  $p<0.001$ ), and HDL-C (ANOVA,  $p<0.001$ ) progressively increased from HA, to Controls to HAL, according to selection criteria.

As already shown and previously described [27], no differences were observed as regards plaque number (both in carotid and femoral arteries), ABI, cIMT, and FMD% between Controls and HAL subjects.

### 2. *PON1 activities in Control, HAL and HA group*

As shown in **Fig.1A**, the trend of **arylesterase** PON1 activities resembled that observed for HDL-C, but the differences were much less evident, with only HAL vs HA difference being significant ( $p<0.05$  for both comparisons). Considering the extent of changes in HAL group compared to Controls, lactonase and arylesterase were increased by 15% and 17%, respectively (HDL-C increase was around 43%), while the decrease in HA was equal to 6% and 16% (HDL-C decrease was 57%). This lack of parallelism between PON1 activity and HDL-C became more

evident after normalization of arylesterase/lactonase for HDL-C concentration (**Fig. 1 B**). Indeed, upon this mathematical transformation, the trend was reverted and the differences between groups (Controls vs. HAL, Controls vs. HA, and HA vs. HAL) became highly significant ( $p < 0.001$  for all). A similar result, with less marked differences, was observed after normalization of PON1 activities for Apo A-1 concentration (ANOVA  $p < 0.001$ ) (**Fig.1 C**).

### 3. *PON1 activities and vascular parameters*

None of the markers of early vascular damage evaluated (i.e. ABI, cIMT, and FMD) was significantly correlated with either arylesterase or lactonase activity (data not shown). On the contrary, the presence of established vascular lesion such as an arterial plaque, was associated with a significantly ( $p < 0.05$ ) lower level of arylesterase (but not of lactonase) (**Fig. 2**). Of interest, no significant association was observed between levels of Apo A-1 or HDL-C and the presence of plaques (data not shown), despite their well-recognized athero-protective function.

APO A-1 is widely suggested to be the most important determinant of HDL function as well as a synergic and essential co-factor of PON1 [32,33]. Consistently with this proposed intimal connection between PON1 and this apolipoprotein, we found that they were strongly related to each other (arylesterase vs Apo A-1:  $r = 0.45$ ,  $p = 0.001$ ). To investigate whether this functional synergy could influence the occurrence of plaques, we compared the prevalence of these lesions with four possible combinations of high/low arylesterase and high/low (median=cut off) APO A-1 levels: 1) Low arylesterase/Low Apo A-1 (n=11) 2) High arylesterase/Low Apo A-1 (n=9); 3) Low arylesterase/High Apo A-1 (n=9) 4) High arylesterase/High Apo A-1 (n=11). As disclosed in **Fig. 3**, the group with low levels of both parameters presented by far the highest frequency (64%) of individuals with plaques; at the opposite, the group with High arylesterase/High Apo A-1 included the lowest (18%). To check if these descriptive findings have statistical relevance, we performed a binary logistic regression, taking Low arylesterase/Low Apo A1 as reference group (**Fig. 4**). This analysis showed that high serum levels of either arylesterase or APO A-1 were not associated with a

1 significant reduction in odds of having artery plaques (O.R: 0.21, 95% C.I.: 0.03-1.20; O.R: 0.14,  
2 95% C.I.: 0.02-1.06, respectively). From the combination of the two dichotomous variables it  
3 emerged that only the group with high levels of both arylesterase and Apo A-1 showed a lower  
4 likelihood of having plaques compared to reference group (O.R: 0.38, 95% C.I.: 0.19-0.94: p<0.01).  
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#### 9 4. PON1 *activities* and cholesterol efflux capacity

10 As suggested by a wealth of in vitro evidence, the beneficial function of PON1 may be exerted by  
11 ameliorating the capacity of HDL to promote cholesterol efflux from cells [34,35]. To check this  
12 possibility, we evaluated the possible association between PON1 activities and the cholesterol  
13 efflux pathways measured in this study. We found that Arylesterase **activity** was positively  
14 correlated with AD-efflux, while lactonase was associated with both this pathway and total CEC  
15 (**Table 1**). As expected, Apo A-1 emerged as a major determinant of all efflux pathways, and the  
16 aforementioned relationships observed for PON1 activities disappeared after adjusting for the levels  
17 of this apolipoprotein (**supplemental Table 2 and 3**).  
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## Discussion

1 To the best of our knowledge, the present is the first study showing that HDL of HAL individuals  
2 are relatively poorer in PON1 activities compared to subjects with normal or very-low HDL-C  
3 levels. In turn, this low PON-1 activity may reflect in a minor efficiency of the lipoprotein,  
4 mitigating the potential atheroprotective effect due to the constitutively high concentration of HDL-  
5 C characterizing these individuals.  
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13 In line with our results, growing evidence suggests that HDL function may be more relevant than  
14 HDL-C concentration in protecting against CVD [36,37]. The antioxidant PON1 is widely  
15 suggested to be one of the components of HDL proteome mostly contributing to the lipoprotein  
16 atheroprotective function [34,38]. In line with these considerations, in vivo (mice over-expressing  
17 PON1) [15] and ex vivo (oxLDL pretreated with PON1) [39] studies suggest that PON1 stimulates  
18 cholesterol efflux from macrophages. HDL-mediated cholesterol efflux represents the first and the  
19 rate-limiting step of this complex and multi-step process. This recover of excess cholesterol from  
20 cells (including foam cells) can occur via passive/facilitated (AD-CEC and SR-BI-mediated) and  
21 active pathways (mediated by membrane proteins such as ABCA1 and ABCG1). A previous  
22 investigation conducted on the same Controls and HAL groups, found that all cholesterol efflux  
23 pathways were comparable or only slightly increased in the latter group [27]. Notably, as observed  
24 for PON1, normalizing for HDL-C reversed the trend, and all CEC pathways increased significantly  
25 in Controls compared to HAL. Consistent findings from in vitro studies using recombinant or  
26 plasma purified PON1 [11,34,35], or the overexpressed protein in mice [15] showed that PON1  
27 plays some role in CEC. Encouraged by these intriguing findings, we explored for the first time the  
28 possible association between serum PON1 activity and all CEC pathways. In apparent contrast with  
29 the proofs in support of an interaction of PON1 with ABCA1 [11,39], we did not find any  
30 association between cholesterol efflux mediated by neither this transporter nor by ABCG1 (mainly  
31 involved in cholesterol efflux to mature HDL [40]) and PON1 activities. However, lactonase (i.e.  
32 the activity putatively responsible of generating lysophosphatidylcholine) was weakly associated  
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with both aqueous and SR-BI CEC pathways. As disclosed by multivariate analysis, this association was chiefly influenced by Apo A-1, most likely because of the strong and well known [35] relationship of this apolipoprotein with PON1 function and CEC of HDL. Overall, this result is consistent with the hypothesis that the contribution of PON1 in CEC may depend on its intimate functional and physical interaction with Apo A-1 on the HDL suggested by the solved crystal structure of the enzyme [33].

Consistently with the mutual interaction between PON1 and Apo A-1 [33,41,42], we found that high levels of arylesterase and Apo A-1 are associated with less arterial plaques. This result is in partial agreement with some clinical/epidemiological studies, reporting an association between sub-clinical atherosclerosis and PON1 [43,44]. PON1 has been widely proposed to act, most likely in coordination with phospholipase A2 and vitamin E, as an antioxidant shield of Apo A-1 [32,33]. This protective action is important for preserving the structural and functional integrity of APO A-1. Indeed, it has been observed that oxidative modification primed by myeloperoxidase and other pro-oxidant agents seems to provoke an impairment of APO A-1 properties, affecting efflux capacity and anti-inflammatory function of HDL, thus favoring atherosclerosis development [41,45]. Consistently, Shao et al. found that patients with coronary artery disease have increased levels of oxidized APO A-1 [46].

Differently from us, other studies reported a significant, although generally weak, inverse correlation between PON1 activities and some early marker of artery diseases, such as cIMT [47–50]. However, these discrepancies might be due to differences in the design of the study, sample composition, as well as in the analytical procedures employed to assay PON1 activity. In particular, the exclusive assessment of paraoxonase activity, measured in two of these studies, is considered not suitable for epidemiological/clinical studies [17]. Indeed, this activity is influenced by PON1 polymorphisms (in particular 192 Q/R) much more than arylesterase and lactonase [17]. This is an important source of variability in PON1 data, which somewhat limits inter-laboratory comparisons.

The present study presents some caveats. First of all, our cross-sectional study design, as opposed to

longitudinal approach, precluded our ability to establish any cause/effect relationship. **Second**, the small **size** of our sample and the lack of gender-match between the three groups (**only HAL and Controls were sex-matched as previously reported [27]**) may affect the reliability of our conclusions. **Third**, since PON1 physically interacts with HDL particles, the measurement of enzyme activities on isolated HDL instead of whole serum sample, would be more valuable and informative. However, it is well established that, at least in healthy subjects, almost the totality of PON-1 is associated with HDL [51]; thus, any alteration in its serum activity should mirror that of HDL-associated enzyme.

A fourth possible limitation of our study is related to the lack of a characterization of the HDL phenotype of our subjects, especially in terms of CETP and LCAT activity. However, a previous genetic analysis of our HAL subjects [27] showed a high prevalence of a CETP polymorphism associated to reduced CETP mass and activity [52]. In addition, the ratios unesterified/total cholesterol that we found in serum seem to suggest a normal cholesterol esterification process, reflex of physiological LCAT activity/mass [31].

**Fifth**, the method for lactonase activity assessment, though presenting important advantages over the others, has some limitations. Moreover HDL-associated PON3 elicits lactonase activity [12] and, even if presumably to a low extent (the enzyme and related activity, is two orders of magnitude less abundant than the other extracellular isoenzyme), it might interfere with the results.

In conclusion, our study showed that arylesterase and lactonase activities of PON1 are not significantly increased in individuals with HAL compared to Controls. This unequal change in PON1 may account for the lack of benefit in terms of CVD risk reduction reported in HAL despite the important increase in their plasma HDL-C. This concept is indirectly supported by our finding of a significant inverse association between PON1 (together with Apo A-1) and the presence of artery plaques. Further studies on larger sample are required to corroborate these preliminary findings.



1 **Conflict of interest**

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3 The authors declared they do not have anything to disclose regarding conflict of interest with  
4  
5 respect to this manuscript.  
6

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13 **Author contributions**

14  
15 Our work is a product of the intellectual environment of the whole team; all members have  
16  
17 contributed in various degrees to its birth.  
18

19  
20 Conception and design of the study - AP, CC.  
21

22  
23 Acquisition of data - GBV, AP, CC, JMS.  
24

25  
26 Data analysis and interpretation - CC, AP, AT.  
27

28  
29 Drafting of the article - CC, AP.  
30

31  
32 Critical revision of the article - GBV, MLM, JMS, EDN, and GZ.  
33

34  
35 All authors read and approved the final manuscript.  
36

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## Legends

1 Fig. 1. Arylesterase and lactonase before (A) and after normalization for HDL-C (B) or Apo A1 (C)  
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3 in CONTROLS, HA and HAL.

4  
5 Fig. 2. Arylesterase and lactonase levels in subjects with or without artery plaques.

6  
7 Fig. 3. Percentage of subjects with arterial plaques in groups with high/low levels (than median  
8 value = 96 KU/L) of serum arylesterase and/or with high/low levels (than median value = 174  
9 mg/dL) of serum Apo A1.

10  
11 Fig. 4. Odds ratio (95% confidence interval) for the presence of arterial plaques in subjects with  
12 high/low levels (than median value = 96 KU/L) of serum arylesterase and/or with high/low levels  
13 (than median value = 174 mg/dL) of serum Apo A1.

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15 Table 1: Correlation coefficients for the potential association of Apo A1, Arylesterase, and  
16 lactonase with serum HDL cholesterol efflux pathways (whole sample, n= 60).

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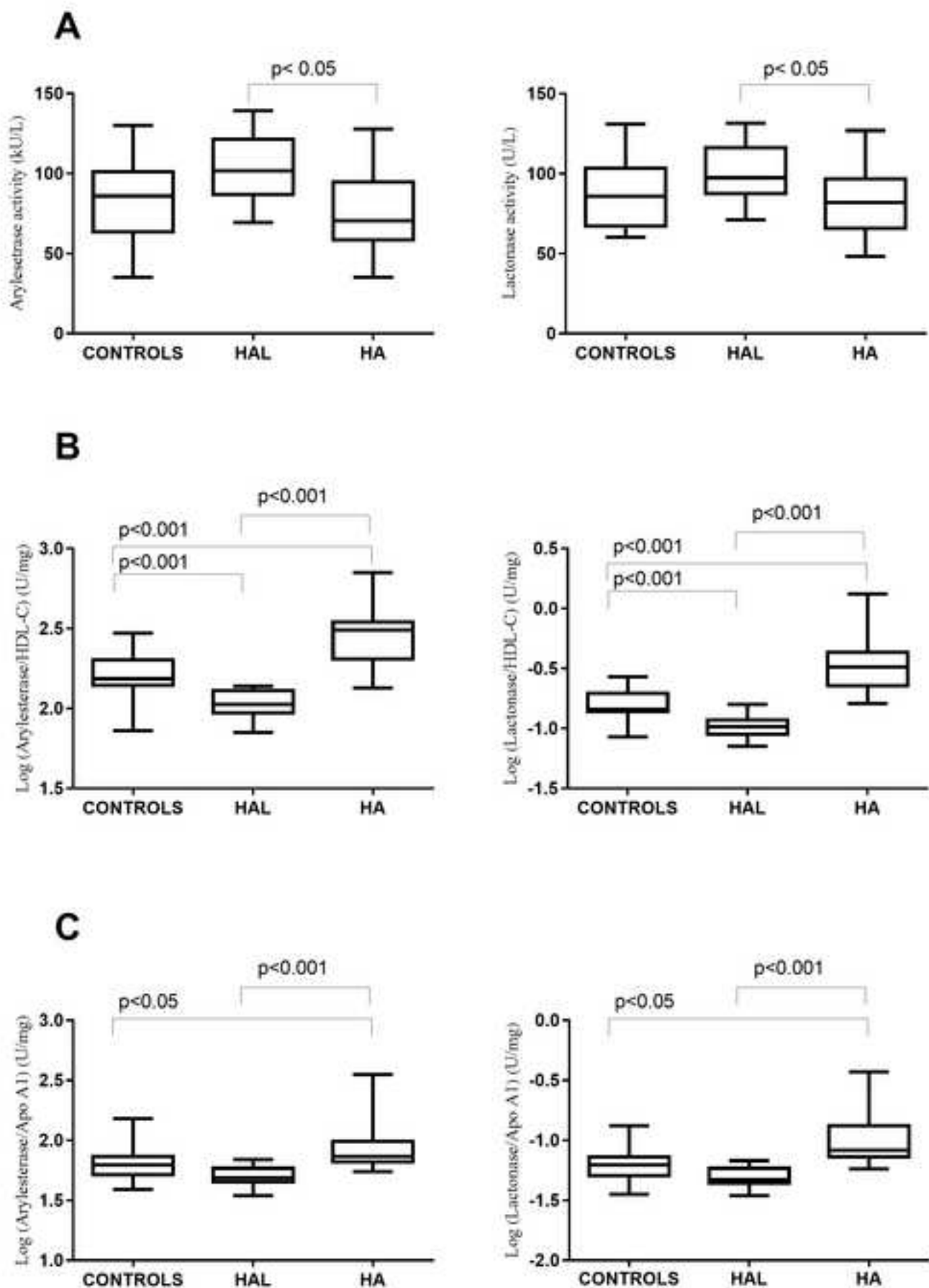
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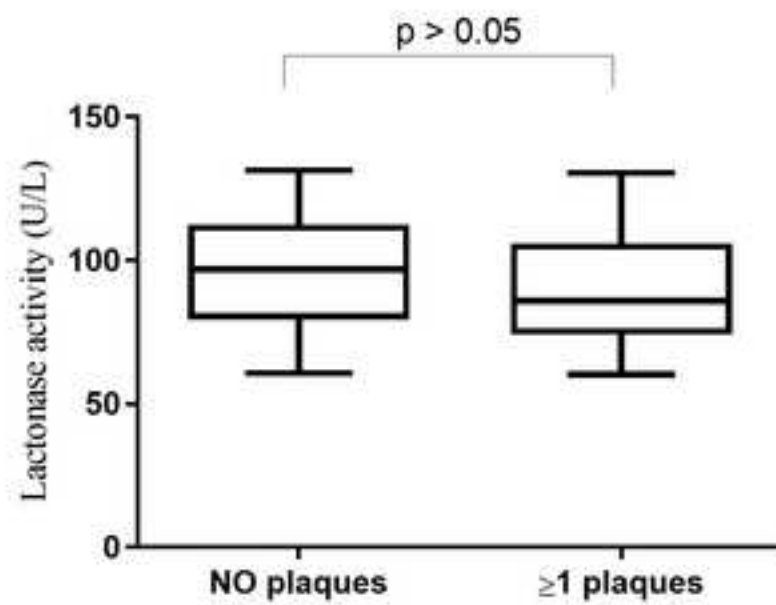
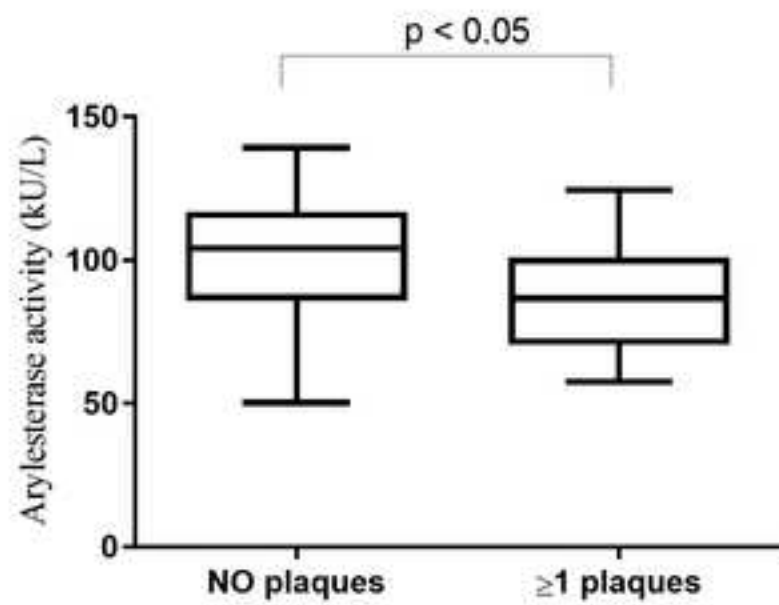
Table 1: Correlation coefficients for the potential association of Apo A1, Arylesterase, and lactonase with serum HDL cholesterol efflux pathways in Controls and HAL subjects

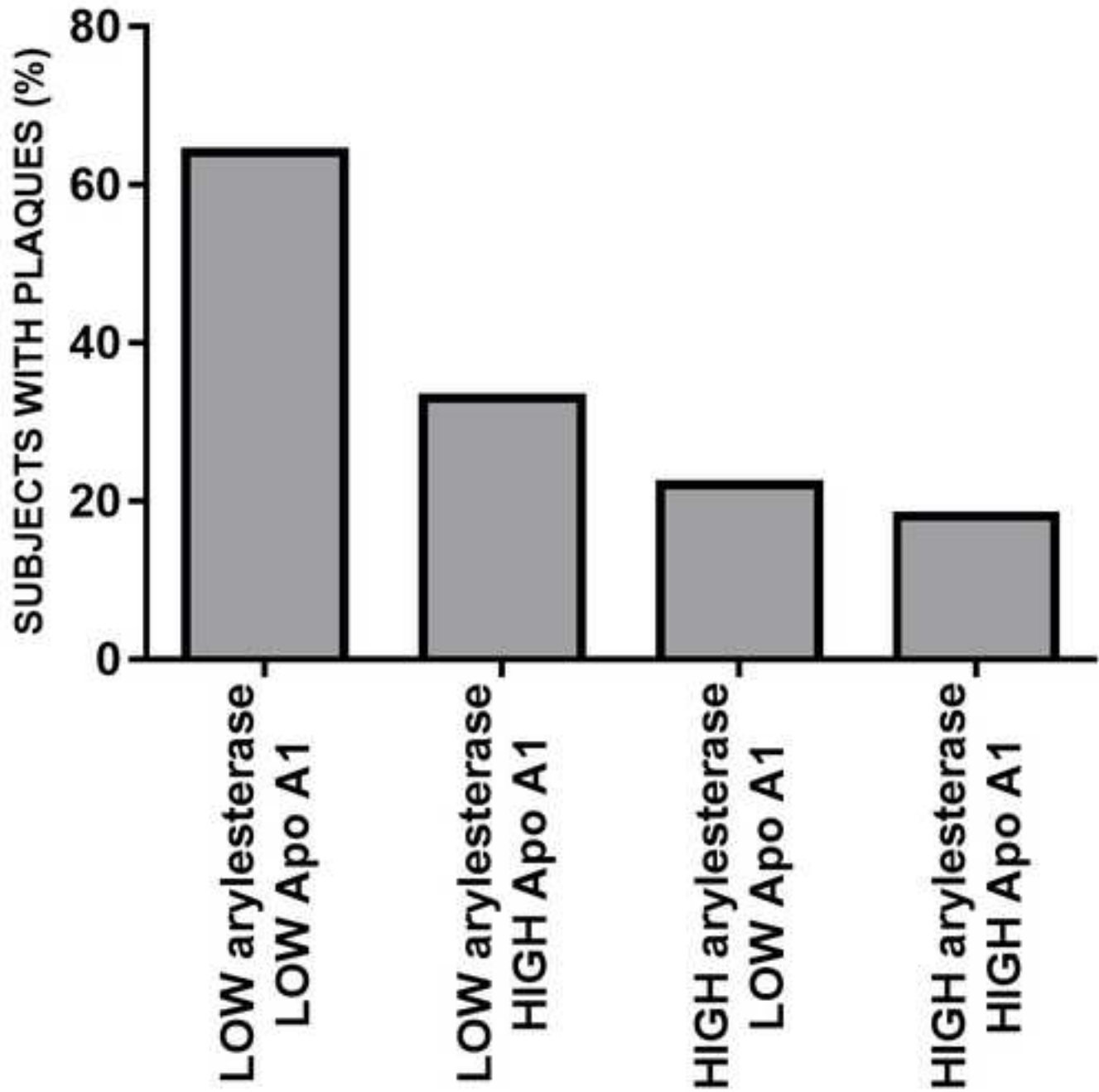
	Controls n=20			HAL n=20		
	Apo A1 (r)	Arylesterase (r)	Lactonase (r)	Apo A1 (r)	Arylesterase (r)	Lactonase (r)
Total efflux/4 h	0.352	0.278	0.178	0.446*	0.327	0.328
AD-efflux/ 4 h	0.398	0.236	0.178	0.431***	0.326	0.319
SR-B1-efflux/4h	0.661**	0.065	0.167	0.618**	0.227	0.103
ABACA1-efflux/4 h	0.252*	-0.318	-0.017	0.178*	0.135	0.186
ABCG1-efflux/6h	0.510*	0.086	-0.017	0.516*	-0.145	0.186

\*p<0.05; \*\*p<0.01;\*\*\*p<0.001

Abbreviations: AD, aqueous diffusion; SR-B1, scavenger receptor class B type 1; ABACA1, ATP-binding cassette A1; ABCG1, ATP-binding cassette A1







Figure(s)

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