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1 **ABSTRACT**

2 **Objective** To evaluate the anesthetic effects and the reliability of three different alfaxalone doses
3 not previously used to induce anesthesia in goldfish.

4 **Study design** Prospective, randomized, clinical study.

5 **Animals** Thirty goldfish undergoing skin scraping, gill exam and stool collection.

6 **Methods** Each fish was transferred to an individual 4 L induction tank. The fish were randomly
7 allocated into three groups (n = 10) in which anesthesia was induced with alfaxalone 6 mg/L, 7
8 mg/L and 9 mg/L. The depth of anesthesia was evaluated by assessing reactivity, activity,
9 maintenance of equilibrium, opercular movement and response to noxious stimuli. Sedation, light
10 anesthesia and surgical anesthesia induction times and recovery time were recorded. The fish length
11 (from snout to fork in caudal fin) was measured. The data were analyzed using ANOVA. Statistical
12 significance was set at $p < 0.05$

13 **Results** All fish achieved surgical anesthesia stage. Goldfish induced with alfaxalone 7 mg/L and 9
14 mg/L showed a mild excitement phase. Sedation induction time of 6 mg/L dose was significantly
15 longer compared to 7 mg/L and 9 mg/L doses. Light anesthesia and surgical anesthesia induction
16 times of 9 mg/L dose were significantly faster compared to 6 mg/L and 7 mg/L doses. No
17 significant differences were recorded in recovery time. Induction and recovery times had no
18 correlation with goldfish size. The cessation of opercular movement was recorded in two fish
19 induced with 7 mg/L and in two induced with 9 mg/L. At 15 days post anesthesia, the fish had a
20 normal physical appearance and no death was observed.

21 **Conclusions and clinical relevance** Alfaxalone is a reliable agent for immersion anesthesia in
22 goldfish. Immersion in water concentration of 6 mg alfaxalone/L provides a smooth induction of
23 anesthesia without side effects. Higher doses shorten the time required for induction, and cause
24 respiratory depression and excitatory movements.

25

26 **Keywords** goldfish, *Carassius Auratus*, fish, immersion anesthesia, alfaxalone.

27 INTRODUCTION

28 Veterinarians always more often treat ichthyic sector. Goldfish (*Carassius auratus*) is the most
29 commonly fish kept as pet. Goldfish is a long-lived species and it may be affected by cutaneous,
30 intestinal, metabolic disorders and neoplasm (O'Hagan & Raidal 2006). Sedation and general
31 anesthesia are required to handling, diagnostic tests and surgical procedures (Sneddon 2012).

32 The key points of fish anesthesia are how to assess the depth of anesthesia, which route to use for
33 administering drugs and what drug to employ. The depth of anesthesia may be evaluated assessing
34 reaction to external stimuli, swimming, gill ventilation rate and reflex response (Sneddon 2012).
35 Furthermore, the depth of anesthesia is related to the anesthetic agent, dose and time exposure
36 (Fleming et al. 2003; West et al. 2007).

37 Immersion anesthesia is more often performed than parenteral anesthesia (West et al. 2007).
38 Sedative and anesthetic agents used for fish are very different from those used for mammals. The
39 most common drugs used to induce general anesthesia in fish are as follow: tricaine (MS-222),
40 benzocaine, isoeugenol and quinaldine (Sneddon 2012). Many agents, which are usually employed
41 for parenteral anesthesia in dogs and cats, are water-soluble and may be administered through the
42 water because fish ventilate the anesthetic agent in solution (Sneddon 2012). Medetomidine-
43 ketamine (Fleming et al. 2003), atipamezole (Williams et al. 2004), propofol (Fleming et al. 2003;
44 GholipourKanani & Ahadizadeh 2013), metomidate (Iversen et al. 2003), and diazepam (Kumlu &
45 Yanar 1999) produced variable results in fish.

46 A formulation composed of a mixture of alfaxalone and alphadolone acetate solubilized in 20%
47 polyethoxylated castor oil (Cremophor-EL) has been previously employed as an intramuscular or an
48 immersion anesthetic agent in fish (Harvey et al. 1988; Peters et al. 2001). A new water-soluble
49 formulation of alfaxalone solubilized in 2-hydroxypropyl-beta cyclodextrin (Ferre et al. 2006) has
50 been employed as an immersion anesthetic agent. In koi carp (*Cyprinus carpio*), immersion in water
51 concentration of 10 mg alfaxalone/L caused fast induction, but the cessation of opercular movement
52 was detected during maintenance (Minter et al. 2014). In oscar fish (*Astronotus ocellatus*),

53 immersion anesthesia with alfaxalone 5 mg/L was reliable for collection of blood samples, despite it
54 significantly reduced the respiratory rate (Bugman et al. 2016). The use of alfaxalone in goldfish
55 was described in two case reports (O'Hagan & Raidal 2006; Fernández-Parra et al. 2017). Induction
56 and maintenance were achieved with alfaxalone, respectively, 10 mg/L and 5 mg/L, but respiratory
57 depression occurred (Fernández-Parra et al. 2017). Moreover, the findings of Bauquier et al. (2013)
58 underscored that administering alfaxalone 5 mg/L induced surgical anesthesia in 5/6 goldfish and
59 light anesthesia in 1/6 goldfish whereas 7.5 mg/L dose induced surgical anesthesia in 6/6 goldfish
60 but caused delayed recovery compared to 5 mg/L dose (Bauquier et al. 2013).

61 The purpose of this study is to evaluate the reliability of three different alfaxalone doses not
62 previously used to induce general anesthesia in goldfish. The present clinical study aims to assess
63 the anesthetic effects of 6 mg, 7 mg and 9 mg alfaxalone/L and to compare the effect of these doses
64 on induction and recovery times.

65

66 **MATERIALS AND METHODS**

67

68 **Animals**

69 The study was performed in accordance with the XXX legislation on animal care (XXX) and XXX
70 law (XXX).

71 Thirty goldfish of unknown gender, one year-old, obtained from the same fish private pond, that
72 underwent skin scraping, gill exam and stool collection, were enrolled. For the acclimation period
73 of 15 days, thirty goldfish were housed indoors together in 130 L tank filled with municipal water
74 constantly aerated with an air stone on a mechanical pump (Haquoss airline3 180 L/h, Haquoss,
75 XXX). Water temperature, pH and nitrates were daily measured and ranged between, respectively,
76 22.4-24°C, 6.9-7.5 and 0.1-0.23 mg/L. The fish were fed with balanced dry fish food given twice a
77 day. They were considered healthy following a visual exam based on the evaluation of equilibrium,
78 swimming, opercular movement and physical appearance.

79

80 **Study design**

81 The fish were withheld for 12 hours before anesthesia. Each fish was transferred to an individual 4
82 L induction tank filled to 75% and aerated with an air stone on a mechanical pump. The water used
83 for trials was the same in which the fish were housed. Two g/L of NaCl was added to each
84 induction tank to help fish osmoregulation.

85 The fish were randomly (simple randomization) divided into three groups ($n = 10$) in which
86 anesthesia was induced, using three different concentrations of alfaxalone (Alfaxan, Dechra, XXX)
87 as follow: 6 mg/L (group G6), 7 mg/L (group G7) and 9 mg/L (group G9).

88 Before adding alfaxalone to the anesthetic tank, the fish was observed for approximately 10
89 minutes and the approach reaction score was evaluated (Table 1). After adding alfaxalone to the
90 induction tank, the assessment of approach reaction, equilibrium and operculum movement scores
91 (Table 1) was performed every minute until loss of equilibrium and reaction to tactile stimulus. The
92 anesthetic stages were evaluated according to criteria outlined in Table 2. When fish reached the
93 anesthetic stage 3, response to noxious stimuli was assessed by the same practitioner that squeezed
94 the caudal fin between two fingers. When response to noxious stimuli score reached 4 (Table 1), the
95 fish was removed from the anesthetic bath, laid down on a gauze dampened with water and
96 photographed. The fish length (from snout to fork in caudal fin) was measured and skin scraping,
97 gill exam and stool collection were performed in two minutes. At the end of these procedures, the
98 fish was transferred to an individual 4 L recovery tank filled to 75% and aerated with an air stone on
99 a mechanical pump. The fish was gently moved with a steel stick in a swimming motion until
100 righting reflex resumed. If opercular movement was not detected, a steady flow of water was
101 directed through the oral cavity and across the gills. The fish was moved to the original 130 L tank
102 after recovery.

103 A daily visual exam based on the evaluation of equilibrium, swimming, opercular movement and
104 physical appearance was performed for 15 days.

105

106 **Data collection**

107 The recorded times (minutes) were defined as follow: sedation induction time (the time from adding
108 alfaxalone to water to the time of sedation stage), light anesthesia induction time (the time from
109 adding alfaxalone to water to the time of light anesthesia stage), surgical anesthesia induction time
110 (the time from adding alfaxalone to water to the time of surgical anesthesia stage), sedation – light
111 anesthesia (the time from sedation stage to light anesthesia stage), light anesthesia – surgical
112 anesthesia (the time from light anesthesia stage to surgical anesthesia stage), sedation – surgical
113 anesthesia (the time from sedation stage to surgical anesthesia stage) and recovery time (the time
114 from the fish transfer to the recovery tank to the time of recovery of normal approach reaction,
115 equilibrium and operculum movement scores).

116

117 **Statistical analysis**

118 Fish length was compared using Students *t* test. Fish length was reported as mean \pm standard
119 deviation (SD).

120 ANOVA analysis was performed to evaluate the times using the general linear model (GLM)
121 procedure with software package IBM SPSS Statistics vers. 20 (IBM Corp., Armonk, NY, USA)
122 with the alfaxalone concentration (three levels: 6 mg/L, 7 mg/L and 9 mg/L) as fixed factor. Later,
123 the times were covariated with the fish length. The times in minutes (min) were reported as least-
124 squares means (LSM) \pm standard error of the mean (SEM). *P* values $<$ 0.05 were considered
125 significant.

126

127 **RESULTS**

128

129 **Fish length**

130 Mean fish length was 78.1 ± 15.4 mm. Mean fish lengths in the three groups were as follow: $81.8 \pm$
131 6.8 mm (group G6), 72.2 ± 23.6 mm (group G7) and 80.2 ± 12.5 mm (group G9). There were no
132 significant differences between groups.

133

134 **Anesthetic effects**

135 After adding alfaxalone to the induction tank, the fish belonged to the groups G7 and G9 showed a
136 short excitement period during which they rapidly swam forward and backward across the tank or
137 upright with its mouth facing the surface of the water or the bottom of the tank. This behavior
138 approximatively lasted two minutes.

139 Two fish belonged to the group G7 showed the cessation of opercular movement when surgical
140 anesthesia stage was achieved. The fish were immediately removed from the anesthetic bath and a
141 steady flow of water was directed through the oral cavity and across the gills. Two fish belonged to
142 the group G9 required the same treatment because of the cessation of opercular movement when
143 they were transferred to the recovery tank. All four fish recovered slow regular opercular movement
144 in one minute after the administration of the steady flow of water across their gills.

145

146 **Induction and recovery times**

147 All the goldfish achieved surgical anesthesia stage.

148 Sedation, light anesthesia, surgical anesthesia induction times and recovery time are reported in
149 Table 3. In the group G6, sedation induction (6.00 min) time was significantly longer compared to
150 those of the groups G7 (3.80 min) and G9 (4.00 min). Light anesthesia and surgical anesthesia
151 induction times in the group G9 (8.00 and 10.20 min) were significantly faster compared to those of
152 the groups G6 (14.40 and 20.80 min) and G7 (12.60 and 19.60 min). No significant difference was
153 recorded in recovery time between groups.

154 The change in anesthetic depth was significantly faster in the group G9 compared to those of the
155 groups G6 and G7 (Table 4).

156 Fish length did not influence statistical significances (Tables 5 and 6).

157

158 **Follow up**

159 At 15 days post anesthesia, all the fish had normal equilibrium, swimming, opercular movement
160 and physical appearance, and no death was recorded.

161

162 **DISCUSSION**

163 This study demonstrates that alfaxalone 6 mg/L, 7 mg/L and 9 mg/L doses induce surgical
164 anesthesia stage in goldfish. Immersion in water concentration of 9 mg alfaxalone/L provides a
165 faster induction of light and surgical anesthesia stages compared to 6 mg/L and 7 mg/L doses. The
166 cessation of opercular movement may occur with immersion in water concentration of 7 mg and 9
167 mg alfaxalone/L.

168 Many water-soluble agents have been used for immersion anesthesia in fish (Harms 1999). The
169 ideal anesthetic agent must provide a reliable induction and adequate recovery and it should be
170 routinely stocked by the practitioners in their clinics. Propofol and alfaxalone were used to induce
171 and maintain general anesthesia in goldfish (Fleming et al.; 2003; Bauquier et al. 2013;
172 GholipourKanani & Ahadizadeh 2013; Fernández-Parra et al. 2017).

173 Alfaxalone is a neuroactive steroid that induces sedation and anesthesia because it selectively
174 modulates gamma aminobutyric receptors (Cottrell et al. 1987). These receptors have also been
175 identified in the brain of fish (Cottrell et al. 1987). Nevertheless, there are doubtful results as regard
176 alfaxalone concentration to induce surgical anesthesia in goldfish. Bauquier et al. (2013) showed
177 that alfaxalone concentration of 5 mg/L was satisfactory to induce surgical anesthesia in 83% of the
178 sample and 7.5 mg/L dose produced surgical anesthesia in 100% of the sample. In other findings,
179 alfaxalone 5 mg/L induced surgical anesthesia and the fish were able to maintain neutral buoyancy
180 at 15-35 minutes after the end of anesthesia (O'Hagan & Raidal 2006; Fernández-Parra et al. 2017).
181 Our data underscore that fish anesthetized with alfaxalone 6 mg/L, 7 mg/L and 9 mg/L achieved

182 surgical anesthesia. Potential causes of these differences include physical-chemical characteristics
183 of the water, and biological factors such as age, size, body condition and ratio of gill area to body
184 mass.

185 Water temperature, pH and osmolality influence metabolism in fish and therefore the values of
186 these parameters are important to understand the results. Lower water temperature is associated
187 with prolonged anesthetic induction and delayed recovery (Neiffer & Stamper 2009). Given that
188 goldfish are housed in cold water, it is reasonable to presume that induction times are prolonged and
189 recovery times are delayed compared to those of fish housed in warm water. Lower water pH
190 increases ionization and decreases anesthetic agent efficacy (Neiffer & Stamper 2009). The addition
191 of salt to the anesthetic bath is recommended because anesthesia alters fish osmoregulation
192 (Sneddon 2012). Furthermore, the sudden change in water temperature, pH and osmolality causes
193 stress which results in an increased gill blood flow producing a greater anesthetic absorption (West
194 et al. 2007). To reduce the influence of these factors on immersion anesthesia and to minimize the
195 stress for the fish, we used the water obtained from the house tank for the anesthetic bath and
196 recovery.

197 In this study, induction and recovery times had no correlation with goldfish size. In the
198 veterinary literature, there are no previous findings focused on relationship between goldfish size
199 and anesthetic requirements. In Atlantic salmon and brown trout, size did not influence induction
200 and recovery times (Sneddon 2012). Conversely, larger body size in whitefish, Senegalese sole and
201 Atlantic cod was associated with increased or decreased induction and recovery times (Sneddon
202 2012). The likely reason for these differences could be that each anesthetic agent has a specie-
203 specific effect because of lipid solubility and lipid content of fish, basal metabolism and gill surface
204 area (Gressler et al. 2012; Sneddon 2012).

205 The induction times recorded in this study were those to be expected based on the available
206 literature (O'Hagan & Raidal 2006; Bauquier et al. 2013; Fernández-Parra et al. 2017). Moreover,
207 as expected, 9 mg/L dose showed faster induction times compared to 6 mg/L and 7 mg/L doses. The

208 explanation of these results can be that high doses of alfaxalone may cause hypotension and reflex
209 tachycardia, which lead to an increase in blood flow through the gills and, consequently, an increase
210 in alfaxalone intake.

211 Our results underscored that alfaxalone dose had no significant influence on recovery time and
212 contradicted an expectation that higher alfaxalone concentration could increase recovery time. In
213 fact, in koi carp a higher alfaxalone dose (2.5 mg/L) produced delayed recovery compared to 1
214 mg/L dose (Minter et al. 2014). Additionally, although an ideal recovery from general anesthesia
215 must be lower than 10 minutes, the recovery times recorded in our sample were satisfactory and
216 lower compared to those reported in goldfish (O'Hagan & Raidal 2006; Bauquier et al. 2013;
217 Fernández-Parra et al. 2017) and koi carp (Minter et al. 2014). The likely reason for the observed
218 differences is that surgical anesthesia was maintained for two minutes in the present study, a time
219 significantly shorter compared to other reports (O'Hagan & Raidal 2006; Minter et al. 2014;
220 Fernández-Parra et al. 2017).

221 Higher alfaxalone doses caused excitatory movements. Side effects previously described in fish
222 anesthetized with alfaxalone are as follow: unilateral horizontal nystagmus in koi carp (Minter et al.
223 2014) and increased hearth rate in oscar fish (Bugman et al. 2016). We did not record any of these
224 side effects, but an interesting observation is that a hyperactivity period was recorded after adding
225 alfaxalone to the induction tank. In goldfish, the authors are unaware of previous findings
226 reporting a hyperactivity period after adding alfaxalone. However, a short excitement period was
227 described during induction phase of immersion anesthesia in some fish species (Stetter 2001). The
228 mild excitement phase may be due to the chemical irritation of the gills.

229 Respiratory depression is the most common side effect recorded during immersion anesthesia. In
230 the present study, cessations of opercular movement were recorded with high alfaxalone doses,
231 similarly to a previous finding (Fernández-Parra et al. 2017). One explanation of these results is that
232 higher doses speed up the change in the depth of anesthesia and, if anesthetic induction is too fast,
233 differentiating one anesthetic stage from another may be difficult and side effects may rapidly arise.

234 The cessation of opercular movement can lead to hypoxia that induces gills filament collapse and
235 delayed recovery (Harms 1999). Notwithstanding the cessation of opercular movement can be
236 resolved by transferring the fish to a recovery tank (Fernández-Parra et al. 2017), we preferred to
237 deliver a steady flow of oxygenated water through the oral cavity and across the gills in order to
238 stimulate the buccal flow/heart rate reflex. Although goldfish are anoxia tolerant compared to other
239 species (Bickler & Buck 2007), a constant monitoring of anesthetic depth is required to avoid
240 cardiorespiratory complications.

241 A limitation of this study is the assessment of depth of anesthesia. Activity, equilibrium, reaction
242 to tactile or noxious stimuli and opercular movement have been widely used to determine the depth
243 of anesthesia in fish, but these parameters depend on species, drug and dose (Neiffer & Stamper
244 2009; Sneddon 2012; Bauquier et al. 2013). In the present study, the response to noxious stimulus
245 was assessed with a pressure applied to the caudal dorsal fin. The disadvantage of this method was
246 the inaccurate repeatability of applied force.

247 In conclusion, the present study shows that alfaxalone may be considered a reliable anesthetic
248 induction agent in goldfish. Immersion in water concentration of 6 mg alfaxalone/L provides a
249 smooth induction of surgical anesthesia stage in approximately 20 minutes without side effects.
250 Higher doses shorten induction times, and cause respiratory depression and excitatory movements.
251 Induction and recovery times have no correlation with goldfish size. Additional studies are needed
252 to evaluate the efficacy and safety of alfaxalone for a longer duration.

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302 **TABLES**

303 Table 1. Assessment of approach reaction, equilibrium, operculum movement and reaction to
 304 noxious stimuli using ordinal scales (Bauquier et al. 2013, modified)

Approach reaction score	
0	Normal: swims away when approached
1	Reduced: slower to react and slower swimming
2	Low: slow to react and may not react reliably
3	Lost: no reaction to approach or contact
Equilibrium score	
0	Normal: strongly retains upright position when still and swimming
1	Reduced: wobbles from upright position when still and swimming
2	Low: leans or lies on side or may turn upside down, returns to upright when swimming or when stimulated
3	Lost: leans or lies on side or may turn upside down, does not return to upright and generally does not swim
Operculum movement score	
0	Normal: rate will be recorded
1	Reduced: slowing of the operculum rate
2	Slow: slow but steady rate
3	Lost: no operculum movement
Response to noxious stimuli score	
0	Normal: strong tail wiggle (at least 5 movements in 1 to 2 s)
1	Reduced: reduced tail wiggle (3 to 5 movements in 1 to 2 s)
2	Slow: weak tail wiggles (1 to 3 movements in 1 to 2 s)
3	Faint: tail flinch only (1 movement in 1 to 2 s)
4	Lost: no tail wiggles or movement

305 s, seconds.

Stages	Level of consciousness	Behavior
Stage 0	Normal	Normal equilibrium, normal operculum movement, normal response to visual and tactile stimuli
Stage 1	Sedation	Normal to reduced equilibrium, reduced operculum movement, reduced response to visual and tactile stimuli
Stage 2	Light anesthesia	Loss of equilibrium, reduced to slow operculum movement, loss reaction to visual stimuli, slow to faint response to tactile stimuli
Stage 3	Surgical anesthesia	Loss of equilibrium, slow operculum movement, loss of reaction to visual or tactile stimuli
Stage 4	Medullary collapse	Loss of equilibrium, loss of operculum movement, loss of reaction to visual and tactile stimuli, cardiac arrest and death

308 Table 3. Sedation, light anesthesia, surgical anesthesia induction times and recovery time in minutes
309 (expressed as LSM \pm SEM) in the three groups

Anesthetic stages	Group			SEM	<i>p</i>
	G6	G7	G9		
Sedation	6.00 ^b	3.80 ^a	4.00 ^a	0.400	0.031
Light anesthesia	14.40 ^b	12.60 ^b	8.00 ^a	1.040	0.020
Surgical anesthesia	20.80 ^b	19.60 ^b	10.20 ^a	1.843	0.022
Recovery	14.80	18.20	16.80	1.077	0.465

310 ^{a,b}Significant differences in the row between groups. The alphabetical order indicates the order of
311 the data.

312 Table 4. Intervals of time in minutes (expressed as LSM \pm SEM) elapsed between sedation and light
313 anesthesia, light anesthesia and surgical anesthesia, sedation and surgical anesthesia

Intervals of time	Group			SEM	<i>p</i>
	G6	G7	G9		
Sedation - light anesthesia	8.40 ^b	8.80 ^b	4.00 ^a	0.896	0.038
Light anesthesia - surgical anesthesia	6.40 ^b	7.00 ^b	2.20 ^a	0.981	0.084
Sedation - surgical anesthesia	14.80 ^b	15.80 ^b	6.20 ^a	1.741	0.032

314 ^{a,b}Significant differences in the row between groups. The alphabetical order indicates the order of
315 the data.

316 Table 5. Data covariated with the fish length. Sedation, light anesthesia, surgical anesthesia
 317 induction times and recovery time in minutes (expressed as LSM \pm SEM) in the three groups

Anesthetic stages	Group			SEM	<i>p</i>
	G6	G7	G9		
Sedation	5.89 ^b	3.97 ^a	3.94 ^a	0.400	0.046
Light anesthesia	13.79 ^b	13.55 ^b	7.65 ^a	1.040	0.000
Surgical anesthesia	19.75 ^b	21.24 ^b	9.60 ^a	1.843	0.001
Recovery	14.40	18.82	16.57	1.077	0.288

318 ^{a,b}Significant differences in the row between groups. The alphabetical order indicates the order of
 319 the data.

320 Table 6. Data covariated with the fish length. Intervals of time in minutes (expressed as LSM \pm
 321 SEM) elapsed between sedation and light anesthesia, light anesthesia and surgical anesthesia,
 322 sedation and surgical anesthesia

Intervals of time	Group			SEM	<i>p</i>
	G6	G7	G9		
Sedation - light anesthesia	7.90 ^b	9.58 ^b	3.71 ^a	0.896	0.002
Light anesthesia - surgical anesthesia	5.96 ^b	7.69 ^b	1.95 ^a	0.981	0.028
Sedation - surgical anesthesia	13.86 ^b	17.27 ^b	5.66 ^a	1.741	0.002

323 ^{a,b}Significant differences in the row between groups. The alphabetical order indicates the order of
 324 the data.