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Modified PNA design and synthesis: a novel
approach using Molecular Dynamics and
Metadynamics

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1. Introduction

1.1. Modern challenges in drug discovery

Every year, millions of people all over the world come into contact with hard reality of a genetic disease or a tumor. Treatment of this pathologies is a great challenge for science, comparable to that against bacteria at the beginning of last century.¹ Penicillin, discovered by Fleming in 1928, started revolution of antibiotics that led, in a few time, to cure most of bacteriological infections. Treatment of genetic diseases is a complex problem that requires a multidisciplinary approach involving many different fields, ranging from medicine, biology, biotechnology, which provide pre-clinical and clinical studies, to chemistry, which allows the synthesis of new compounds, and to physics and engineering, which are necessary for the development of new materials. During the last decades, huge efforts have been done to make therapies more effective, to find new ones or, in general, to improve the quality of life of patients and their life expectation. Many steps forward have been done in particular for tumors, for which 5 years surviving is nowadays averagely of about 60%. Alzheimer, Cystic Fibrosis, Glioblastoma, Pancreatic cancer are just some, among many, that unfortunately are nowadays still almost incurable. Prevention is the first mean to sensitize population about bad habits and thus reducing incidence of some pathologies. Studies on mechanism of action of diseases allowed in many cases to better understand causes and to isolate new targets for the therapies.^{2,3} Advancements in Science have indeed also provided new tools for early diagnosis and targeted therapy. The major approach for the development of new bioactive compounds have traditionally concerned small molecules. Drug discovery process was simplified using combinatorial approach or screening of molecules derived from natural products; prediction of the behavior of these could be done computationally using docking techniques.⁴ Small molecules are relatively easy to synthesize and cheaper to scale up. On the other hand, often they could bind multiple targets causing many side effects difficult to predict and thus failure of many drugs at clinical phase tests can occur. Examples of commonly used drugs for treatment of cancer are reported in Figure 1.1.

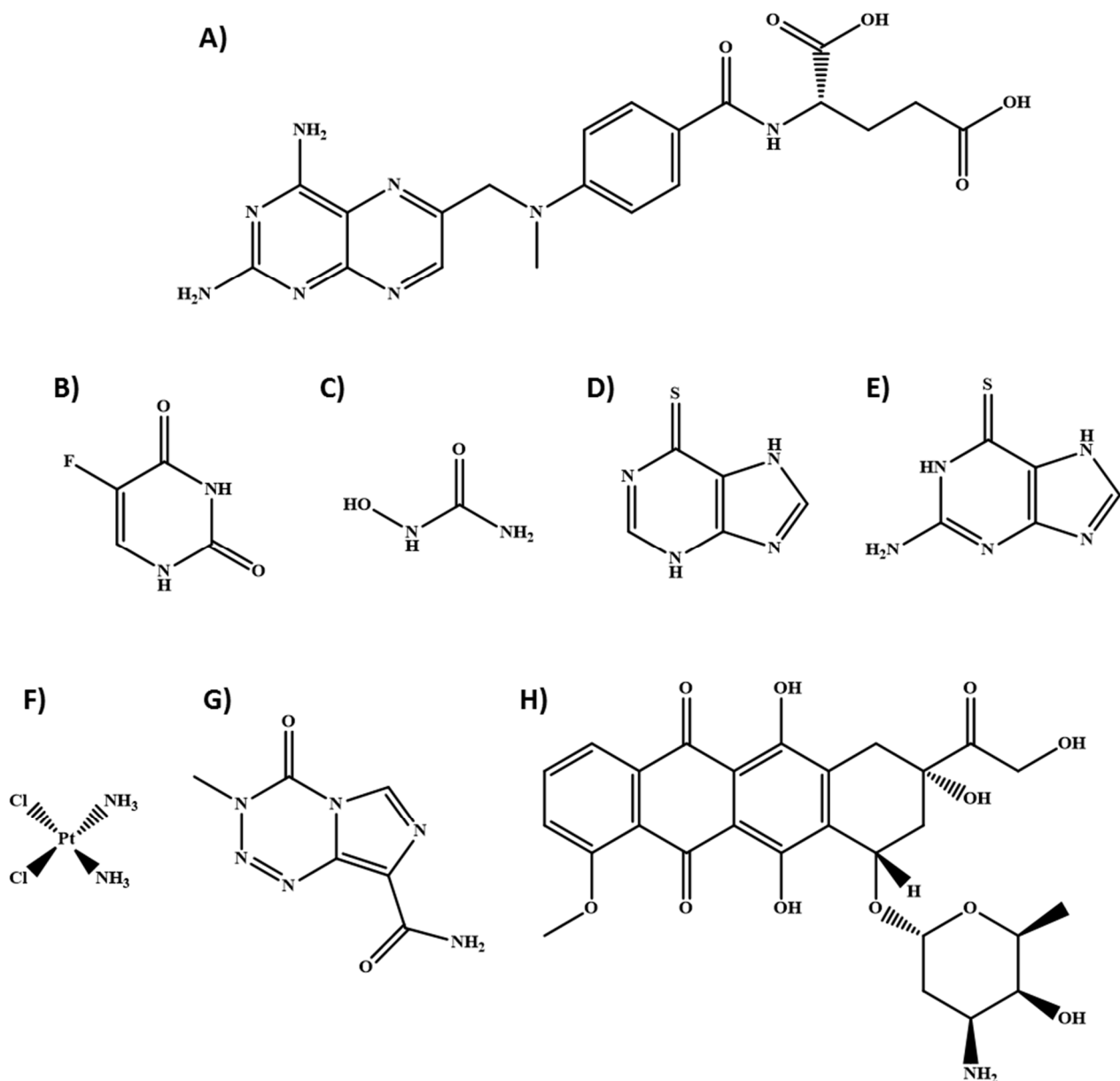


Figure 1.1: examples of antitumor agents used for therapy. **A)** Methotrexate; **B)** 5-Fluoro Uracil; **C)** hydroxyurea; **D)** Mercaptopurine; **E)** Thioguanine; **F)** Cisplatin; **G)** Temozolomide; **H)** Doxorubicin.

Generally, they follow two mechanisms of action: stop of DNA building blocks synthesis or DNA and RNA alkylation. Compounds from **A** to **E** belong to the first class. Methotrexate inhibits folic acid reductase that is responsible for conversion of folic acid to tetrahydrofolic acid.⁵ This compound is a common intermediate necessary for the production of a cofactor used for nucleoside synthesis. Compounds **B**, **C**, **D** and **E** inhibit selectively one nucleotide formation and respectively thymine,⁶ cytosine,⁷ adenine⁸ and guanine.⁹ Compounds from **F** to **H** belong instead to the second class of chemotherapeutic agents. Cisplatin is one of the first and simplest drugs used in tumor treatment, but is still used together other platinum based compounds.¹⁰ Temozolomide is used as adjuvant against

some brain cancers,¹¹ and Doxorubicin and its derivatives are used as therapy for leukemias or lymphomas.¹²

Alongside this small-molecule approach for the treatment of cancers, new innovative strategies are being developed in the recent years in order to maximize effectiveness and selectivity. For example, the cell therapy, which consists in diseases treatment by injection of cellular material, has recently reached a translational level. Many different types of cells are used and, among others, Stem Cells and T Cells have showed interesting therapeutic properties. Stem Cells have been proposed for their ability to differentiate in different specialized cells. This particular behavior finds then application in regenerative medicine, allowing to restore damaged tissues. Nowadays these cells are widely studied for treatment of neurodegenerative diseases,^{13,14,15} with the aim to recover damaged neurons and restore normal cerebral activity. T cells are a type of lymphocytes and have been used for immunotherapeutic approach.¹⁶ These cells are able to kill cancer cells and, in particular, their resulted effective also in treatment of metastasis.^{17,18} For example, Rosenberg and coworkers reported use of engineered T lymphocytes for effective reduction of metastatic melanoma,¹⁹ inducing rapid reduction of the tumor and absence of tumor recurrence one year after the treatment. However, cure of diseases with cells presents some drawbacks, in particular costs, since culture results in extremely low yields, and difficult delivery.

Use of therapeutic antibodies is another approach diffused in recent years. In particular several monoclonal antibodies (mAbs) have been studied and nowadays are applied in diagnostic^{20,21} and treatment of different pathologies.²² Antibody engineering allowed to generate new artificial compounds able to selectively bind targets, but at the same time to avoid rejection and clearance by patient's immune system.²³ Chimeric antibodies represent the first generation of compound in which human constant domains are fused with murine variable ones (human portion ~70%). Humanized antibodies represent instead the second generation and present only a little fraction of murine domains (human portion 85-90%). Mechanisms of action of these compounds is not always clear or can derive from multiple effects. Most common pathways consist in a direct action of mAbs that can interact with specific receptors on the surface of cells or in stimulating immune-mediated tumor cell killing. Monoclonal antibodies can be used also in conjugation with drugs or toxins that can be released in cells.²⁴ Rituximab is one of the best examples of commercial antibody. It is a chimeric compound against protein CD20, which is mainly present on the surface of B cells. Therefore, is used in case of excess or over reactivity or dysfunctionality of this type of cells. Nowadays Rituximab is commonly administered in treatment of different diseases like lymphomas,

leukemias and other autoimmune disorders.^{25,26} Despite their unique properties of activity and selectivity, monoclonal antibodies are large proteins (~ 150 kDa) and their synthesis is still very expensive. Moreover, they must be administered in large amounts to achieve clinical results due mainly to scarce permeation toward target cells.

Nanomedicine is another field of research well explored in recent years. Advance of chemistry in the field of materials allowed to easily produce a large variety of compounds. Nanomaterials apply to both diagnostics,^{27,28} and therapeutics,^{29,30} often with the goal of achieving both effects (theranostic).³¹ Due to their properties of penetration in cells, nanoparticles are good candidates for drug delivery.³² A first generation of nanocarriers consisted in passive delivery systems (Figure 1.2A). The most known compounds of this type, that are in clinical use, are liposomes,³³ in which drugs are trapped in vesicles constituted by lipid bilayers, and mixed structures such as nab-paclitaxel,³⁴ in which toxic paclitaxel is bound to albumin nanoparticles. This kind of systems often are nonspecific toward tumors, causing undesired side effects. This led to a second generation of nanomaterials in which nanocarriers are functionalized with antibodies,³⁵ peptides,^{36,37} ligand^{38,39} and other biomolecules^{40–42} in order to discriminate target cells. Many examples are available in the literature, but the use of these compounds in clinic is still limited. Particular attention has been given to biocompatibility of these compounds, since some types of nanoparticles may result very toxic.⁴³

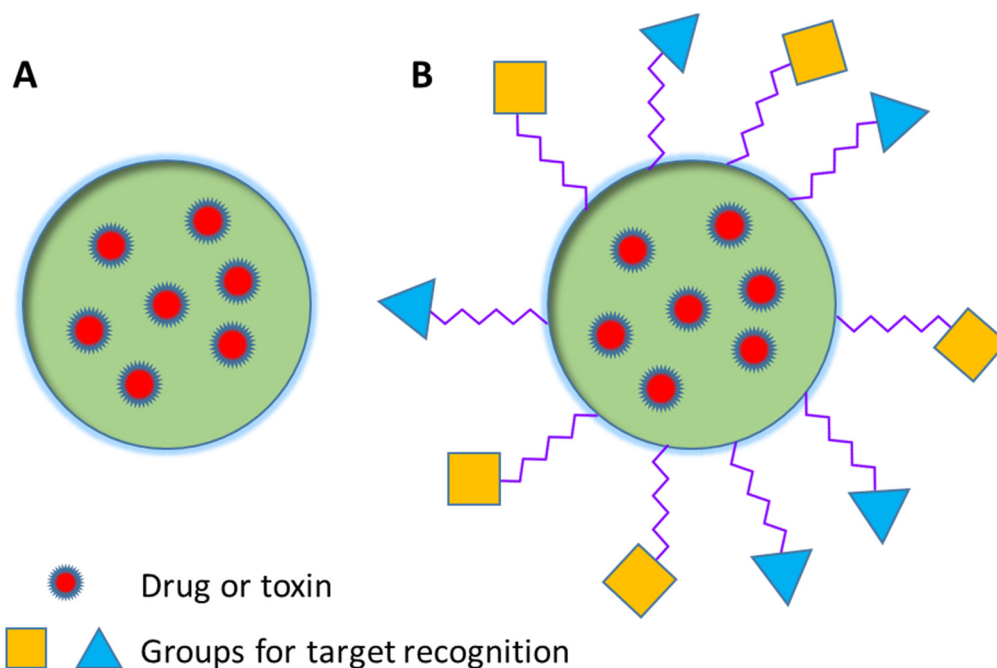


Figure 1.2: Schematic representation of first (A) and second (B) generation nanostructures for drug delivery.

Finally, the treatment of diseases by rationally targeting of nucleic acids by bio-macromolecules is a long-standing, and still very active, field of research, which is the subject of this thesis and will be discussed in detail in the following paragraphs.

1.2. Nucleic Acids Targeting

Since many diseases are linked to DNA expression and mutation,^{44,45} a part of the research focuses on genes or on transcription, transport and translation of their information. Main processes involved in gene expression are represented in Figure 1.3

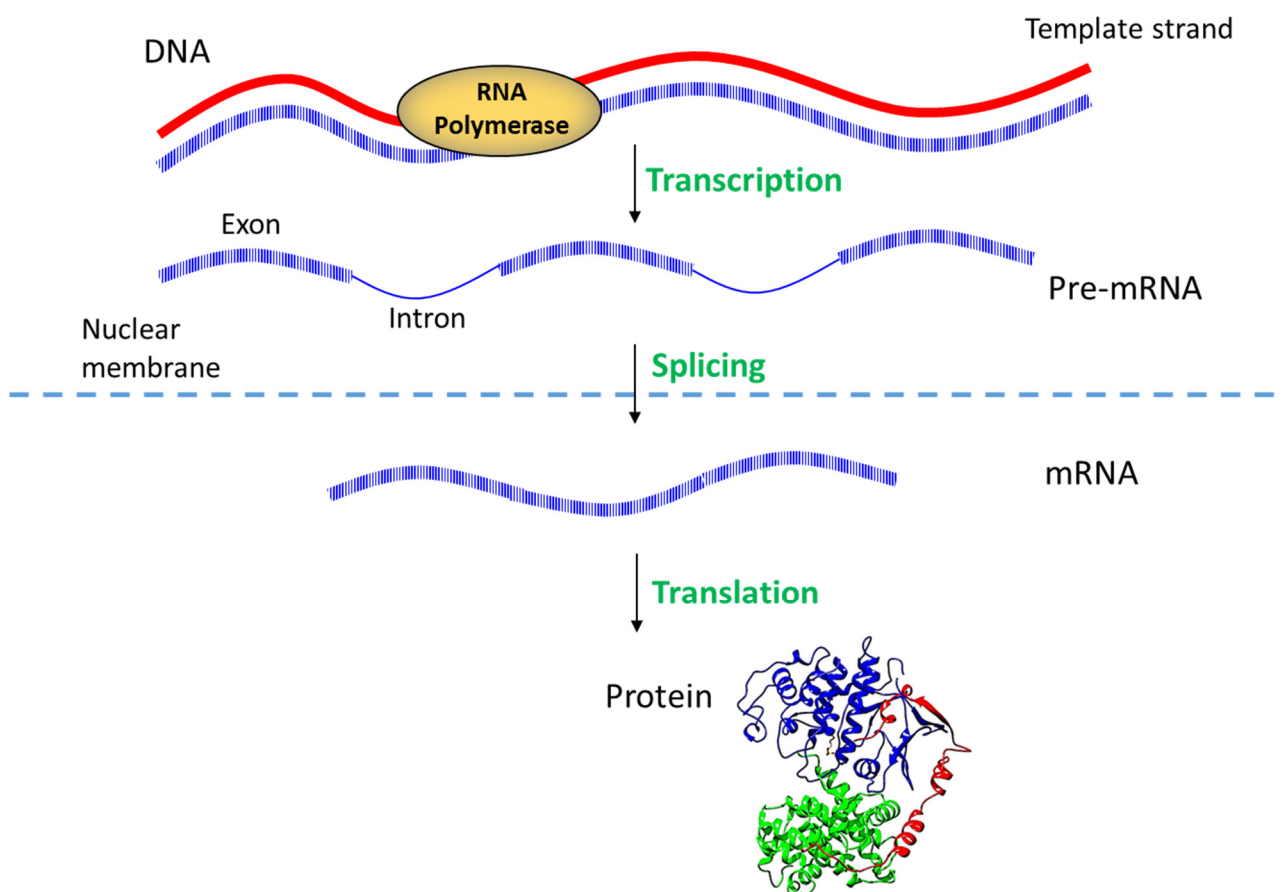


Figure 1.3: Schematic representation of gene expression processes.

The antigene approach consists in directly targeting DNA.⁴⁶ Indeed, it is possible to bind double stranded DNA with oligonucleotide fragments or artificial analogues, forming triplex structures or invading double helix and thus interfering with transcription. However, direct interaction with DNA is difficult because the targeting molecules must enter into cells and then into nucleus.⁴⁷ Another

possibility for affecting transcription is the use of decoy compounds⁴⁸ for transcription factors. These proteins are able to recognize specific gene sequences and then promote or block RNA polymerase activity. Decoys bind to these factors preventing or reducing interaction with target DNA and thus controlling transcription process.⁴⁹

Pre-mRNA is a precursor of messenger RNA in which exons and introns are both present. To obtain mature mRNA is necessary to remove introns and correctly join exons (splicing); this process is usually performed by spliceosomes and often is possible to obtain different mRNAs from the same pre-mRNA, by changing exons recombination. Physiological conditions can then influence production of different proteins, but it is also possible to have errors in this phase leading to non-functional or partially functional proteins (aberrant splicing). Oligonucleotides or analogues are used for directing splicing process⁵⁰ or to block aberrant slicing favoring normal one.⁵¹

The antisense approach⁵² consists in targeting messenger RNA. Formation of a duplex with mRNA prevents translation and RNase-H mediated cleavage of RNA strand.⁵³ mRNAs are found in cytoplasm and are then easier to target. Similarly, it is also possible to bind micro RNA using an anti-miR approach (this methodology will be discussed further in section 1.3).^{54,55}

DNA or RNA oligonucleotides can directly target specific complementary sequences by the Watson-Crick base pairing scheme and can thus provide a rational design of new drugs; the use of this approach using naturally occurring nucleic acids in pharmaceutical applications is difficult because DNA, and even more RNA, are quite unstable in biological media due to the presence of nucleases. Moreover, the cellular uptake of naturally occurring nucleic acids was found to be extremely poor due to the phosphate negative charge. To overcome these problems, it has been necessary to move to modified artificial DNAs. These systems must retain, and hopefully increase, ability to bind to the complementary sequence of DNA and at the same time present better resistance, cellular uptake and, possibly, new functionalities. Among the first proposed compounds (Figure 1.4), phosphorothioates⁵⁶ or alkyl phosphonate⁵⁷ presented modifications on the phosphate group; in the first case an atom of oxygen was substituted with sulfur, while in the second with an alkyl group. In both cases modifications induced chirality in the system leading to two different diastereoisomer with different behaviors.⁵⁸

2'-O-Alkyl⁵⁹ (and in particular 2'-O-methyl)⁶⁰ or 2'-fluoro-oligonucleotides⁶¹ were the first examples of sugar modified nucleotides proposed to substitute RNA. Indeed, since the OH in 2' position greatly enhance hydrolysis rate, compared to DNA, due to possible intramolecular attack on phosphate, substitution at this site enhances the stability of these derivatives.

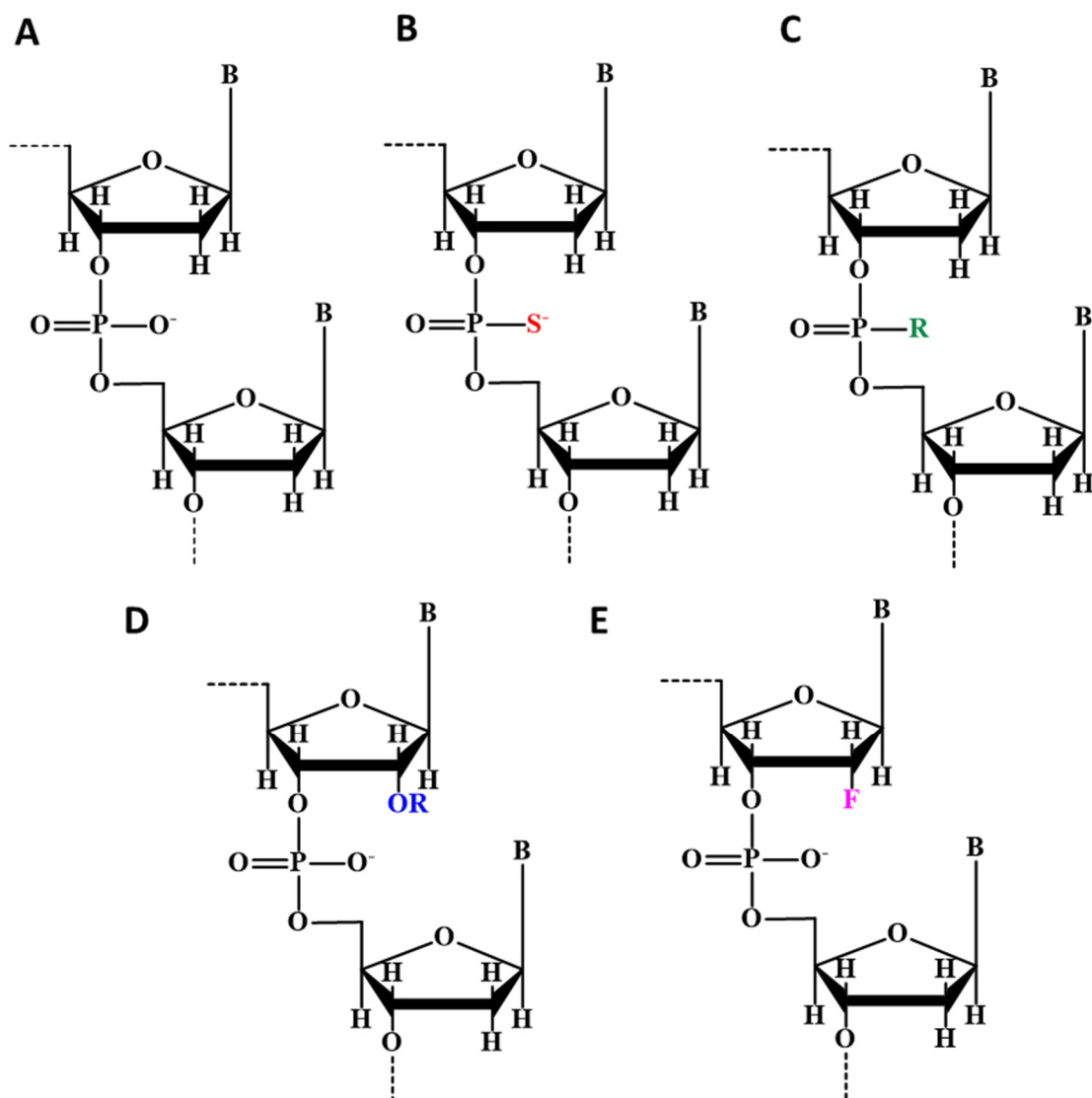


Figure 1.4: A) standard DNA; B) Phosphorothioate; C) alkyl phosphonate; D) or 2'-O-Alkyl RNA; E) 2'-Fluoro RNA modifications.

Tricyclo DNA⁶² (Figure 1.5) are examples of conformationally constrained DNA that are not recognized by nucleases and recognize RNA with high affinity.⁶³ Threose Nucleic Acids⁶⁴ are analogues in which ribose is substituted with threose sugar ring. Although TNAs are artificial compounds they are thought as possible precursor of DNA or RNA.^{65,66} Among sugar modified systems, Locked Nucleic Acid (LNA) are very effective ones and present high affinity and selectivity toward target sequence.⁶⁷ These type of compounds present oxygen in 2' position linked to 4' carbon with a methylene bridge, thus reducing hydrolysis. Moreover, this linkage blocks the

nucleoside unit in a C3' endo (N-type) conformation, which is typical of RNA. Many variations of this system have been proposed during the years.⁶⁸

It is also possible to replace sugar rings and phosphate with different units. The class of morpholino^{69,70,71} oligonucleotides is an example in which the nucleic acid backbone is substituted with methylenemorpholine rings and phosphorodiamidate linkages; triazole DNAs^{72,73} present a imidazole ring covalently bound to ribose.

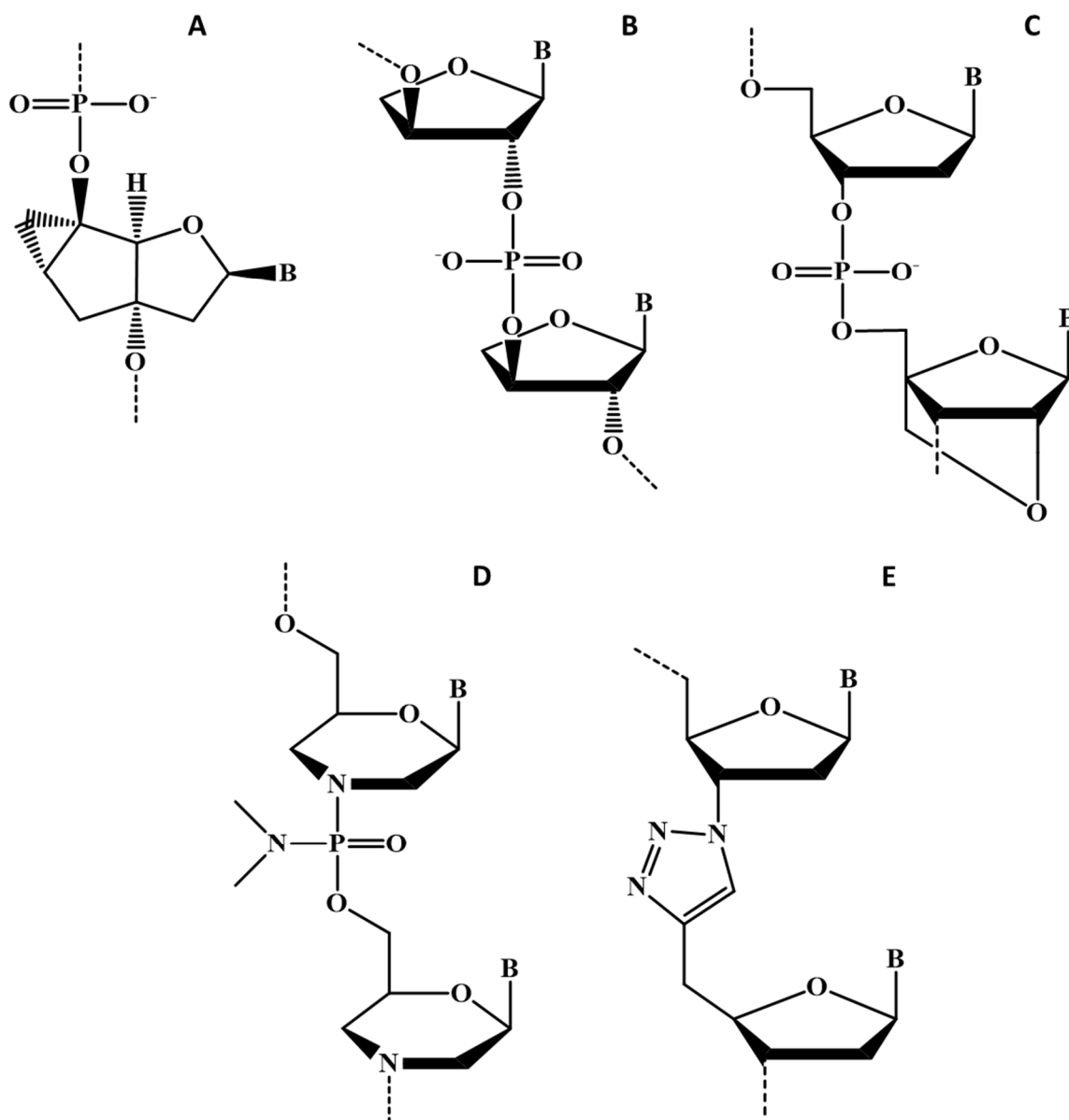


Figure 1.5: A) Tricyclo DNA; B) TNA; C) LNA; D) Morpholino phosphodiamidides; E) Triazole DNA modifications.

1.3. Micro-RNA as therapeutic targets

MicroRNAs (miRNAs) are small RNAs, 21–25-nucleotide long, that regulate gene expression at the post-transcriptional level.⁷⁴ miR up- or down-regulation has been correlated with many different diseases like hearing loss,⁷⁵ hearth failures^{76,77,78} and, most notably, different types of cancer,^{79,80} including leukemia.^{81,82,83} Gene expression is regulated by guiding RNA-induced silencing complex (RISC) that is able to bind target mRNA and to degrade it (Figure 1.6).

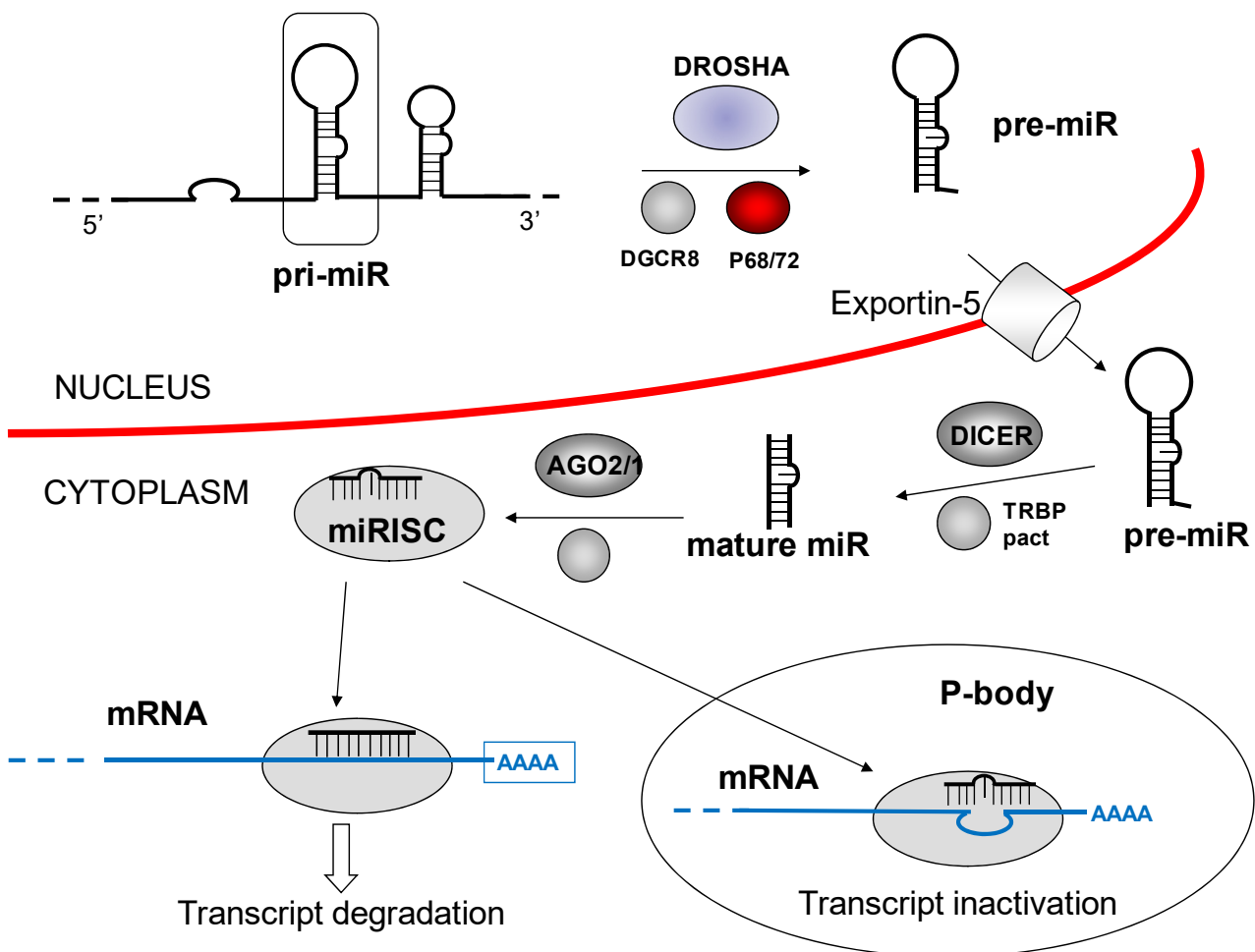


Figure 1.6: Mechanism of maturation of microRNAs and their activity through RISC complex.

As an example of the potentiality of miR-based therapeutics, the work by Giacca and coworkers,⁸⁴ who studied miR connected to cardiac growth, is worth citing. In embryos enlargement of the hearth is principally due to proliferation of cardiomyocytes; this process ends at birth and growth is then connected to hypertrophic enlargement of myocytes. miR 590 and miR 199a were found to stimulate cardiomyocyte proliferation and were used to promote cardiac regeneration. Infarcted

mice treated with this two miR showed a regeneration of cardiac tissue and an almost complete recovery of functional parameters (Figure 1.7). Thus, this miR based approach is the only available example of possible cure of infarctuated heart by administration of drugs.

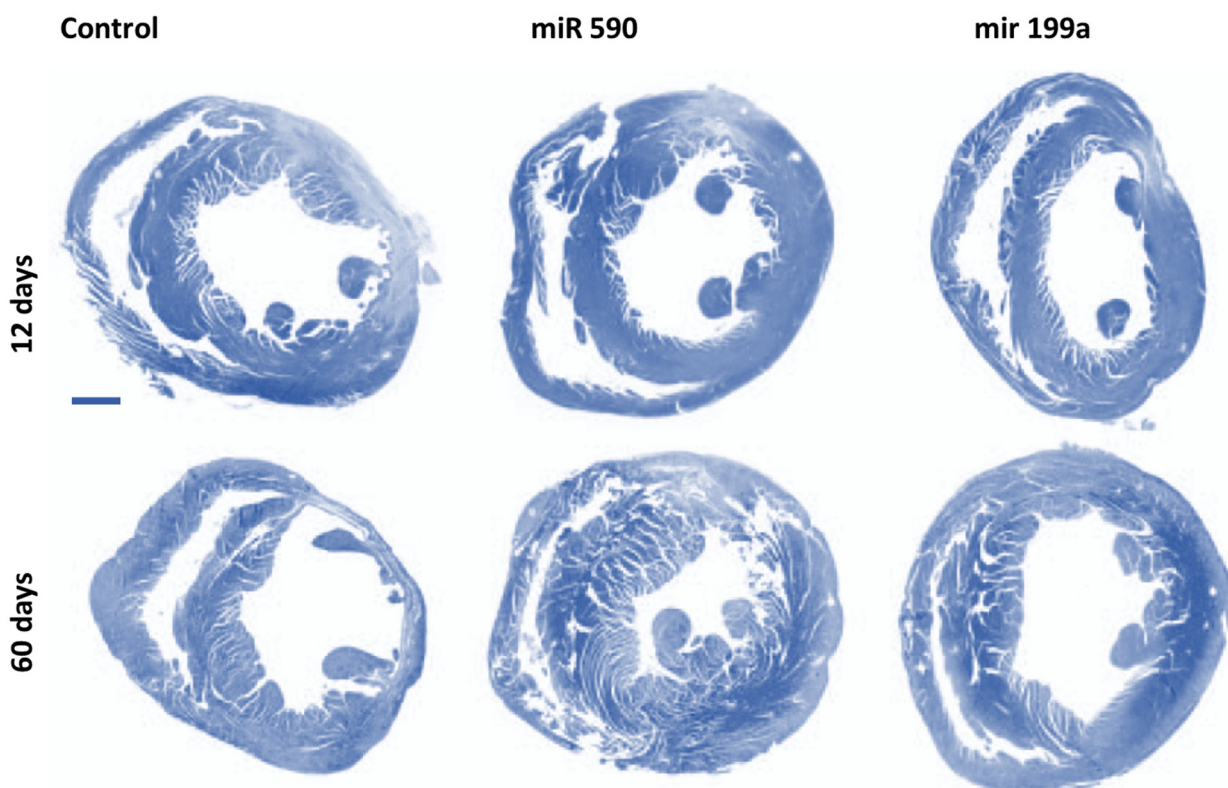


Figure 1.7: Heart cross section of infarcted mice after 12 days and after 60 days. From left, control (untreated mouse), mouse treated with miR 590 and mouse treated with miR 199a. Reprinted from ref. 84 with the permission of Nature Publishing Group.

Due to problems explained before, usually use of modified systems is preferred. AntagomiR are synthetic 2'-O-methyl RNA (Figure 1.4D) that are able to strongly bind micro RNA.⁸⁵ In order to further increase resistance to nucleases usually phosphorothioate modifications are inserted on the first two and last four bases and cholesterol is linked at 3' end to enhance distribution and cell permeability.⁸⁶ These type of compounds is commercially available and is actually tested for treatment of several pathologies.^{87,88,89}

LNA have also been successfully used to target microRNA, for example in treatment of Hepatitis C Virus.⁹⁰ Miravirsen is a LNA/phosphorothioate hybrid that is able to efficiently bind miR-122 (Figure 1.8) that is overexpressed in affected liver cells, and activates virus replication.⁹¹ This drug has demonstrated broad antiviral activity and it is actually in phase 2 clinical trial.

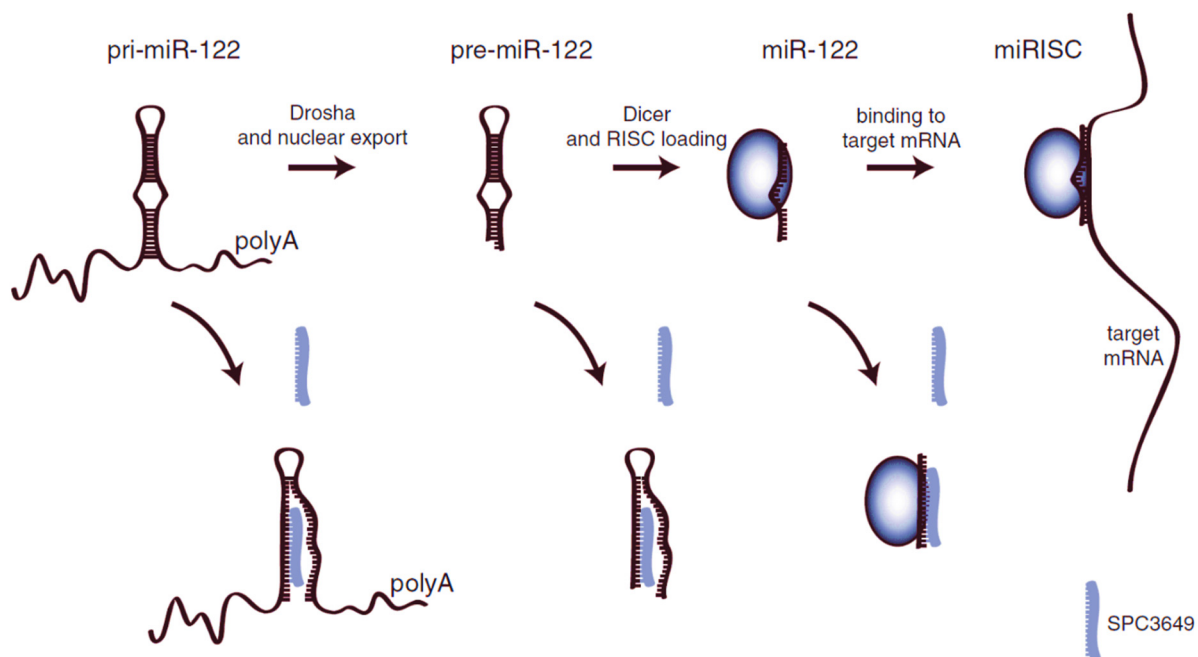


Figure 1.8: Proposed mechanism of action of anti-miR drugs; Miravirsin (SPC3649) is taken as an example. Reprinted from ref. 90 with the permission of Oxford University Press.

An emerging challenging application is use of peptide nucleic acids to inhibit action of micro-RNAs by targeting their sequences.

1.4. Peptide Nucleic Acids

1.4.1. General properties

Peptide nucleic acids (PNAs)⁹² are particularly interesting DNA mimicking compounds, in which the deoxyribose phosphate backbone is substituted by *N*-(2- aminoethyl)glycine units and bases are linked to the backbone with methylenecarboxylic moieties (Figure 1.9).

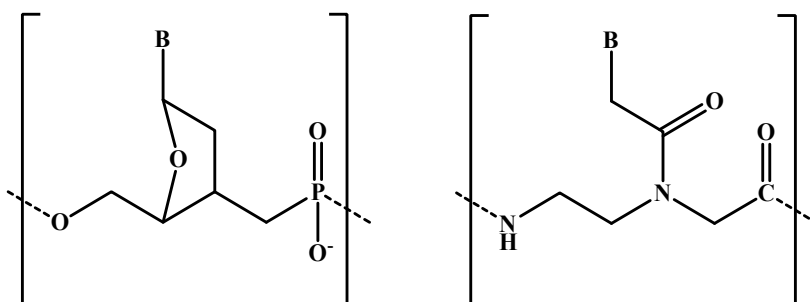


Figure 1.9: left, structure of DNA; right, general structure unmodified Peptide Nucleic Acids.

Like DNA, generally PNAs form a double helix with the target DNA sequence through Watson and Crick bonds;⁹³ in the case of homopyrimidine sequences PNA can also form triplex structures with homopurine DNA strands by forming both Watson-Crick and Hoogsteen bonds.⁹⁴ Depending on the sequence, pairing in PNA:DNA duplexes can occur both in parallel and antiparallel orientations (as defined in Figure 1.10), but generally the latter is preferred.⁹⁵

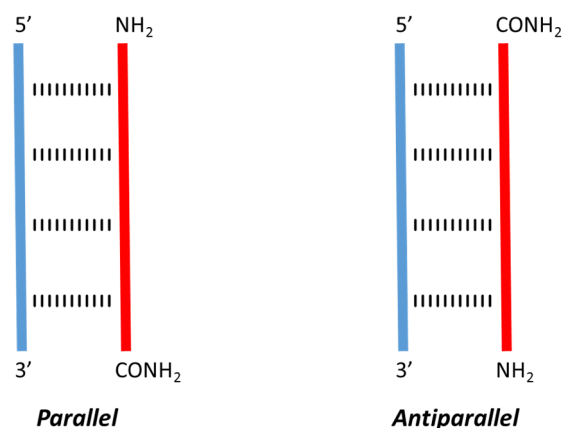


Figure 1.10 : Possible orientations for PNA:DNA (RNA) duplexes. Blue strand is DNA or RNA, while red strand is PNA.

The amidic backbone of PNA is neutral, therefore repulsive electrostatic effects, present in DNA or RNA duplexes, are avoided, and this induces high stability of the PNA duplexes with complementary DNA or RNA. Furthermore, compared to oligonucleotides or their analogs, PNA present higher specificity in sequence recognition, even for a single base mismatch.⁹⁵ Moreover previous studies revealed that PNAs display high chemical and biological stability,⁹⁶ making them particularly suitable for *in vivo* use.

PNA can be obtained with protocols similar to those used for peptides: synthesis is routinely performed on a resin and monomers require presence of a temporary protecting group, that permit chain elongation, and of semi-permanent groups, that protect nucleobases, avoiding side reactions or multiple chains grow (Figure 1.11). Many strategies have been proposed, but Fmoc/Bhoc⁹⁷ and Boc/Cbz⁹⁸ ones are the most used and thus the corresponding protected monomers are commercially available.

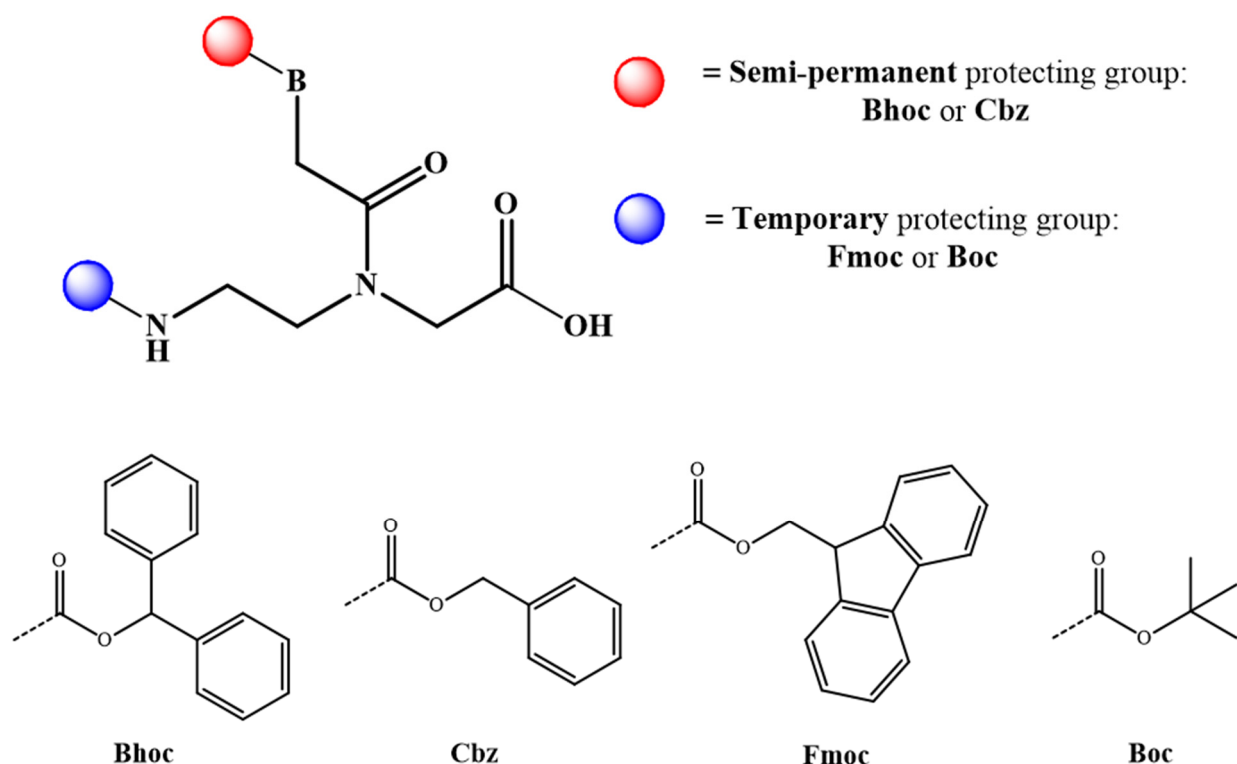


Figure 1.11: Representative monomer structure for solid phase synthesis of PNA.

PNAs have been successfully used in different field ranging from analytical methods, in which they are used as probes in sensors for DNA recognition,^{99,100,101} to diagnostics, in which they are coupled with reporter group for imaging,^{102,103} and, recently, to therapeutics, as potential new drugs.^{104,105}

1.4.2. Modifications

Since their introduction in 1991, many modifications have been proposed to improve PNAs properties or to introduce new functionalities. Modifications can be introduced on the backbone,^{106,107} or on the nucleobases.^{108,109} Those on the backbone usually were designed to affect the conformational freedom of the system and, in general, to reduce entropy loss during duplex formation.¹¹⁰ Insertion of chiral substituents in C2 or C5 position of the backbone (Figure 1.12) can modify handedness of the helix,¹¹¹ favoring or disfavoring the binding with DNA.

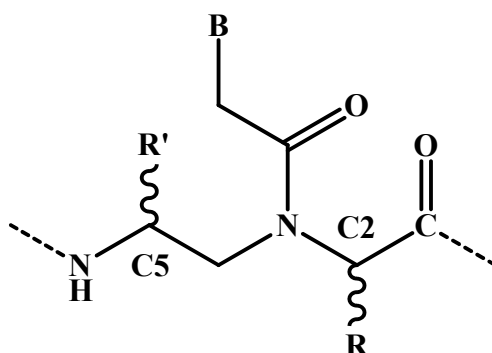


Figure 1.12: Schematic representation of PNA bearing modifications in C2 and C5 carbon atoms.

Substituent groups in C2 position, derived from D-amino acid synthons, induce preference to form a right handed helix, while L groups induce preferred left-handedness; for C5 position the stereochemical preference is inverted, since predisposition to form right-handed helices was observed for the derivatives obtained from L-amino acids. Maximum positive effect on PNA:DNA duplex stability was found with a combination of both modification: a D-amino acid derived group in C2 position and a L-amino acid derived group in C5, leading to a stabilization of $\sim 7^\circ\text{C}$ compared to unmodified PNA. Opposite case, with a constructive left-handed induction, caused a drop of T_m of more than 30°C (Table 1.1).¹¹¹

Table 1.1 : Effect of chiral substituents in C2 and C5 positions. Table is taken from ref (111).

PNA	Helical induction C2	Helical induction C5	Overall helical preference	Chiral accordance/conflict	PNA–DNA Thermal stability [$^\circ\text{C}$]
2D,5L	right-handed	right-handed	right-handed	accordance	57
5L	–	right-handed	right-handed	–	56
2L,5L	left-handed	right-handed	right-handed	conflict	52
2D	right-handed	–	right-handed	–	52
Achiral	–	–	–	–	50
(2L	left-handed	–	left-handed	–	47
2D,5D	right-handed	left-handed	left-handed	conflict	33
5D	–	left-handed	left-handed	–	32
2L,5D	left-handed	left-handed	left-handed	accordance	20

Insertion of groups in C2 position modify also the selectivity of PNA probe. “Chiral box” PNA (Figure 1.13) with three consecutive D-lysine monomers in the middle of the sequence led to a high drop of stability in presence of a single mismatch.^{112,113} D-Lysine containing chiral box PNA was effective in

inducing preference for the antiparallel orientation (with high direction control) while L-Lysine containing PNA favored the parallel one.

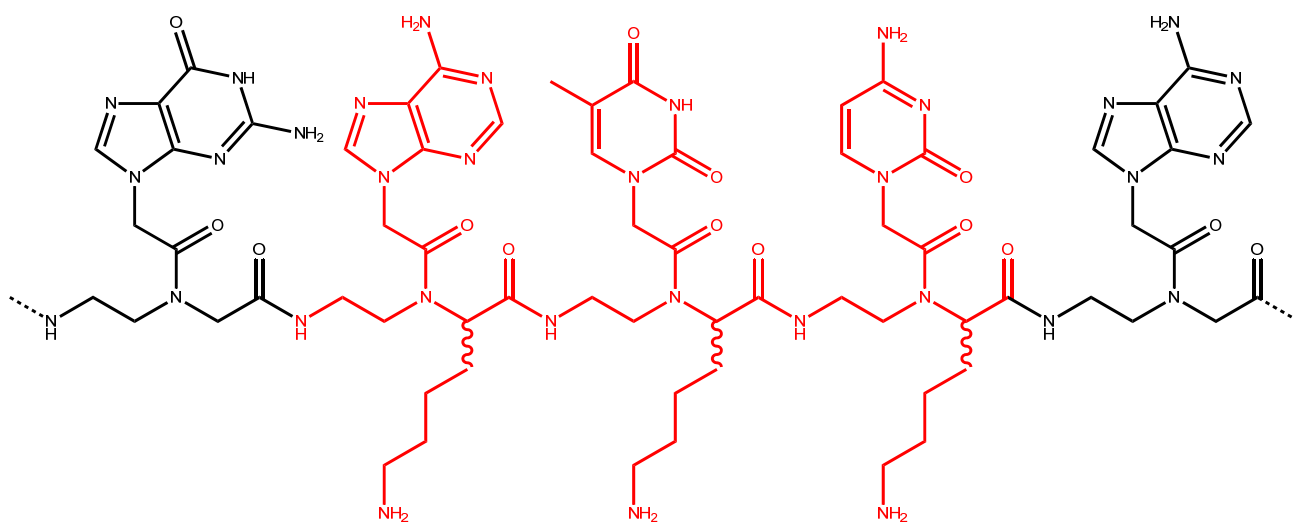


Figure 1.13: "Chiral Box PNA" bearing Lysine group in C2 position.

On the contrary, C5 modification influence principally the preorganization of the system, proved by strong circular dichroism (CD) signals for single strand PNA.¹¹⁴ Insertion of positively charged groups in the same position favored cellular uptake of the PNA,^{115,116} it can also be improved by conjugation with carrier peptides,¹¹⁷ like octa-arginine¹¹⁸ or NLS.¹¹⁹ However, incorporation (embedding) of the amino acid side chains into the PNA structure was shown to increase resistance to peptidases.¹¹⁸ Modifications of bases allow to improve PNA properties or to introduce new functionalities. In order to improve stability of the complex is possible, for example, to use bases with larger π -surface area, keeping H-bonding pattern.^{120,121}

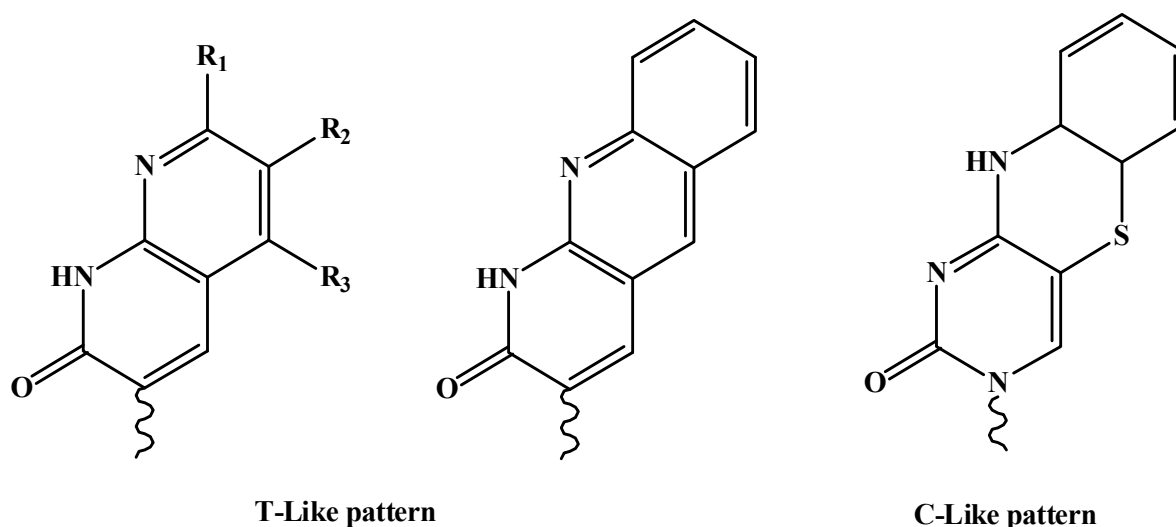


Figure 1.14: Examples of extended π -surface bases with thymine or cytosine bonding pattern.

Modified bases can be used also to insert reporter groups,^{122,123} allowing their use for analytical or diagnostic purposes. Particularly interesting are modifications that take advantage of Hoogsteen sites of nucleobases in addition to Watson-Crick interactions, thus increasing the number of H-bonds and therefore stability of the duplex. The so-called “G-clamp” bases (Figure 1.15), introduced by Matteucci and co-workers,¹²⁴ belong to this type and in particular are cytosine analogues with extra moieties able to “chelate” target guanine. Many systems have been reported in literature,¹²⁵ some examples are shown in Figure 1.15. It has been demonstrated that system **B** is able to form one additional H-bonds with guanine, while **C** is able to form two extra H-bonds.¹²⁶ A very effective system is **D**, which, if introduced in a PNA sequence, led to a stabilization of 12 °C compared to unmodified PNA.¹²⁷

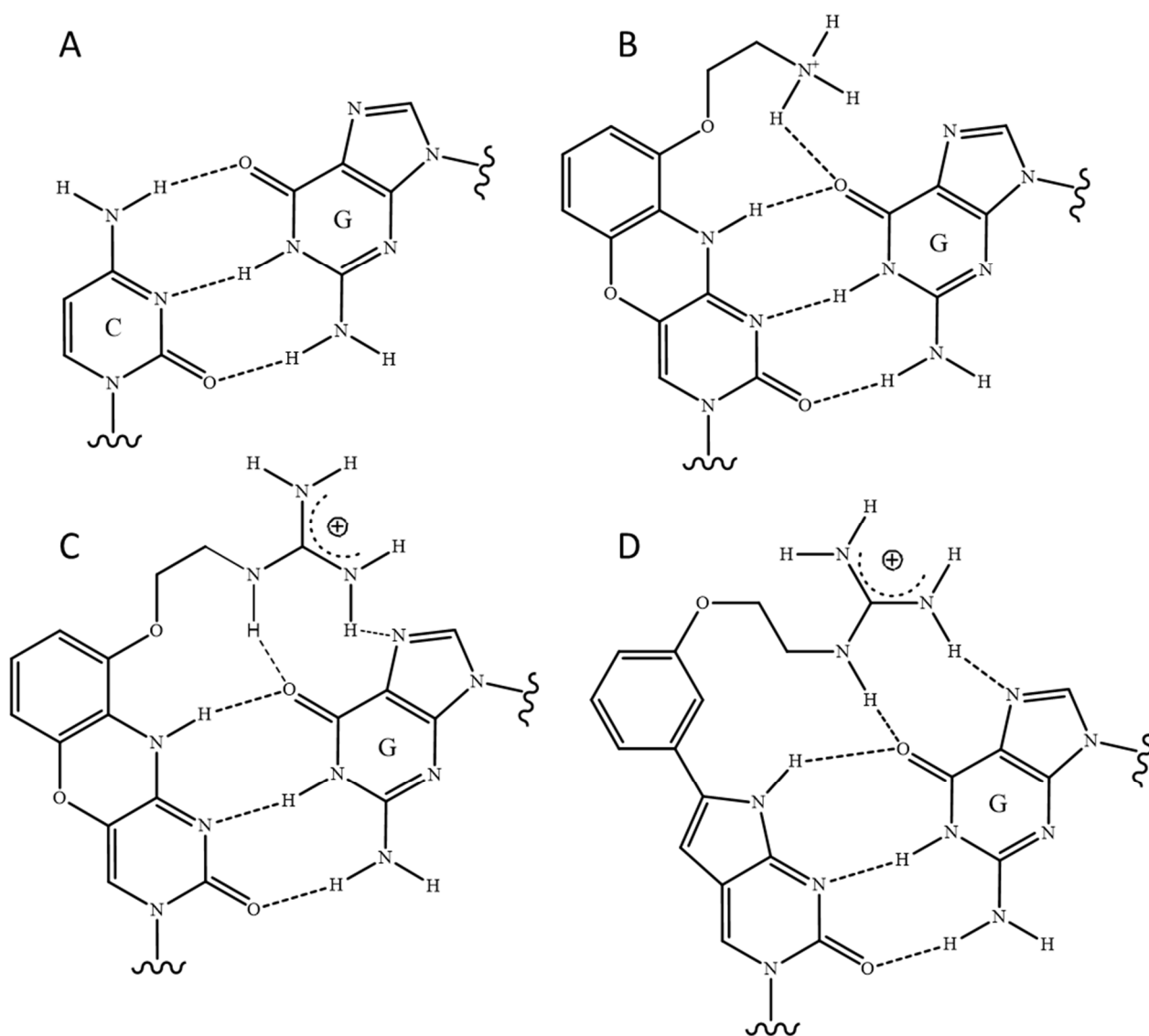


Figure 1.15: **A)** Standard cytosine-guanine base pairing; **B)** modified base presents an extended π -surface and an amino ethyl group able to interact with guanine; **C)** same base of B, but with guanidinium instead of amino group; this base is able to interact with Hoogsteen site; **D)** this base presents guanidinium unit like in case of C, but different polycyclic aromatic part.

Combination of modified backbone and base can lead to very effective systems and, for example, use of **C** with a γ -modified PNA allowed B-DNA double strand invasion.¹²⁸

Uracil is a well-studied system because substituents in 5- position don't alter duplex stability, since the side chains are placed in major groove of duplexes. Proof of this are many works, reported in the literature, in which polymerase enzymes incorporate modified nucleotides.^{129,130} For these reasons many modifications of uracil moiety have been proposed,¹³¹ bearing in position 5 reporter groups, metallic centers, reactive groups, etc.. Some examples are reported in Figure 1.16.

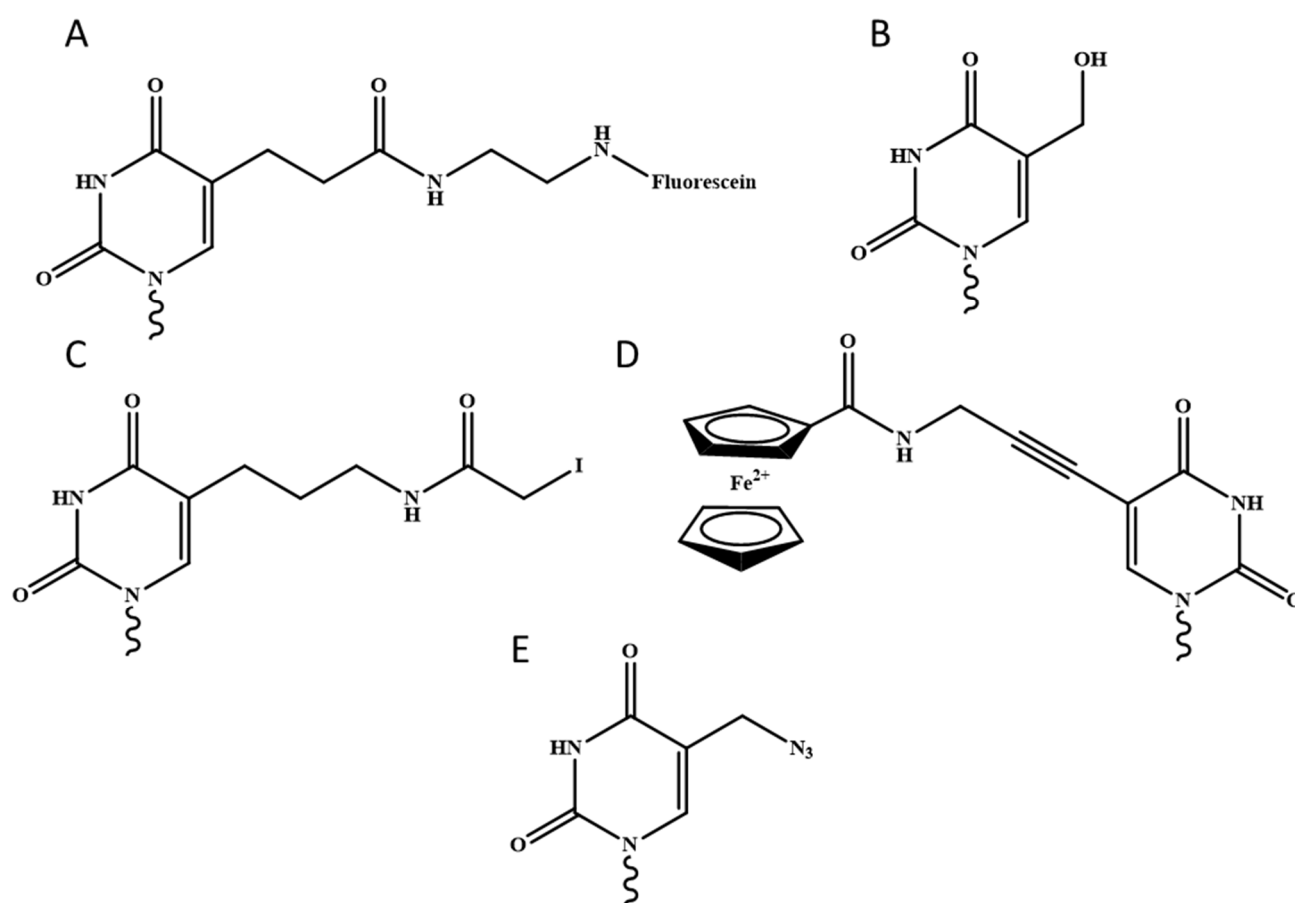


Figure 1.16: Examples of 5 modified uracil. **A)** uracil modified with reporter group; **B)** 5-hydroxymethyluracil **C)** uracil-based cross-linking agent; **D)** uracil coupled with ferrocene unit; **E)** 5-azidomethyluracil.

A is an example of system bearing a reporter group,¹³² coupled through a linker, useful for diagnostic or analytical purposes. **B** presents a modification that increases uracil hydrophilicity, favoring PNA solubility.¹³³ **C** is an example of base able to make cross linking with DNA and in particular it makes a covalent bond only with guanine.¹³⁴ Sonogashira's reaction¹³⁴ has been used to create a wide range of modified monomers starting from 5-iodouracil and this type of reactivity has been used also for the introduction of metal centers (**D**).¹³⁵ Finally **E** is a versatile monomer that allows to introduce a wide range of groups by means of click reactions or coupling after azide reduction.¹³⁶

1.5. Design of new modified PNA

A widely used approach for design of new modified PNA is that based on rational design combined with “trial and error” method. Availability of crystal structures containing PNA^{93,137–143} allowed to better focus on the conformational properties and to design modified PNA monomers more accurately. Several review papers report a complete overview of the backbone^{110,144} and of the nucleobase-modified PNA structures¹⁰⁸ proposed in the literature. Examples, relevant for the thesis work will be discussed in detail.

An interesting case of modifications derived from crystal analysis was reported by Ganesh and co workers.^{145,146} Standard aminoethylglycine (aeg)-PNA is conformationally flexible and can adapt to target DNA or RNA preferred helical conformation. A drawback of this adaptability is the high negative entropic change in duplex formation. To avoid this problem, many constrained systems have been proposed. From the observation of characteristics angles of PNA (Figure 1.17) in PNA:DNA and PNA:RNA duplex structures, changes in the β angle going from DNA to RNA heteroduplex and from duplex to triplex were identified. In particular in PNA:DNA duplex β is about 140° while in PNA:RNA or PNA₂RNA range from 65 to 70° .

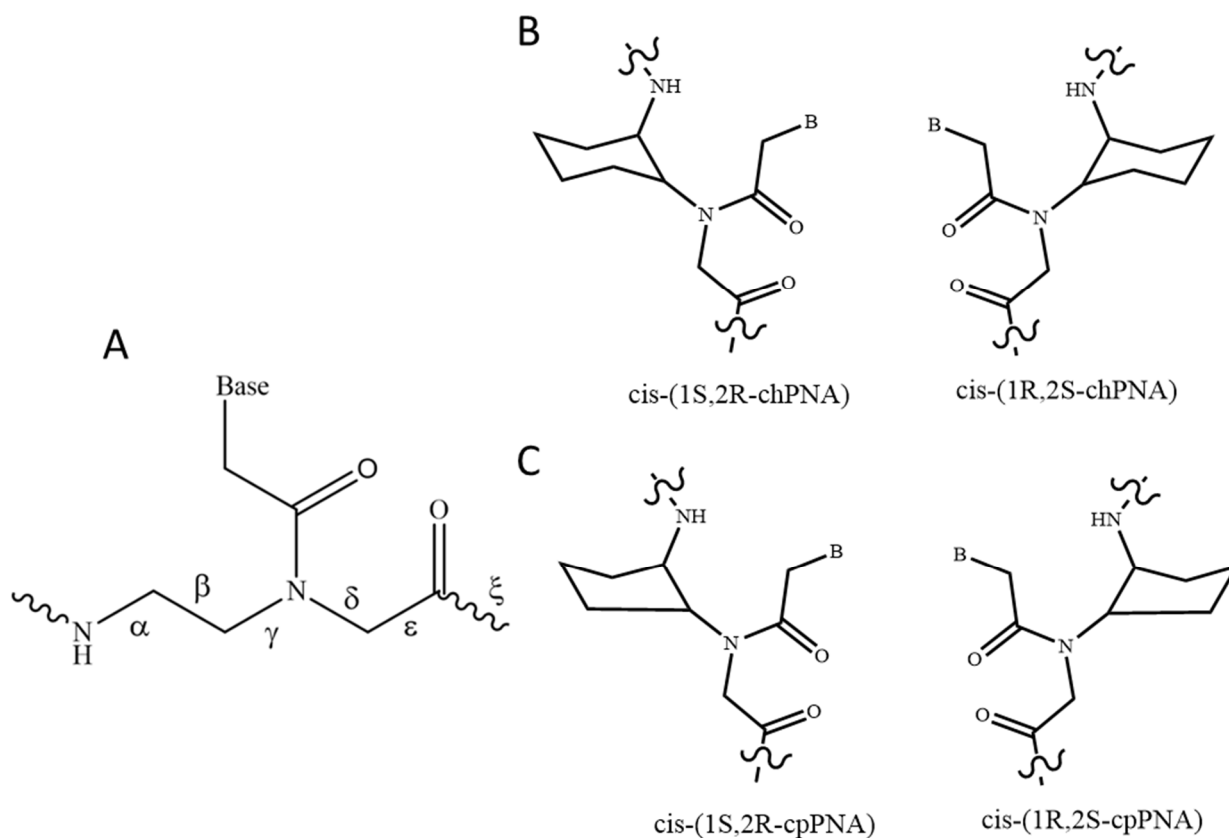


Figure 1.17: A) PNA characteristic angles; B) ch-PNAs structures; C) cp-PNA structures.

From this observation cyclohexanyl PNAs (chPNAs) were designed, by introducing an aminocyclohexyl unit instead of the standard aminoethyl one. This modification reduced backbone mobility and thus conformational freedom of the duplex. Two different systems, cis-(a,e)(1S,2R/1R,2S) were considered; the former presented a beta angle equal to -63° , the latter equal to 66° . Study of stability revealed that ch-PNAs destabilized the formation of DNA:PNA duplex, while the PNA:RNA stability was not affected. In particular complexes with RS monomers resulted in a higher melting temperature compared to SR analogues, confirming the importance of the structural design based on β angle. Thus, although these systems were not particularly effective in binding, they allowed to discriminate between DNA and RNA, showing differences in melting temperatures up to 50°C . Poor binding was probably a consequence of the rigidity of cyclohexyl ring that remained trapped in either of the two chair conformations, limiting pairing during duplex formation. To resolve this drawback, Ganesh and coworkers proposed a cyclopentyl-modified backbone (cp-PNAs), in which endo-exo puckering allowed better adaptability. Effectively this led to an increased duplex stability for both DNA and RNA, but to the detriment of the selectivity shown in case of ch-PNAs.

Although this was an example of successful rational design of new backbone-modified PNA structures, this approach presents the drawback to use information obtained from crystal structures that are rigid models, which is only a partial set of all possible conformations present in solution, where most processes of interest occur. A new interesting approach, based on dynamic study of systems using Molecular Dynamics, will be shown in following chapters.

1.6. Design of anti-miR PNA: a challenge

Anti-miR PNAs have been used for modulating differentiation,¹¹⁸ arrest cellular proliferation^{147,148} and inducing apoptosis.¹⁴⁹ Detection of miRs in living cells has also been achieved by means of PNA probes.¹⁵⁰

In miR nomenclature, “seed region” refers to the part of miR that goes from 2nd to 7th nucleotide of sequence. It has been demonstrated that targeting this region is sufficient to inhibit the activity of the miR without the necessity to have full sequence complementarity.¹⁵¹ Binding short sequences, like micro-RNA, requires very effective systems and even more efficient ones if the objective is to target the seed region, leading to demand of new modified oligonucleotides or PNAs. The present

PhD thesis was developed in the context of a regional project, PNA NOVA, with the purpose of developing new effective PNAs for miR targeting.

PNAs used in anti-miR approach essentially act by reducing micro RNA availability in the cell through duplex formation, but thus requiring equimolecular quantity or an excess of PNA to observe an effect (usually micro molar scale). A possibility to overcome this problem is to insert catalytic moieties able to cleave RNA. This would permit PNA to bind RNA and, once duplex is formed, to cleave phosphodiester bond. Duplex with two RNA fragments is less stable than duplex with whole miR. In this way it would be possible for PNA to dissociate from the complex and then bind again RNA strand and to act in catalytic way (Figure 1.18) like a sequence selective artificial nuclease.^{152,153}

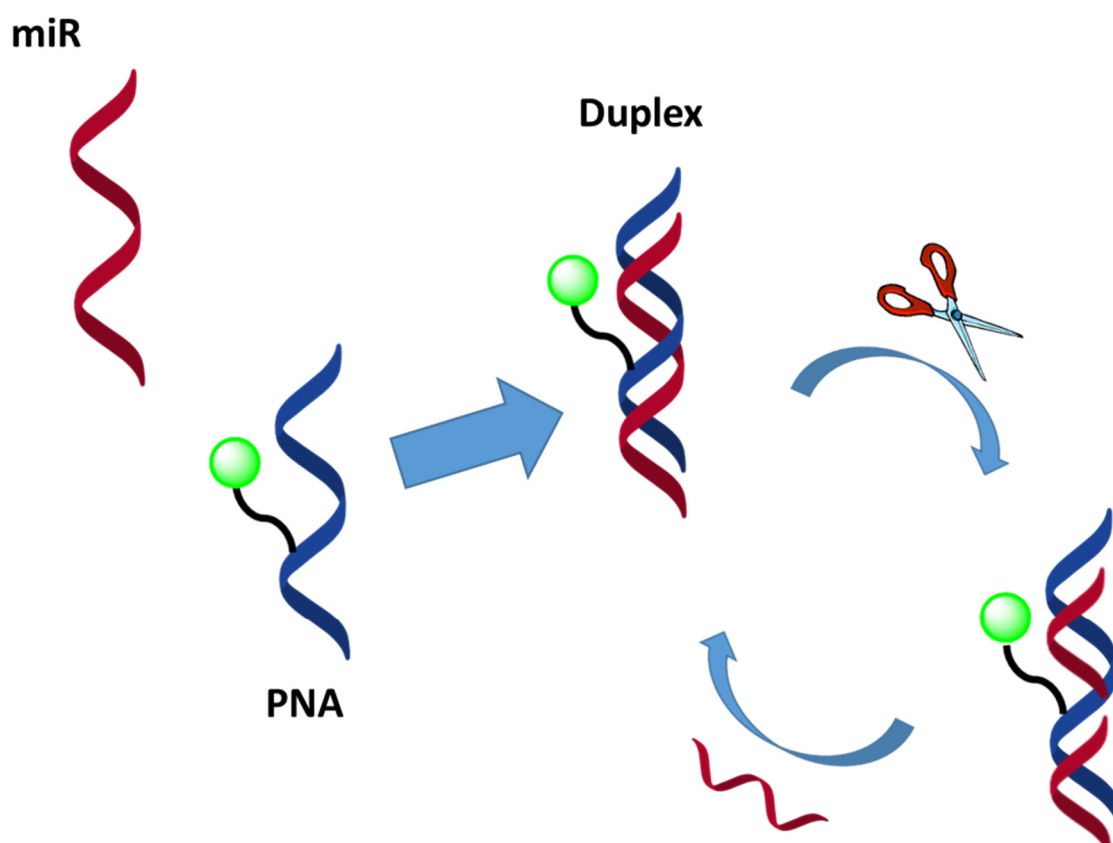


Figure 1.18: Schematic representation of activity of a PNA bearing a hydrolytic moiety.

A useful chemical toolbox for achieving this goal is provided by the extensive studies carried out in the field of artificial nucleases, and in particular of metallo-nucleases.

1.7. Artificial Metallonucleases

DNA must preserve genetic information and to ensure that its phosphodiester bond is quite resistant to hydrolysis. Actually, estimated half-life of DNA at 25 °C and pH 6.8 is of 30 millions of years.¹⁵⁴ RNA, which presents the role of transporting information, is more unstable, due to the possibility of intramolecular catalyzed hydrolysis and under the same conditions exhibits a half-life of “only” four years.¹⁵⁵ However, in biological media DNA and RNA can be rapidly degraded due to the presence of nucleases. These enzymes are able to greatly accelerate the hydrolysis reaction. Many types of enzyme can be found, but generally, their activity is ensured by presence of a metal center.

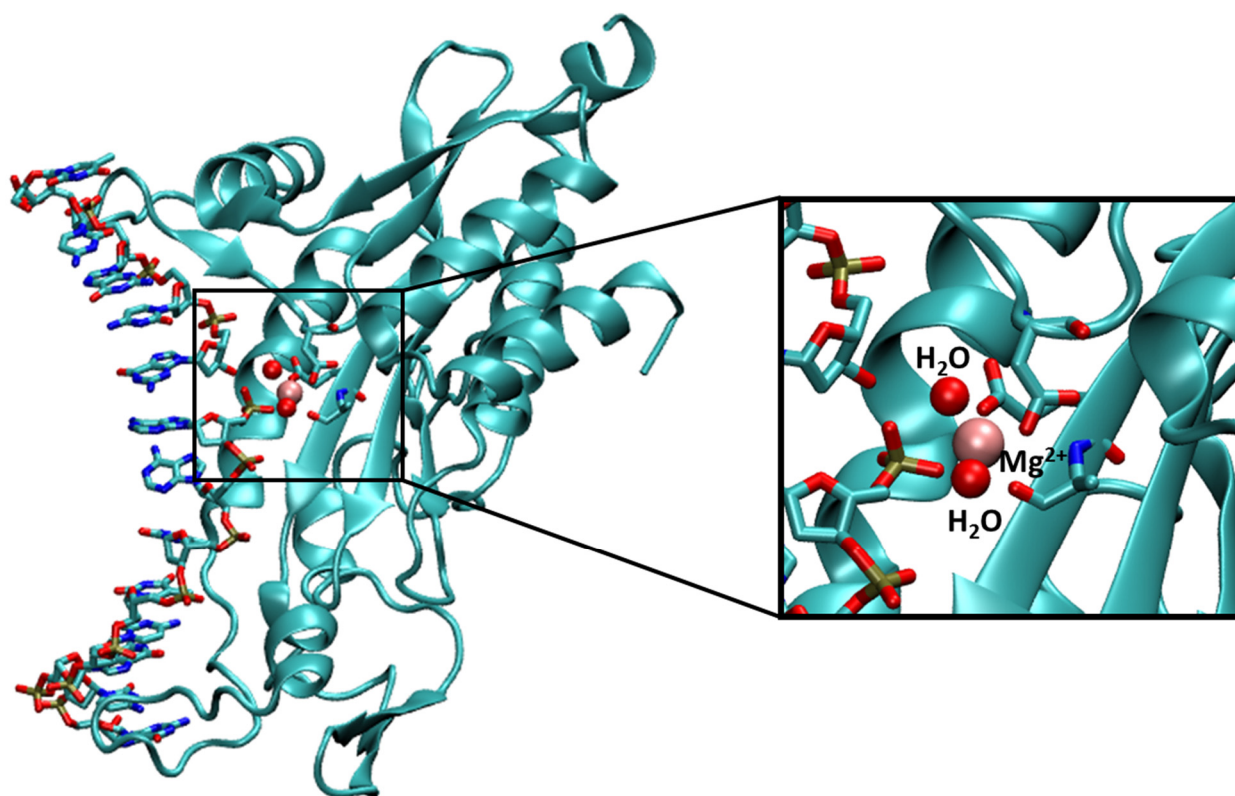


Figure 1.19: Crystal structure of EcoRI complex with DNA taken from PDB (1QPS)¹⁵⁶ and insight of catalytic center showing the role of the magnesium ion.

In Figure 1.19 an example of structure of a nuclease isolated from *Escherichia coli* is shown. Enzyme is able to bind DNA strand and to hydrolyze it through water molecules coordinated to Mg²⁺. Many other examples of nucleases can be found in literature^{157–159} and Mg, Mn, Fe and Zn are commonly

found in this type of enzymes. Several efforts have been spent trying to mimicking nucleases activity. There are two principal cleavage methods: oxidative and nucleophilic.

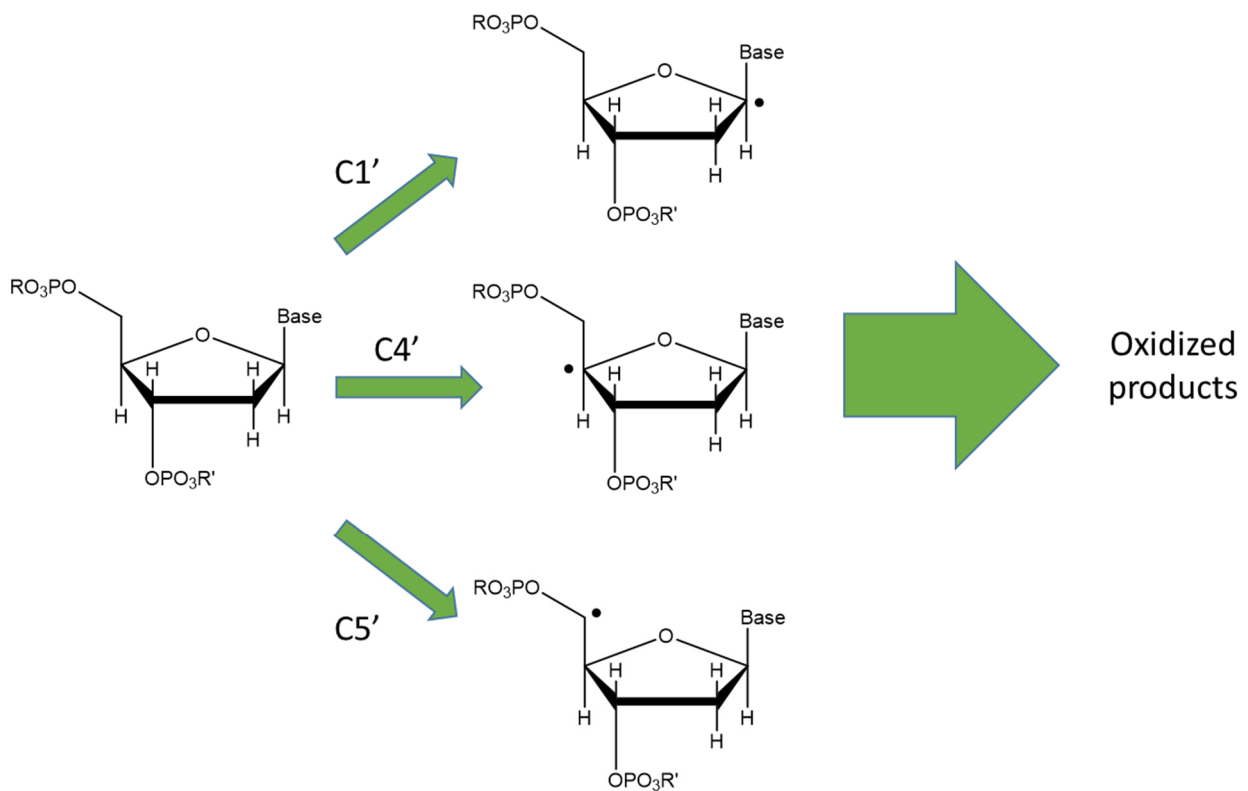


Figure 1.20: Schematic representation of oxidative cleavage pathway by hydrogen abstraction.

Oxidative cleavage (Figure 1.20) proceeds via different pathways, of which, the most effective is through hydrogen abstraction, that can occur in different positions, mainly in C1', C4' and C5'. After radical formation, DNA can be oxidized to obtain different products.¹⁶⁰

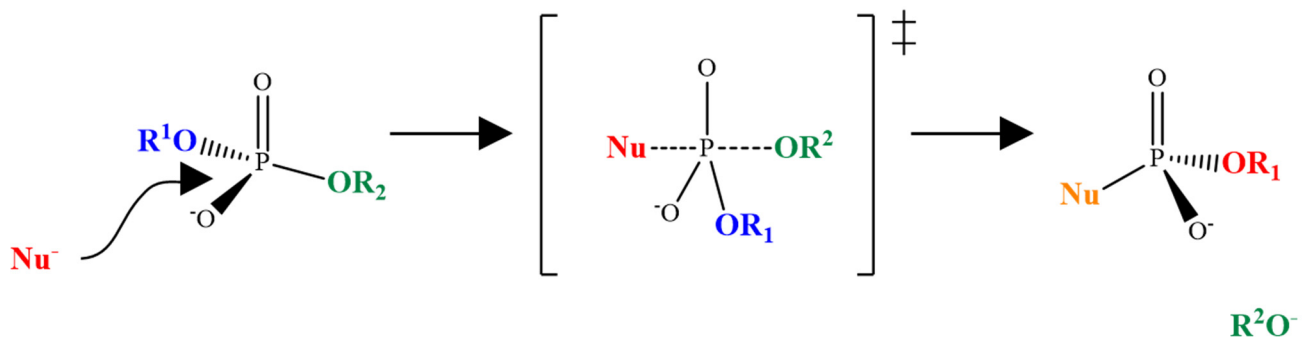


Figure 1.21: Schematic representation of hydrolytic cleavage pathway.

At contrary, nucleophilic cleavage proceeds through attack of a nucleophile on phosphorus, followed by scission of phosphodiester via pentacoordinate transition state (Figure 1.21). Nucleases acts following this pathway and nucleophile is usually a coordinated water molecule or a side chain of an amino acid residue. Many examples of metallic complexes, exhibiting hydrolytic effects toward DNA or RNA, have been reported in the literature.¹⁶¹ This type of compounds are called artificial metallonucleases. Most effective complexes contained cobalt,¹⁶² nickel¹⁶³ and cerium. In particular complex of Ce(IV)/EDTA^{164,165} resulted very effective in DNA single strand hydrolysis and have also been used in combination with PNAs:¹⁶⁶ PNA conjugated with NLS peptide was used to perform DNA double strand invasion and at that point was possible hydrolysis by Ce(IV)-EDTA complex. Drawback of using these metals is their cytotoxicity if released from ligands. Iron would be a perfect candidate, since is not toxic, however complexes with Fe(II) or Fe(III) resulted scarcely active or needed basic conditions to perform cleavage.¹⁶⁷ Interest of scientific community has been focused toward two alternatives: copper and zinc. Toxicity is higher compared to iron, but limited metal release is tolerated by biological systems, in particular in case of zinc. Complexes with these metals showed a sufficient activity due to coordinated water molecules with pKa within the range of 7-9. Many examples of complexes with this metals have been tested as nucleases.¹⁶⁸ Artificial nucleases can be mononuclear complexes,¹⁶⁹⁻¹⁷¹ with only one metal center, or multinuclear complexes (Figure 1.22).^{172,173} Generally complexes with copper were more active than those with zinc, and di- or multi-nuclear complexes exhibited better properties than mononuclear counterparts. Molecules bearing pendant nucleophiles have been synthesized (Figure 1.22): this groups, usually alcohols, react in place of coordinated water molecule with a higher hydrolysis rate, but intermediate phosphate is stable and it is not possible to achieve turnover.^{174,175}

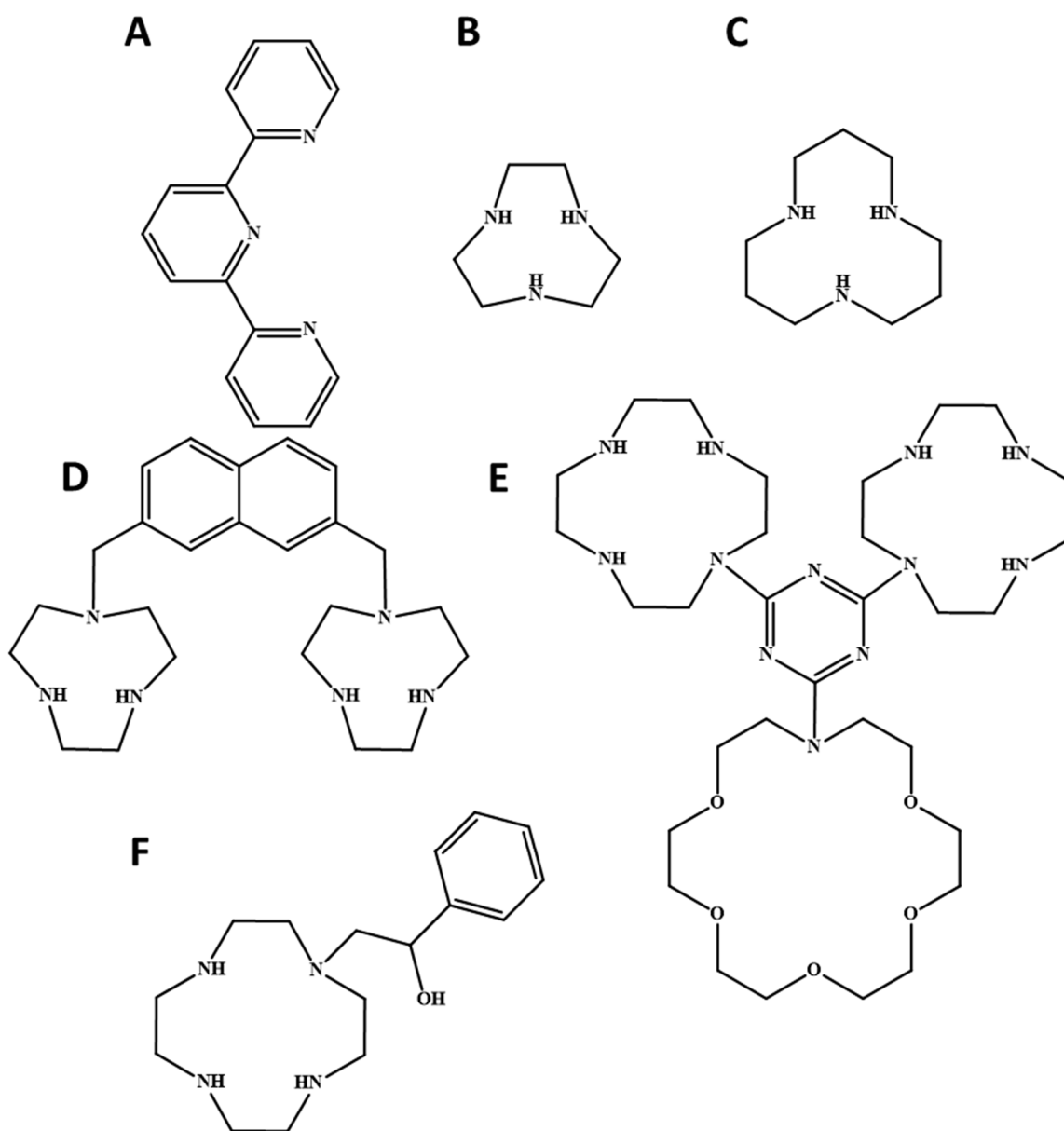


Figure 1.22: Examples of mononuclear and multinuclear metal ligands used as artificial metallonucleases. **A)** terpyridine; **B)** 1,4,7-triazacyclononane; **C)** 1,5,9-triazacyclododecane; **D)** dinuclear triazacyclononane derivative; **E)** trinuclear 1,4,7,10-tetraazacyclododecane derivative; **F)** 1,4,7,10-tetraazacyclododecane bearing a pedant alcohol.

Scrimin and coworkers reported an interesting example of polypeptide bearing two Zn^{2+} triazacyclononane hydrolytic units.¹⁷⁶ Folding of peptide is necessary to bring two catalytic rings sufficiently close together in order to enhance phosphodiester bond (Figure 1.23). Another artificial nuclease was developed by the same group using gold nanoparticles functionalized with hydrolytic moieties.¹⁷⁷ On particle surface, these units are sufficiently close to act cooperatively increasing hydrolysis rate.

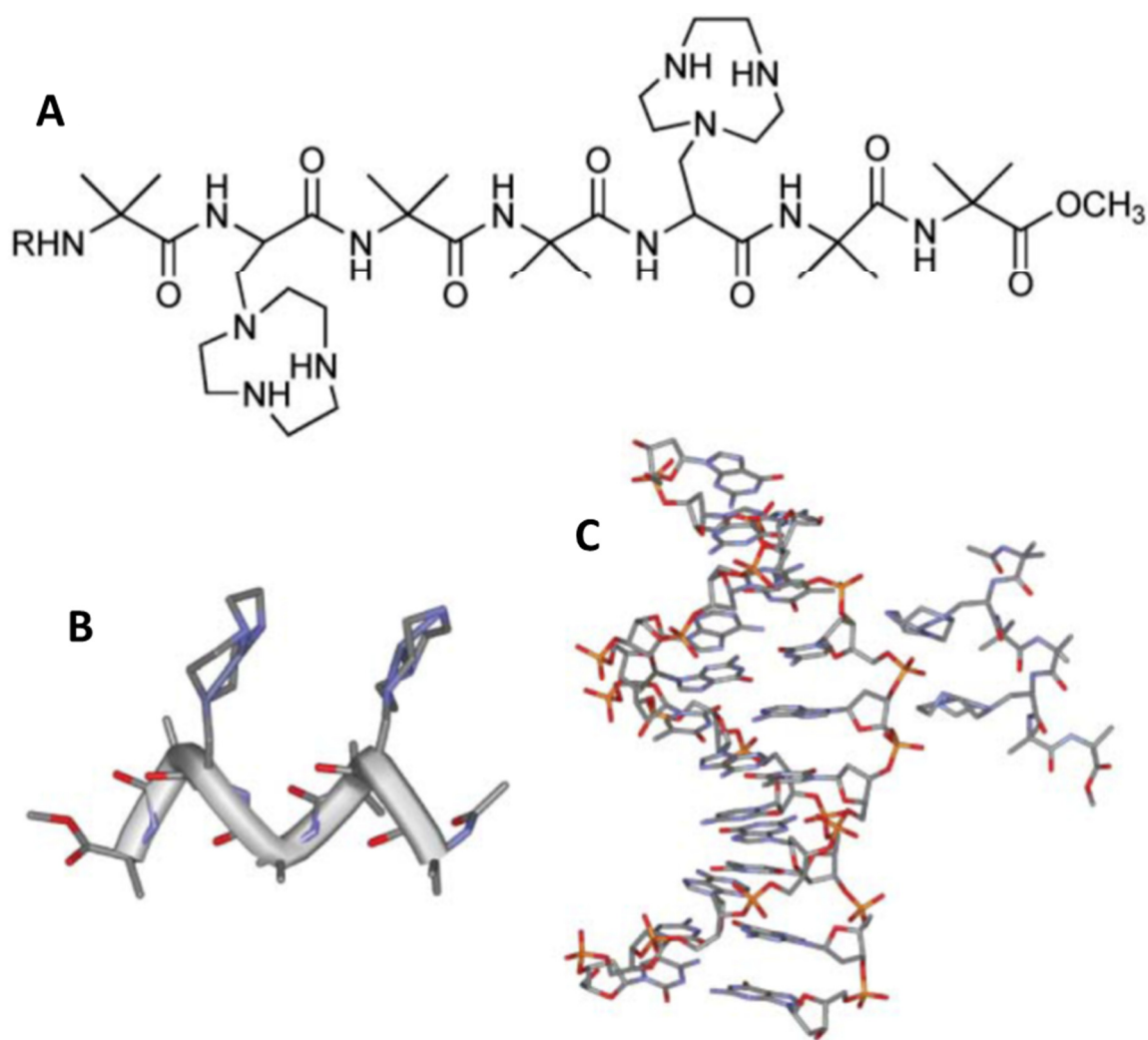


Figure 1.23: Modified Peptide bearing two hydrolytic units. **A)** Chemical structure; **B)** secondary structure; **C)** proposed mode of binding to DNA. Reprinted with the permission of Royal Society of Chemistry.¹⁷⁸

During PhD thesis synthesis of a mononuclear ligand was performed in order to insert a hydrolytic unit in PNA strand. Discussion of PNA-based artificial nuclease will be reported in more details in Chapter 6.

2. Methodology background: Molecular Dynamics and Metadynamics

2.1. Simulations of large molecular systems

Theoretical basis for simulation of molecular properties have been developed since the very beginning of computer era.¹⁷⁹ However, only in recent years exponential growth of computational power permitted to widely expand the use of simulations to solve complex problems. *Ab initio* methods describe the motion of electrons in atoms and molecules using wave functions and avoiding any experimental data. In this way, it is possible to compute quantum mechanical properties of molecular systems at a high level of accuracy, but lacking in efficiency due to the usually high computational costs. For this reason, *ab initio* simulations are generally restricted to small molecules/systems with less than 100-1000 atoms. However, medicine, biology and biotechnology challenges require tools suitable for much bigger and complex systems such as proteins, enzymes, receptors, membranes, etc. with hundreds of thousands of atoms. Classical Molecular Dynamics (MD) is an alternative approach which can be used to address this type of problems; in MD atoms are described as spheres, represented by their Van der Waals radii, connected with rigid bonds and subjected to classical physics laws. This approach simplifies several calculation steps avoiding the resolution of large Hamiltonian matrixes as it happens in *ab initio* MD, thus neglecting quantum mechanical effects. MD techniques resulted in a successful approach for simulating large molecular systems, allowing to achieve precious insights in the molecular structures and motions. Many years have passed from the first simulation of 9 picoseconds, made by Karplus on bovine pancreatic trypsin inhibitor (BPTI),¹⁸⁰ and nowadays it is possible to reach simulation times ranging from tens to thousands of nanoseconds, thus approaching reliable simulated measurable properties. In this chapter we will describe the methodologies of the work carried out during a training period on computational tools spent at the Department of Chemistry and Applied Biosciences ETH Zurich, USI Campus, Lugano, in the research group of Prof. Michele Parrinello, one of the pioneers and most prominent scientist in the field of Molecular Dynamics of large systems, including Biosystems, under the supervision of Dr. Vincenzo Verdolino.

2.2. Introduction to Molecular Dynamics

Use of classic physic principles, neglecting quantum effects, allows to simplify energy calculation that is represented by the sum of kinetic (K) and potential (V) energy (2.1):

$$E(\mathbf{r}, \mathbf{p}) = K(\mathbf{p}) + V(\mathbf{r}) \quad (2.1)$$

where \mathbf{r} and \mathbf{p} are the coordinate and momentum vectors, respectively. Kinetic energy has usually a very simple form (2.2) easy to calculate, while potential is a more complex function.

$$K = \sum_{i=1}^N \frac{1}{2m_i} (p_{ix}^2 + p_{iy}^2 + p_{iz}^2) \quad (2.2)$$

where m_i is the mass of i^{th} particle and p_{ix} , p_{iy} , p_{iz} are the vector components of \mathbf{p}_i .

In MD there are different way to express the potential $V(\mathbf{r})$, but in any case it comes from the sum of different terms, defined as bonded (V_{bond} , V_{angle} , V_{dihedral}) and non-bonded (V_{improper} , V_{vdw} , V_{elec}).¹⁸¹

$$V = V_{\text{bond}} + V_{\text{angle}} + V_{\text{dihedral}} + V_{\text{improper}} + V_{\text{vdw}} + V_{\text{elec}} \quad (2.3)$$

$$V_{\text{bond}} = K_b(b - b_0)^2 \quad (2.4)$$

$$V_{\text{angle}} = K_\theta(\theta - \theta_0)^2 \quad (2.5)$$

$$V_{\text{dihedral}} = K_\chi[1 + \cos(n\chi - \delta)] \quad (2.6)$$

$$V_{\text{improper}} = K_\psi(\psi - \psi_0)^2 \quad (2.7)$$

$$V_{\text{vdw}} = \epsilon \left[\left(\frac{R_{ij}}{r_{ij}} \right)^{12} - 2 \left(\frac{R_{ij}}{r_{ij}} \right)^6 \right] \quad (2.8)$$

$$V_{\text{elec}} = \frac{q_i q_j}{4\pi\epsilon_0 r_{ij}} \quad (2.9)$$

where K_b is the bond force constant and $b - b_0$ is the distance from equilibrium position. K_θ is the angle force constant and $\theta - \theta_0$ is the angle from equilibrium between 3 bonded atoms. K_χ is the dihedral force constant, n is the multiplicity of the function, χ is the dihedral angle and δ is the phase shift. K_ψ is the force constant and $\psi - \psi_0$ is the out of plane angle. ϵ is the depth of the potential well, R_{ij} is the distance in which potential reach the minimum, and r_{ij} the distance between atoms. q_i and q_j are atoms charges, ϵ_0 is the dielectric constant and r_{ij} is the same distance of V_{waw} . Bonds, angles and impropers terms are expressed in the form of harmonic potential (2.4, 2.5, 2.7). Usually spring constants assume quite high value, thus inducing a great energy variation for little displacements from equilibrium position. Dihedral potentials are conversely softer, allowing broader ranges and assuring necessary flexibility to the molecule (2.6). Electrostatic contribute is expressed in the familiar Coulomb equation (2.8) and dispersive interactions (Van der Waals) are regulated by Lennard-Jones potential (2.9). These two terms constitute long-range interactions and are the most expensive in terms of computational effort. In order to accelerate MD calculations a cutoff value for VdW interactions is usually set to 10 Å since this contribution decays as r^{-6} and at long distances is almost zero. For Coulombic potential is not possible to adopt the same cutoff since it decays as r^{-1} and the error generated would be fairly high. A common technique to this problem is the use of Particle Mesh Ewald (PME) method that consists in introducing a neutralizing charge distribution for every point charge of the molecule and subtracting this contribution to the total electrostatic potential.¹⁸² This term decays faster and is then possible to apply a cutoff similar to that one used for dispersive forces. Use of PME protocol requires also application of periodic boundary conditions (PBC)¹⁸³ that means molecular systems replicated in the three spatial directions, forming an infinite *lattice*. This approach ensures that all particles leaving the simulation box from any side are re-entered from its periodic image (Figure 2.1). PBC box allows also to avoid surfaces effects¹⁸¹ due to finite size systems and is therefore commonly used in Molecular Dynamics. All potential contribution described above must be defined for every molecule to simulate. Different simulation programs have been developed together with their own Force Field (FF) syntax and parameters attribution. Most used programs are GROMACS,¹⁸⁵ AMBER,¹⁸⁶ CHARMM¹⁸⁷ and NAMD.¹⁸⁸ Moreover, for common types of molecules and scaffolds, many different Force Fields are available for users avoiding manual parameter definition. In early versions, each different program used its parameters, leading to development of large variety of Force Fields for the same type of molecules. Nowadays, most of the programs share the code giving to users the possibility to choose; for example GROMACS allows to select its proper FFs (GROMOS)¹⁸⁹ or among FFs developed for

other programs (AMBER,¹⁹⁰ CHARMM,¹⁹¹ OPLS¹⁹²). Since proteins are the most studied systems, in the past years Force Fields in this area were greatly developed.^{193–195}

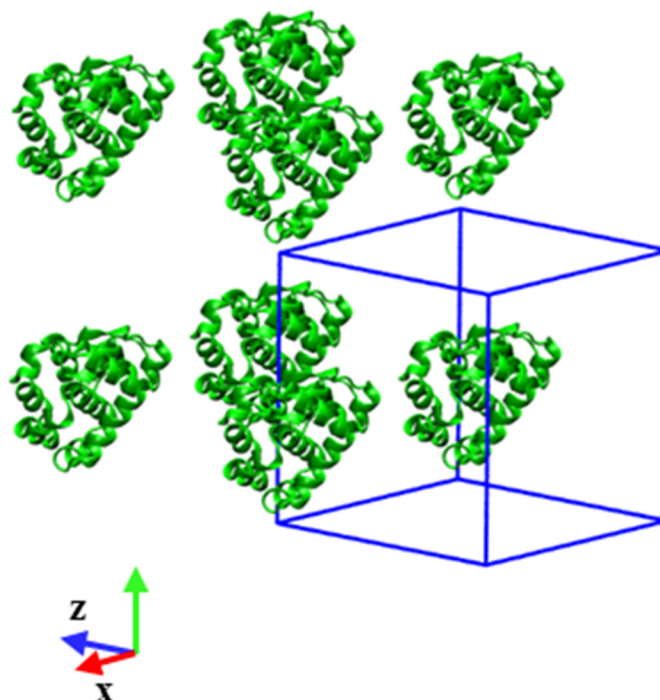


Figure 2.1: Representation of Periodic Boundary Conditions: simulation box is represented in blue (60.0 X 60 X 60 Å³), original protein structure (3HTB)¹⁸⁴ is inside the box and outside +x, +y, +z, replicates are shown.

Nowadays, FFs for organic small molecules start to be expanded for including lipids,¹⁹⁶ sugars,¹⁹⁷ nucleic acids,¹⁹⁸ etc. The choice of the best performing FF is not trivial and usually is based on the experience of the user and on the literature. Compared to proteins, force fields for nucleic acids are barely explored and continuous improvements are in the process. Most recently used FFs for DNA and RNA are Amber *ff99sb*¹⁹⁹ improved with *parmbsc0*,²⁰⁰ CHARMM27,^{191,201} GROMOS53a6¹⁸⁹ and OPLS-AA,²⁰² but the first two gave the best results in reproducing the experimental data.^{198,203,204} Generalized Amber Force Field (*GAFF*)²⁰⁵ and CHARMM General Force Field (*CGenFF*)²⁰⁶ are also popular FFs since they provide parameters for small organic molecules that can be applied to ligands or modifications.

Once the best choice of the FF is set along with all the simulation parameters, it is possible to start the time evolution of the molecular systems. This will follow trajectories that satisfy the solution of Newtonian motion equations under the field of forces selected. This achievement bare mechanistic, structural, thermodynamics and kinetic features that can be post processed. In particular, it is of great interest the potential or the Free Energy Surface (FES) describing the system under study. One effective approach is to define variables or combination of variables called Collective Variables (CV) that based on the experience or the physics of the system can optimally describe one or more

motions of interest. Moreover, the selected CVs have to univocally describe the motions under examination in order to better discriminate between different system states.

Another critical aspect of MD simulations is the conformational space exploration. In order to extract Free Energy Surface information out of the simulated trajectories, it is fundamental that the system under study spent a significant amount of time in the conformation of interest. This criterion that could appear as an arbitrary estimation can be quantitatively evaluated through the free energy surface (FES) convergence analysis. Indeed, if the simulation time is sufficiently long, the molecular system will have reached the equilibrium state. With this, the probability for the system to be in a given configuration is calculated by means of the time spent in that conformation and, most remarkably will be converged onto the free energy surface. Employing the collective variables formalism the FES can be expressed in terms of CV-dependent probabilities through the following equation (2.10):

$$\mathbf{F}(\mathbf{s}) = -k_B T \ln(\mathbf{P}(\mathbf{s})) \quad (2.10)$$

where \mathbf{F} is the free energy, \mathbf{P} is probability function of \mathbf{s} , k_B is Boltzmann constant and T temperature.

However, the equilibrium condition is difficult to practically satisfy and it would require infinite simulation time. With finite simulations, such approximation is particularly dangerous when considering systems with high degrees of freedom or for those where state transitions are relatively slow. The latter situation is common, for example, in proteins²⁰⁷⁻²⁰⁹ in which passage from two different states may imply huge conformational rearrangements. In both cases, the restricted conformational sampling allowed by classical MD can bring to erroneous conclusions. To overcome to these problems, enhanced sampling methods²¹⁰⁻²¹³ must be used. In particular, hereafter we describe the Metadynamics methodology.^{214,215}

2.3. Principles of Metadynamics

Metadynamics has been defined as “filling the free energy wells with computational sand”.²¹⁶

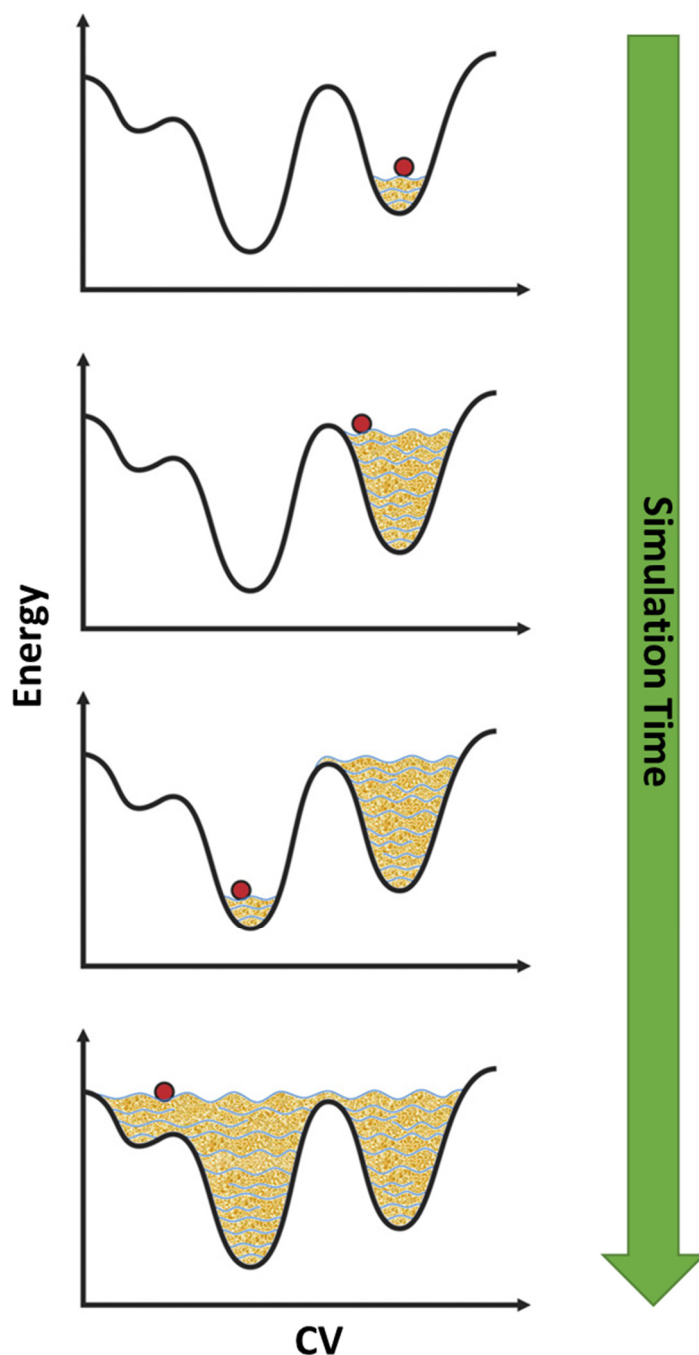


Figure 2.2: Graphical representation of Metadynamics.²¹⁷ Conformational space is expressed as function of a generic Collective Variable, which describes the system, and energy. Position of the system in this space is represented with a red circle. Applied potential is here represented as sand, which accumulates over time.

This statement simplistically represents the operative procedure employed by the Metadynamics in which simulated system is discouraged to visit previously populated states by means of a history-

dependent bias potential.²¹⁴ Figure 2.2 describes an example of energy profile as function of a generic CV in which there are two local minima and a global minimum. With a red circle we identify the *position* of the system under study at a given simulation time. We define this *position* in terms of Collective Variables that optimally describe the system conformational space. In this example, the starting position of the system is set in a local minimum. In order to escape from this basin a significantly tall energy barrier has to be crossed. The probability of populating other minima on the Free Energy Surface depends on the thermal contribution defined by the energy $k_B T$. Usually, standard MD at room temperature rarely allows for overcoming such tall energy barriers and the system will permanently oscillate in the starting basin only for temperature-induced fluctuation. Metadynamics adds, at regular user-defined intervals, a bias potential in terms of so called *hills* that help the system to populate higher energy configurations and eventually to fall in other local minima. Through this procedure, hills accumulate during the simulation “flooding” the free energy surface and allowing all possible minima in the conformational space. The time-dependent potential assumes the form expressed in (2.11).

$$V(\mathbf{s}(\mathbf{t})) = \sum_{\mathbf{t}' \leq \mathbf{t}} \mathbf{w} e^{\left(-\sum \frac{(\mathbf{s}_i(\mathbf{t}) - \mathbf{s}_{i,\mathbf{t}'})^2}{2\sigma_i^2} \right)} \quad (2.11)$$

where \mathbf{w} is the height and σ the width of a single hill, $\mathbf{s}(\mathbf{t})$ is a vector of CVs at time \mathbf{t} and $\mathbf{s}_{i,\mathbf{t}'}$ is the bias potential deposited by the hills at $\mathbf{t}' < \mathbf{t}$, keeping the “bias-history” of the simulation. Recovering the visited states, it is thus possible to reconstruct the FES as the sum of the bias potential (2.12):

$$\mathbf{F}(\mathbf{s}) = - \sum_{\mathbf{t}' \leq \mathbf{t}} \mathbf{w} e^{\left(-\sum \frac{(\mathbf{s}_i(\mathbf{t}) - \mathbf{s}_{i,\mathbf{t}'})^2}{2\sigma_i^2} \right)} \quad (2.12)$$

Reliable FES reconstruction requires an accurate choice of the collective variables. CVs should satisfy the following four characteristics to be used as system descriptors.²¹⁸

- First of all, CVs must be simple or complex functions of atomic coordinates derived from trajectories collected during MD simulations ;

- Second, the function $s(r)$ has to be continuous in the entire configurational space sampled. This condition is partially satisfied by the existence of $ds(r)/dr$ which are the forces acting on each atoms at a given time and responsible for atomic motions. Failure in satisfaction of this condition causes crash of simulation;

These two conditions are of general validity and requested for all kind of MD simulations.

- Third feature: any efficient CV has to univocally discriminate the different states characterizing the system under study. Ideally, at a given value or combinations of CVs it one single system state or molecular structures within a certain confidential interval should be defined. This is a quite arbitrary evaluation that often gives rise to erroneous conclusions. In particular, for complex CV definitions, this condition can hardly be achieved and dedicated discussion should be derived. Nonetheless, if such condition is not satisfied, the statistics collected during the MD run can still be post processed by means of reweighting techniques that allows recovering the unbiased probability distribution of any variable from Metadynamics simulation.²¹⁹
- Fourth, CVs should represent the “slow motion coordinates” of the system. In this way, “fast coordinates” follow a standard unbiased molecular dynamics whereas the time demanding motions will be facilitated by the Metadynamics. For example, if we consider the ligand binding to a protein it might be the case where the hot-site under study becomes available upon slow conformational rearrangement involving opening and closing of peptides functionalities. If that is the case, in order to accurately evaluate the binding energy, it is necessary to collect a sufficiently large number of statistics involving both processes (i.e. pocket opening/closing). Being this process the rate determining of the simulation it turns out that the optimal CVs have to be evaluated accordingly.

2.4. Well-Tempered Metadynamics

Classical Metadynamics employs constant height of the hills w along the whole simulation. This condition usually brings to overestimate the free energies and leaves to the user experience an arbitrary criterion for the statistic accumulation interruption. In particular, high hills allow to rapidly flood the energy surface, accelerating the MD, but resulting in rough FES estimation. On the contrary, low hills lead to accurate FESs requiring long simulation time. Another problem is that once a minimum is filled, the applied bias remains fixed, causing unrealistic sampling and in some cases preventing the observation of slow processes. An improvement of Metadynamics is the Well-Tempered Metadynamics (WT-MTMD).²²⁰ This technique consists in applying variable height of the hills regarding the conformational space already visited. This algorithm, ensures that the hills deposition at a given value of the CVs is history dependent allowing for initial rapid filling of the local minimum and decreasing after several exploration of the same state. In this scheme, the molecular system will eventually explore all the free energy landscape avoiding a local overestimation of the energy barriers involved. Indeed, it is possible to demonstrate that Well-Tempered Metadynamics converges to an exact FES.²²¹

The height of the hills is expressed in its time dependent formulation by the equation (2.13).

$$w = \omega e^{-\left[\frac{V_b(s,t)}{\Delta T}\right]} \tau_G \quad (2.13)$$

where ω is the bias deposition rate, $V_b(s,t)$ is the potential at the current value of the CV for a given time step, and τ_G is the deposition time step.²²²

Metadynamics has been successfully used to simulate complex systems such as protein,²²³ folding,²²⁴ flexible docking,^{225,226} nucleation²²⁷ and mechanism of reaction pathways.²²⁸ For this reason, we have selected this advanced technique for simulations of PNA-based systems.

2.5. Procedures for PNA simulations

All the MD simulations reported in this thesis were carried out with GROMACS 4.5.5,²²⁹ patched with PLUMED 1.3²³⁰ and PLUMED 2.0,²³¹ open source software, employed for the Metadynamics calculations.

Structure 176D,¹⁴³ taken from Protein Data Bank, was chosen as starting structure for the computational studies on unmodified PNAs (sections 3.2, 3.3, 3.5). For simulations where RNA moieties were present, we employed *ff99SB* force field improved with *parmbsc0*, since these force fields have been extensively used^{232,233} leading to reliable and reproducible results.²⁰³ PNAs are not parameterized in the available FFs and we retrieved missing parameters from R.E.DD.B. Server.^{234,235} Details on the parameters used are reported in Appendix 8.1.

γ -modified PNA structures (sections 3.2, 3.4) were generated by modification of structure 176D. A force field for gamma serine modified PNAs was not available in the literature and its derivation will be discussed in section 2.6.

For the study of PNA containing modified bases, it was necessary to generate duplexes with sequences of interest besides the modification. Manual modification of duplexes is time consuming and can lead to unreal structural deformations of the system. The Nucleic Acid Builder (NAB)²³⁶ module of AmberTools¹⁸⁶ is a module that allows to automatically generate DNA:DNA or RNA:RNA duplexes allowing to study of nucleic acids with desired sequence. To overcome problems described above we modified the code in order to automatically generate PNA:DNA and PNA:RNA duplex structures (see section 2.7). Charges and parameters derivation procedure are reported in section 2.8.

All systems were solvated in cubic boxes allowing at least 15 Å from the edges (i.e. 30 Å from the periodic boundary image) and filled by means of TIP3P water molecules. Topology and coordinate files were generated using t-leap module of AmberTools²³⁷ and converted to GROMACS input files using Acpye script.²³⁸ All the simulated systems, after a brief energy minimization procedure, were equilibrated at 300K (or differently where indicated) and 1 atm for 2 ns in the NPT (i.e. isothermal-isobaric) ensemble, employing the Berendsen barostat.²³⁹ The production runs were generated in the NVT (i.e. canonical: isochoric-isothermal) ensemble, by means of the v-rescale thermostat.²⁴⁰ The non-bonded cutoff scheme was set equal to 10 Å and the PME algorithm was employed for the electrostatics contribution.¹⁸² The simulations were performed with a 2.0 femtosecond time step,

constraining all the bonds with the SHAKE algorithm.²⁴¹ Metadynamics simulations were conducted under the same conditions, applying the opportune bias to the CVs. Details are reported in the next chapter.

2.6. Parameters for γ -Modified PNAs

Simulating gamma modified PNA requires determination of charges and parameters for the force field. Restrained electrostatic potential (RESP)²⁴² is recognized as one of the standard in order to assign atom charges for MD simulations. It consists in a penalty-based fitting approach of electrostatic potential (ESP) calculated with *ab initio* methods. Use of different basis sets for calculation may result in different charges values, but using 6-31G* or larger basis set it has been demonstrated that results converge.²⁴³ Therefore, this is the commonly used basis set because is a good compromise between accuracy and computational costs. Charges are generated fitting ESP by means of shell of points (with a density of 1 point/Å²) at 1.4, 1.6, 1.8, 2.0 times the Van der Waals radius of atoms. This charge derivation allows to obtain reliable charge values, sampling “buried atoms” accurately and with reduced dependency to conformation. Better results are nowadays obtained increasing number of shells and of points; in the present work we have used 10 shells for each atom and a density of 17 points/Å². Charge derivation was carried out using the RESP ESP charge Derive (R.E.D.) Server²⁴⁴ and following the protocol developed by Cornell *et al.*²⁴⁵ In particular, to build a complete residue library, for each base there are three types of fragments: N-terminus, central and C-terminus (Figure 2.3). The N-terminus is the first monomer of the sequence (i.e. with free amino group) and C-terminus is the last monomer of the sequence (i.e. with free carboxylic group); into PNA chain, a generic monomer **n** is attached to amino group of residue **n-1** and to carboxylic group of residue **n+1** (central fragments). In order to mimic the peptide linkage between residues and to obtain reliable charges, their N-end tail is acetylated (ACE) whereas the C-end is capped with methyl amide (NMA). Structures were optimized with Gaussian 09 D.01²⁴⁶ using DFT B3LYP level of theory and 6-31G* basis set. RESP charges were then calculated using R.E.D. program²⁴⁴ implemented in R.E.D. server²⁴⁷. Schematic representation of charge derivation procedure is represented in Figure 2.3. Concerning central fragments, RESP charges were obtained imposing neutrality condition (charge equal to 0) after capping groups removal (NMA and ACE). RESP charges on N-terminus were derived from capped monomers and methyl ammonium (NMA⁺) imposing total charge equal to +1 after capping groups removal and joining of **NH₃⁺** to the monomer.

Similarly, C-terminus is derived from capped monomer and acetate (ACE^-) imposing total charge of -1 after capping groups removal and joining of COO^- to the monomer. Moreover, charge fitting was performed imposing rotation of the bases along the linker. This last, homogenizes the fitted charge values as a function of base rotation, leading to more consistent and reliable RESP values.²³⁴ Finally, atoms of the backbone and of the linker were forced to have the same charges except for C8' and relative hydrogens, allowing charge redistribution with different bases bonded. This procedure, the same used for unmodified PNA, permitted to construct a complete and consistent library of gamma modified PNA monomers. Details about R.E.D. usage are reported in the program site.²⁴⁸ Structures and charges are reported in Appendix 8.2.

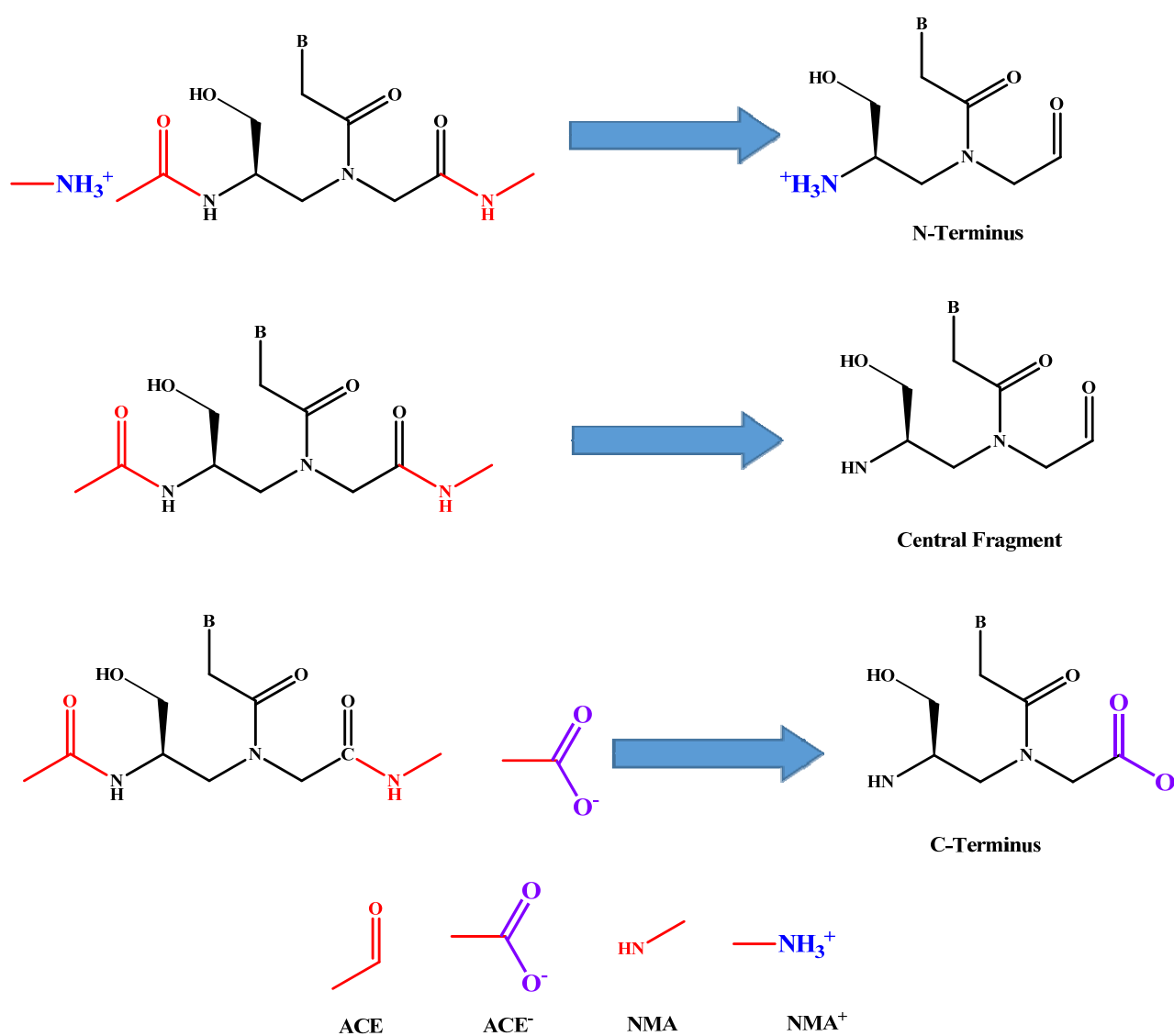


Figure 2.3: Schematic representation of RESP derivation procedure. In red are reported groups that are removed to obtain final fragments. In blue NH_3^+ that is joined to backbone to generate N fragment and in violet COO^- that is joined to backbone to obtain C terminus.

Since the structure of gamma PNA is analogue to the unmodified one and the serine side chain is already implemented in *ff99sb*, it was not necessary to introduce new Force Field parameters for the related simulations.

2.7. Modified NAB

Despite of growing interest toward PNA, one limiting issue in PNA chemistry is the scarce availability of solid-state structures due to the difficulties in obtaining crystals of the duplexes, and the even scarcer number of solution structures obtained by NMR. In particular PNA:RNA duplexes are rarer than those with DNA. To design effective systems is important not only to test the effect of modifications, but also to evaluate the effect of introducing new PNA monomers in duplex with the desired DNA or RNA target sequence. Manual modification of duplex structures requires to substitute bases with the desired ones, both on PNA and DNA or RNA. Moreover, these must be placed in the correct orientation allowing for H-bonding and stacking interactions. Little displacements of atoms and functional groups can induce destabilization of the system leading to erroneous simulations. For all these reasons, a manual approach is strongly discouraged and time consuming. In order to automatize this process we propose a modified version of Nucleic Acid Builder (NAB) implemented in AmberTools14. This program works placing backbone atoms in the appropriate positions forming an undistorted helix. Lastly, the remaining atoms of the molecule are introduced through the coordinates declared in the pdb files for each single monomeric unit. This procedure requires setting of two parameters, reads the helix rise and twisting. The current version of NAB already contains parameters optimized for A-DNA and B-DNA as taken from the literature,²⁴⁹ allowing for studying different DNA forms. PNAs, as found in the already known duplex structures, show conserved conformation with helical arrangement similar to the DNA parents. Such a similarity, allows using the same approach to generate duplexes with PNA, just properly modifying the original code. We included the PNA monomers in the list of the recognized residues and we provided the corresponding structures to be recalled by the program. The parameters needed for the helix construction were taken from data available in literature^{250,143} and the atoms employed for its generation are the corresponding atoms used for DNA and listed below.

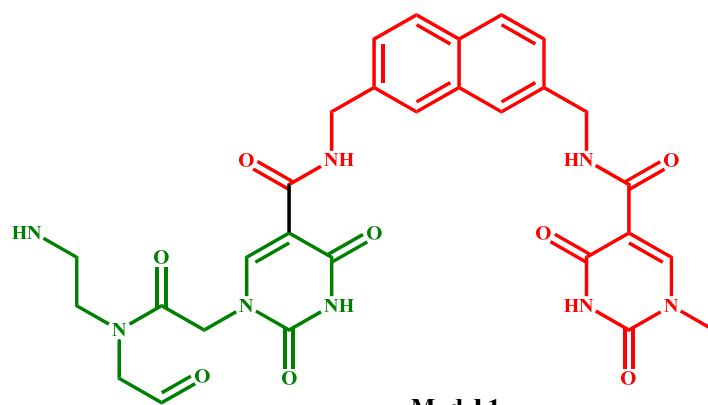
Table 2.1: List of atoms used to construct helix in original Nab program and corresponding PNA atoms used in our program.

DNA atoms	Corresponding PNA atoms
P	N1'
O5	C2'
C5'	C3'
O3'	C'
C3'	C5'
C4'	N4'
C1'	C8'

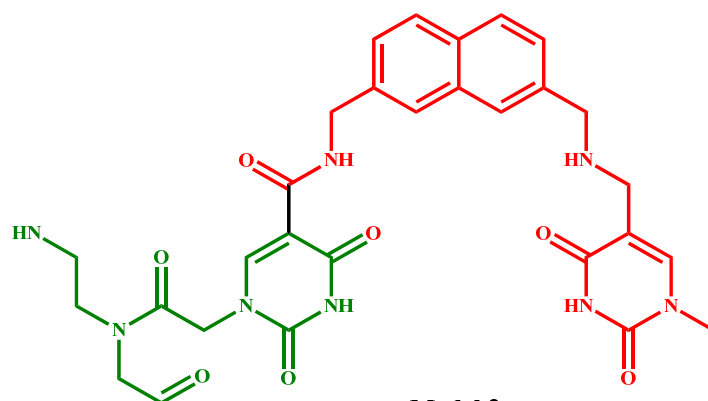
The advantage of our program is to automatize the whole scaffold construction saving a significant amount of time and gaining in accuracy and reproducibility. A full set of structures can be readily prepared in few minutes compared to several weeks of work as it used to be previously. Complete program list is reported in Appendix 8.3.

2.8. Modified bases parameterization

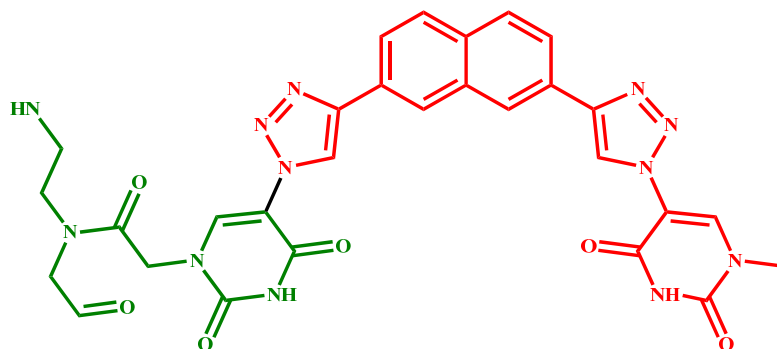
One attractive development is represented by PNAs made by modified monomers. The resulting building blocks can be designed with different complexity and functionalities in order to target the desired biochemical action. Computationally, this translates in more complex structures with non-standard moieties affecting on FF compatibility. Since modified residues are generally inserted in the central part of PNA chain, for each kind of modification (Figure 2.4) we decided to build a partial library focusing only on Central Fragments (Figure 2.3). This allowed to save computational time. However, a full library (containing also N-terminus and C-terminus fragments) would be of great interest and it is currently in the optimization process in our group. Each monomer was capped with ACE and NMA moieties and the geometry optimized with Gaussian 09 using DFT B3LYP level of theory and 6-31G* basis set. RESP calculations were performed with Gaussian 09 at the HF level of theory and using 6-31G* basis set. In this case, we used 10 shells and a density of 10 point/Å². Missing force field parameters for MD simulations were taken for analogy from *ff99sb* and from *GAFF*.



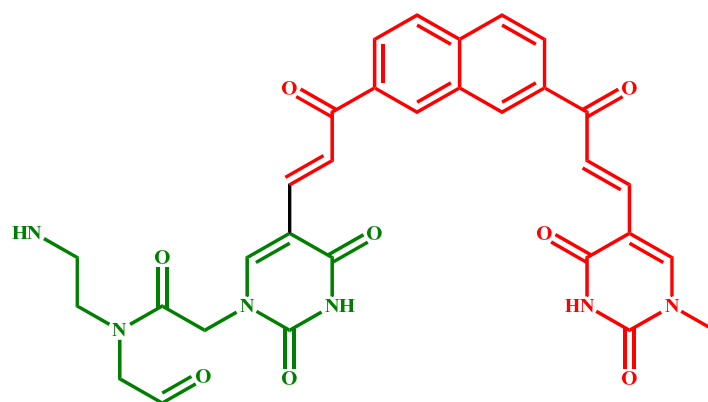
Model 1



Model 2



Model 3



Model 4

Figure 2.4: Structures of modified PNA monomers. Standard PNA fragments are colored in green and modifications are represented in red.

Starting duplex structures for MD simulations have been generated using NAB module. Modified bases can be inserted manually simply modifying the thymine structure or automatically inserted with NAB. In particular, **Model 1** has been already implemented in NAB code as a fundamental part of this thesis whereas all the other monomers will be included soon. A complete list of charges and parameters are reported in Appendix 8.6.

The Design and the relevance of these dimeric systems will be further described in Chapter 4 and 5.

3. Study of single-stranded PNAs and their interaction with RNA

3.1. Introduction

Understanding the role and the mechanism of action of large biomolecules normally relies on known existing solid-state structures as a starting point. However, as mentioned in chapter 1, in the literature only a limited number of structures containing PNAs can be found. For this reason, the possibility of performing reliable Molecular Dynamics simulation and virtual screening on single stranded PNA, as well as duplex structures containing PNA is highly desirable. In particular, this would enable prediction on the dependence of PNA activity upon structural modifications, by observing molecular details of interaction mechanism which are otherwise difficult to be probed by microscopy or spectroscopic techniques. Such approach could allow to selectively choose the most promising PNAs for the synthesis and experimental tests, based on binding properties prediction, thus saving much time and many resources spent on synthetic efforts.

Despite the growing interest of the scientific community on PNAs activity, the published computational studies are still very few. In particular, examples focusing on PNA:RNA models are very rare. Orozco and coworkers^{251,252} were among the firsts to explore the possibility to use Molecular Dynamics as a technique capable to describe PNA properties. Their first study concerned the comparison between PNA:DNA:PNA triplex already reported in literature with their simulation scheme.²⁵² A successive research has focused on PNA:DNA and PNA:RNA duplexes.²⁵¹ Gantchev et al. studied γ -radiation induced cross-linking in PNA:DNA heteroduplexes by means of a 300 ps long MD.²⁵³ Hatcher et al. performed studies of electron transfer in PNA and DNA single strands with 2 ns long simulations,²⁵⁴ here highlighting the greater structural parameter fluctuations of the PNA compared to the DNA and the resulting influence on charge transfer. Mansawat et al. collected 10 ns long simulations concerning the use of pyrene moiety as an universal base in a acpcPNA:DNA duplexes,²⁵⁵ revealing important structural information concerning the spatial arrangement of the inserted modification. Sanders et al. performed a 25 ns long simulation on modified PNA bearing a hypoxantine base for the targeting of KRAS miR and they validated simulated data with experimental evidences.²⁵⁶ Autiero et al. studied PNA:DNA and PNA:RNA duplexes focusing on the structural parameters.^{250,257} However, the understanding of the formation mechanism and of the factors governing stability of duplexes between PNA and RNA (or DNA) is still not complete, mainly

due to the multitude of degrees of freedom involved in this process. It is reasonable to believe that the duplex formation is somehow affected by the PNA single strand conformational freedom and to its ability to adapt to the helical conformation of the duplex. The validation of a MD-based procedure for the description of this process would lead to the dual benefit of providing insights into the duplex formation mechanism, and of developing a virtual screening tool for modified PNA moieties. Here we show our results concerning a systematic approach encompassing PNA single strand, and duplex PNA:RNA re-annealing and melting. Simulations described in this chapter were carried out in collaboration with the group of Prof. Michele Parrinello, at the Department of Chemistry and Applied Biosciences ETH Zurich, USI Campus, Lugano, using the computing facilities there available.

3.2. Validation of the force field by MD simulation of PNA:RNA duplexes

As discussed in Chapter 1, our final goal is to prepare more effective systems able to target micro RNA. For this reason, as a starting model for assessment of the force field for PNA duplexes, we have chosen the 176D¹⁴³ NMR structure (reported in the Protein Data Bank database), which is one of the few PNA:RNA duplex structures reported in literature at the time of this study (crystal structure of PNA:RNA duplex was resolved for the first time at the end of December 2015).²⁵⁸ Moreover, this 6 base long sequence (6 mer) has the appropriate length for our target modeling, since it is computationally non-expensive and its size is comparable to the putative duplex between a PNA and the seed region of miRs. The sequences of PNA and RNA are respectively H-GAACTC-O⁻ and 5'-GAGTTC-3' (Figure 3.1). This duplex was modified by removal of phosphate group bound to 5' residue of RNA, because the chosen force field (*ff99sb*) recognizes the RNA strand only without that group. As mentioned before, in the literature there are no consolidated force fields for PNA, and thus one of our aims was to improve their availability similarly to other biological cases as proteins or nucleic acids that are nowadays widely accepted. In order to test the parameters chosen (Appendix 8.1)²³⁵ we performed a 200 ns long Molecular Dynamics simulation on the PNA:RNA duplex described above, using the protocol described in detail in section 2.5. We solvated the system with explicit solvent, in a cubic box of 41 X 50 X 45 Å³, for a total of 9350 atoms.

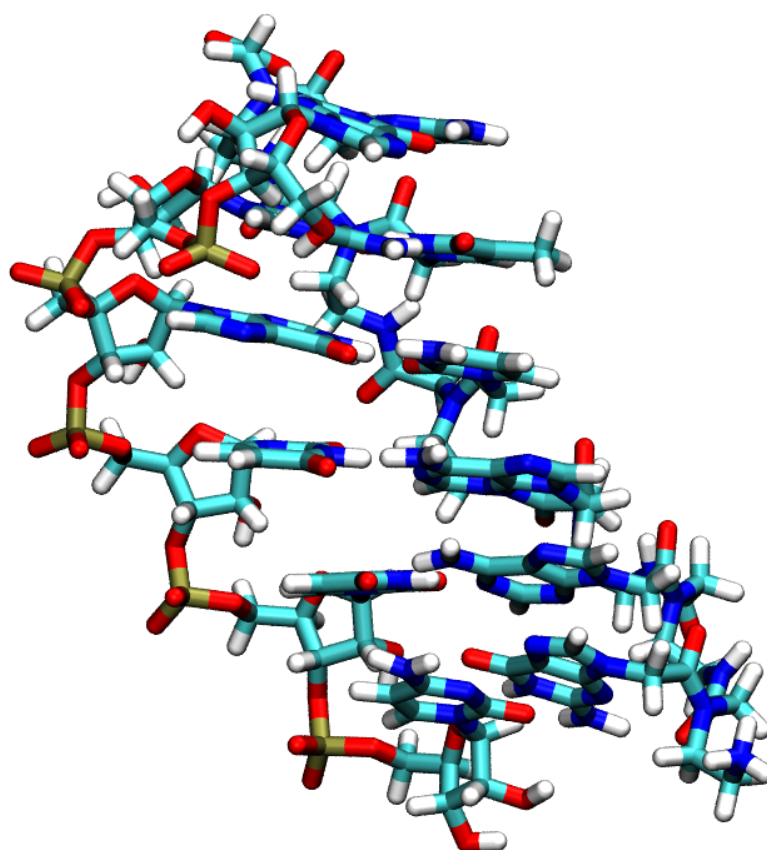
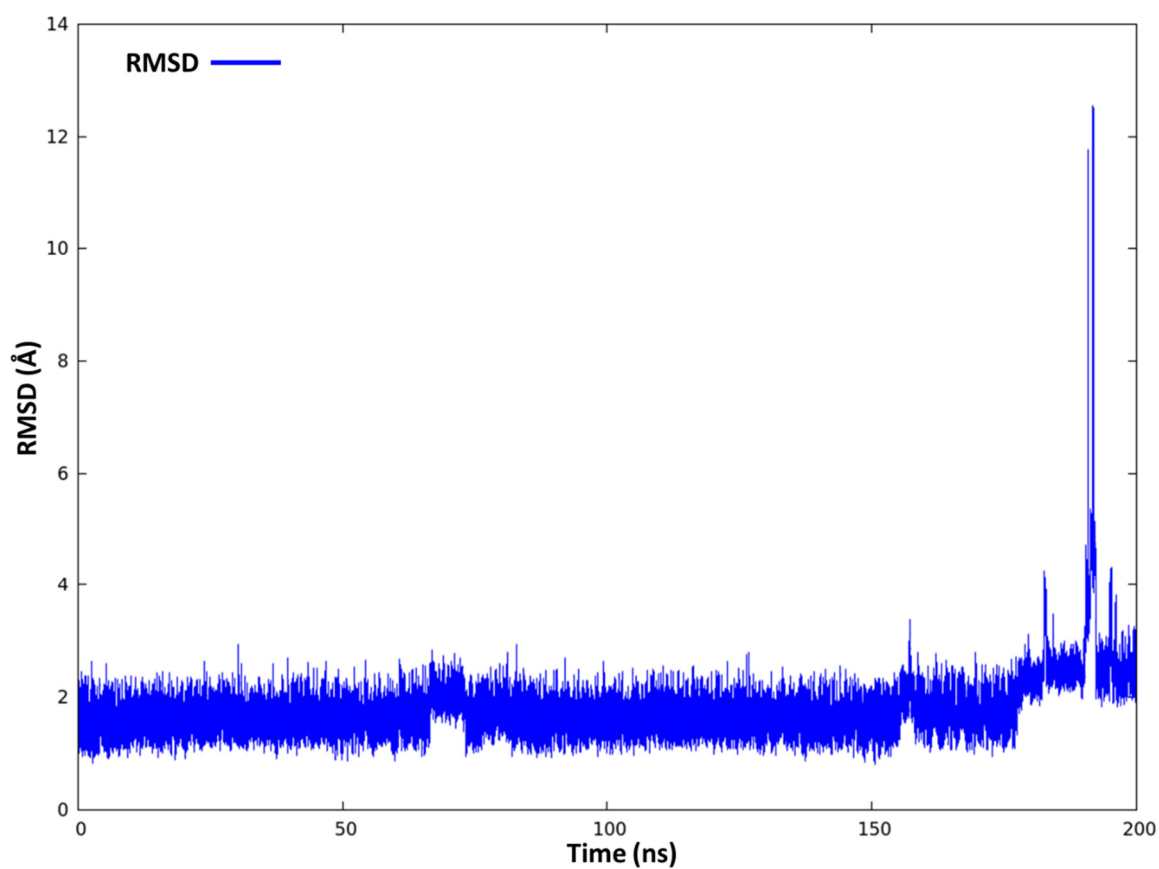


Figure 3.1: Structure of PNA:RNA duplex (176D), taken from Protein Data Bank, used for simulations.

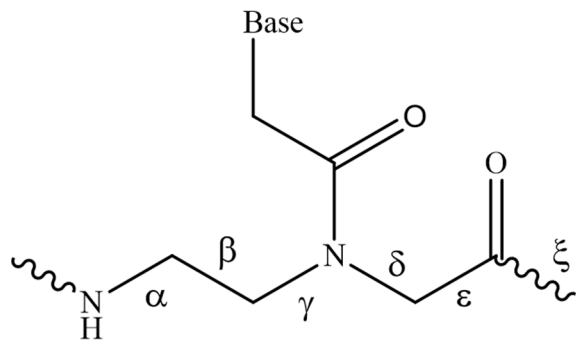


Graph 3.1: RMSD plot of simulated PNA:RNA duplex as function of time.

The duplex conformation was stable for all the simulation length with no significant structural modification. This can be inferred by examining the root mean square deviation (RMSD) that is less than 2 Å except for the very last few nanosecond (Graph 3.1).

The only significant degrees of flexibility of this duplex are represented by the H-Bonding disruption of a terminal couple of bases turning into stacking. This is in agreement with the general view that the two terminal sides of a nucleic acid helical duplex are more exposed than the inner base pairs to the solvent molecules that compete with the H-bond network. If the terminal hydrogen bonds are cleaved the system is destabilized and an efficient way to preserve the overall duplex structure is the stacking between the nucleobases of PNA:RNA terminal fragments. To better test the force field we checked the characteristic torsion angles of PNA obtained in the MD simulation, compared with those reported in literature.²⁵⁹ The results are in good agreement with the experimental ones (Table 3.1).

Table 3.1: left, characteristic PNA angles; right, comparison between simulated angles and those reported in the literature.²⁵⁹



The chemical structure shows a PNA monomer with a backbone consisting of a nitrogen atom bonded to a hydrogen atom and a carbonyl group. The backbone is extended through a series of carbon atoms labeled alpha, beta, gamma, delta, epsilon, and xi. A base is attached to the nitrogen atom. The torsion angles are defined as follows: alpha is the angle between the N-H bond and the C-alpha-C-beta bond; beta is the angle between the C-alpha-C-beta and C-beta-C-gamma bonds; gamma is the angle between the C-beta-C-gamma and C-gamma-N bonds; delta is the angle between the C-gamma-N and C-gamma-C-delta bonds; epsilon is the angle between the C-gamma-C-delta and C-delta-C-epsilon bonds; and xi is the angle between the C-delta-C-epsilon and C-epsilon-C-xi bonds.

Angle	MD (°)	Experimental (°)
α	86.8±10.9	[40 - 100]
β	250.8±15.1	[220 - 280]
γ	70.7±11.2	[40 - 100]
δ	80.8±8.1	[60 - 100]
ϵ	95.6±9.8	[60 - 120]
ξ	148.3±13.2	[130-180]
	339.2±16.9	[0 - 40;320 - 360]
	170.3±12.1	[160 - 195]

Next, we considered the MD simulation of γ -modified PNA with the force field developed. In section 2.6, we discussed the procedure used for charge and parameters derivation for γ -modified PNA monomers. Also in this case we needed the force field was validated before using it for further investigations on PNA properties. Therefore, we performed a 50 ns long simulation on a duplex obtained by manual insertion of serine side chain in γ position of each monomer of the PNA:RNA duplex 176D used in the previous simulation. In this case, the duplex was solvated in a cubic box of 59 X 55 X 57 Å³, for a total of 18100 atoms. This calculation was done with a shorter simulation time because, as for the previous one, duplex durability was expected.

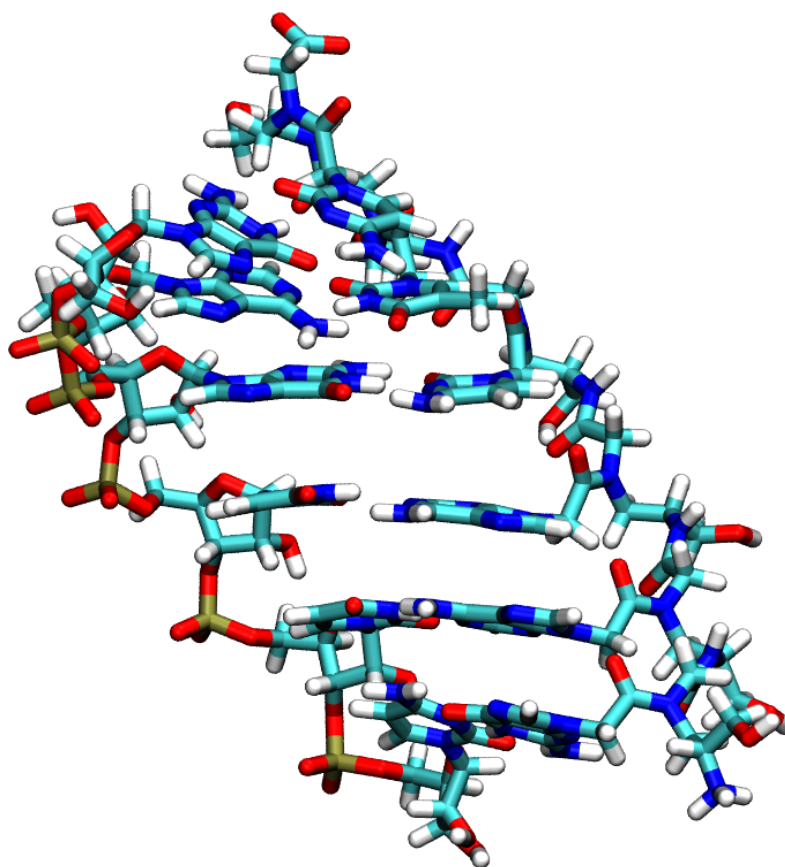
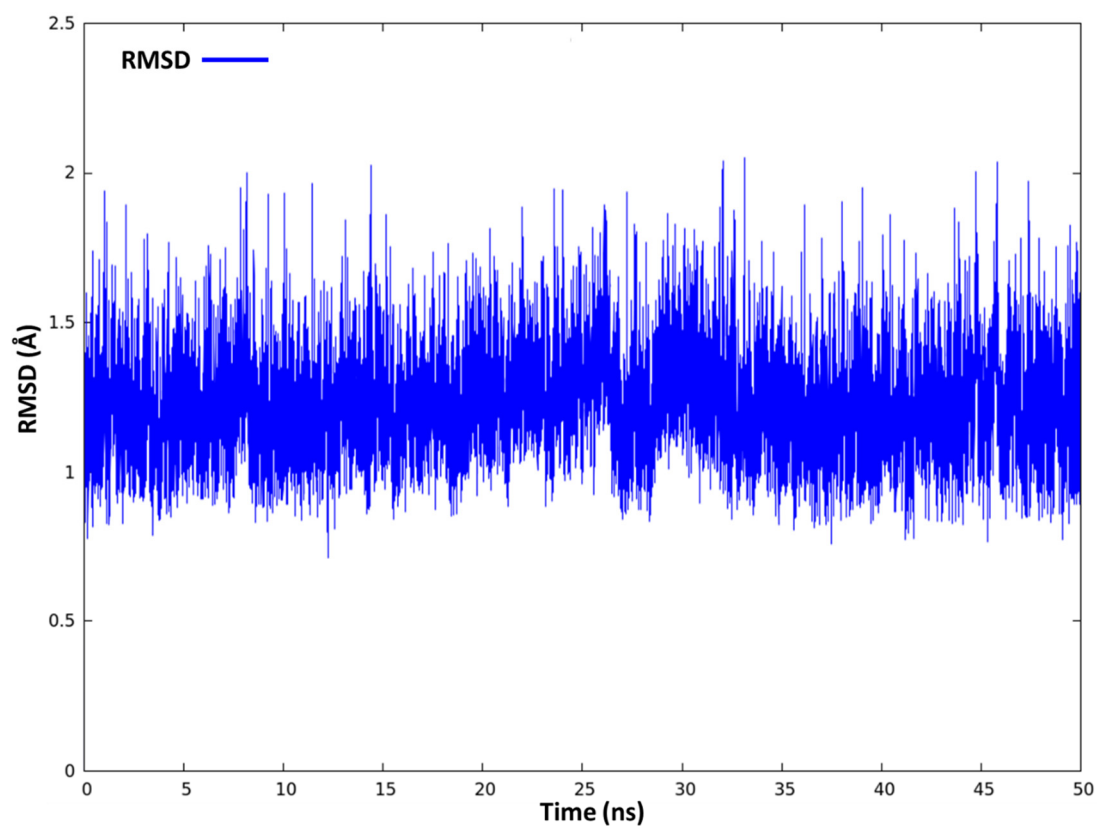


Figure 3.2: Structure of γ -modified PNA. The duplex was generated from system 176D retrieved from Protein Data Bank, by manual insertion of serine side chain in gamma (C5) position.

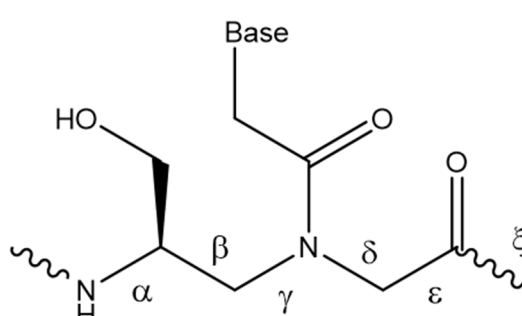


Graph 3.2: RMSD plot of simulated γ PNA:RNA duplex as function of time.

This system was expected to be even more stable than the unmodified one, according to the general properties of γ -modified PNA. Indeed, also in this simulation the duplex resulted perfectly paired for the entire simulation, as proven by the low RMSD (Graph 3.2).

Moreover, characteristic PNA angles were found also in this case to be compatible with those reported in literature (Table 3.2).

Table 3.2: left, PNA bearing serine side chain in gamma position and its characteristic PNA angles; right, comparison between simulated angles and those reported in literature.^{259,138}



Angle	MD (°)	Experimental (°)
α	67.3±7.3	[40-100]
β	69.5±9.3	[40-100]
γ	81.1±7.9	[60-100]
δ	94.7±9.5	[60-120]
ϵ	165.8±10.3	[130-180]
ξ	170.1±8.1	[160-195]

3.3. Simulation of single stranded unmodified PNA (ssPNA)

Understanding the degrees of freedom that characterize the single stranded PNA in solution gives important information on its capability of interacting and eventually binding to RNA. Indeed, conformational freedom of single stranded PNA (**ssPNA**) is directly and strongly related to how the duplex is easily and quickly formed. Moreover, flexibility will also affect the energetic terms in the formation of the final duplex. We investigated the flexibility of a PNA single strand (Figure 3.3) by means of Molecular Dynamics and Metadynamics (MTMD) in order to guarantee a complete sampling of the whole configurational space. Due to the size of this system, we solvated PNA with a larger cubic box of 53 X 67 X 59 Å³, including totally 20775 atoms. Initially, we performed a 200 ns long Molecular Dynamics simulation after thermal equilibration.

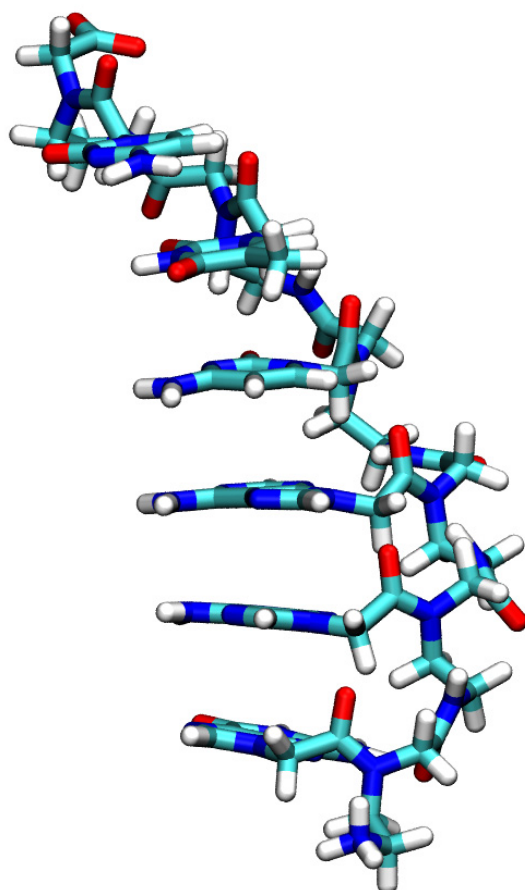
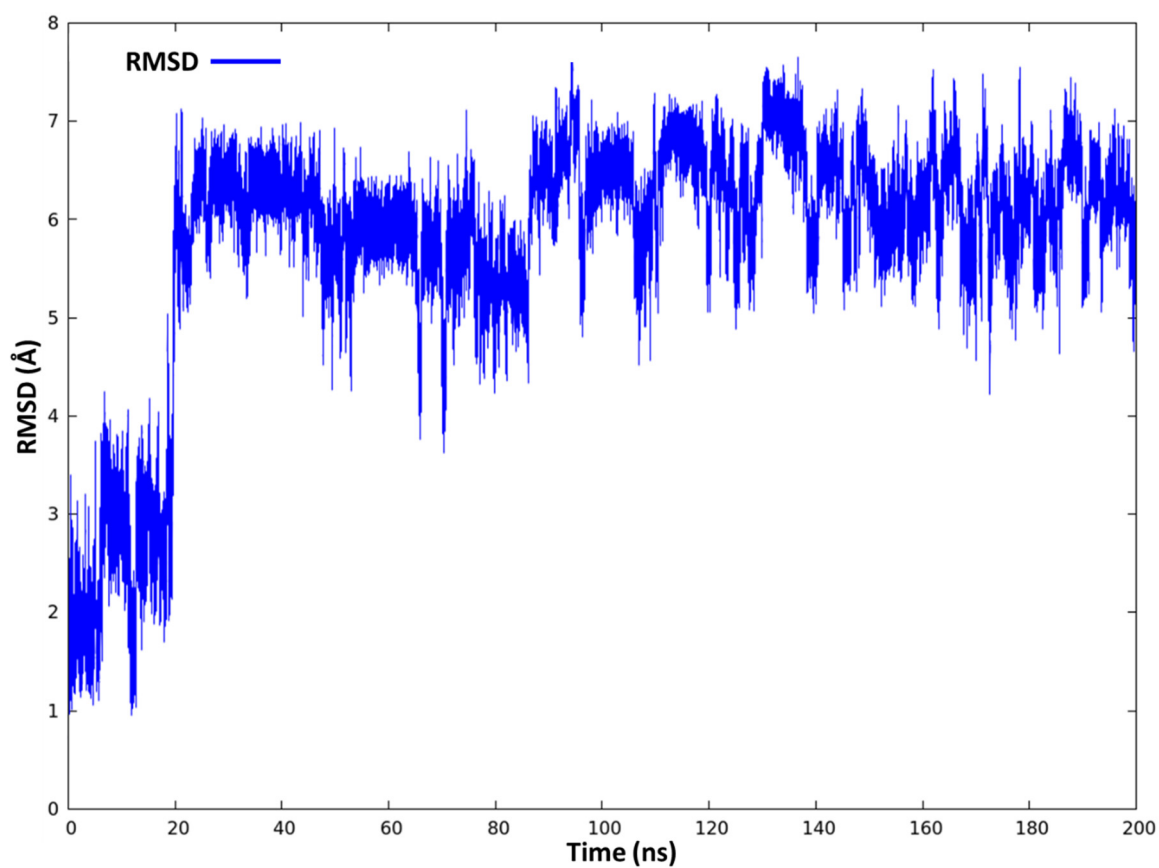


Figure 3.3: Single strand PNA, extracted from duplex 176 D, used as starting structure for simulation.



Graph 3.3: RMSD plot of simulated ssPNA as function of time.

In Graph 3.3, we plot the RMSD of the PNA, with respect to the thermal equilibrated structure, in order to identify relevant structural transitions. Within the first 20 ns the RMSD is relatively low and centered around 2-3 Å denoting a certain degree of stability of the initial helical conformation. Afterwards, a steep variation of the RMSD denotes a critical conformational change that lasts until the end of the simulation. This rapid variation is connected to a kind of “folding” of the PNA. We analyzed this structural transition and we discovered that structures showing RMSD values oscillating between 5 and 8 Å correspond to several conformers among which the system is oscillating. This analysis suggests that the initial helical conformation does not last very long and that in water solution the single PNA strand can be found in a multitude of different conformations. In order to have a complete picture of these conformers, we also investigated the populated structures in greater detail using collective variables (CV) (general description in section 2.3). First, we used the head-to-tail (H-T) distance between the centers of mass of the first and of the last monomer (\mathbf{R}). This variable describes the opening and closing motion of the system and it is related to “folding” and “unfolding” of the PNA. This CV is not sufficient to describe effectively the system because once this is folded there are many possible different stacking interactions. In order to better discriminate these stabilized stacked conformations, we decided to use a local order parameter previously developed for describing crystal nucleation.²⁶⁰ For each base, we defined a vector lying in the plane of the rings. This CV takes into account the distance, the angle and the coordination number between these vectors. Coordination number represents the number of vectors close within a defined cutoff. This parameter is important when studying nucleation since it determines the presence of an aggregate. The PNA considered in our study is 6 mer long and the maximum theoretical coordination number for each vector is then 5. However, our purpose is not to define an aggregate, but to discriminate stacking interactions. Therefore, for each couple of bases, we defined variables in order to have the coordination function ρ_i (equation 3.2) always set to one, thus ruling out association. Distances and angles between these vectors are extremely important in stacking description: when two bases are stacked their distance should be defined in a given cutoff range and the angles should assume discrete values. An appropriate definition of this combination allows determining whether two or more bases are stacked (Figure 3.4 A) or fully (Figure 3.4 B – C) and partially unstacked (Figure 3.4 D).

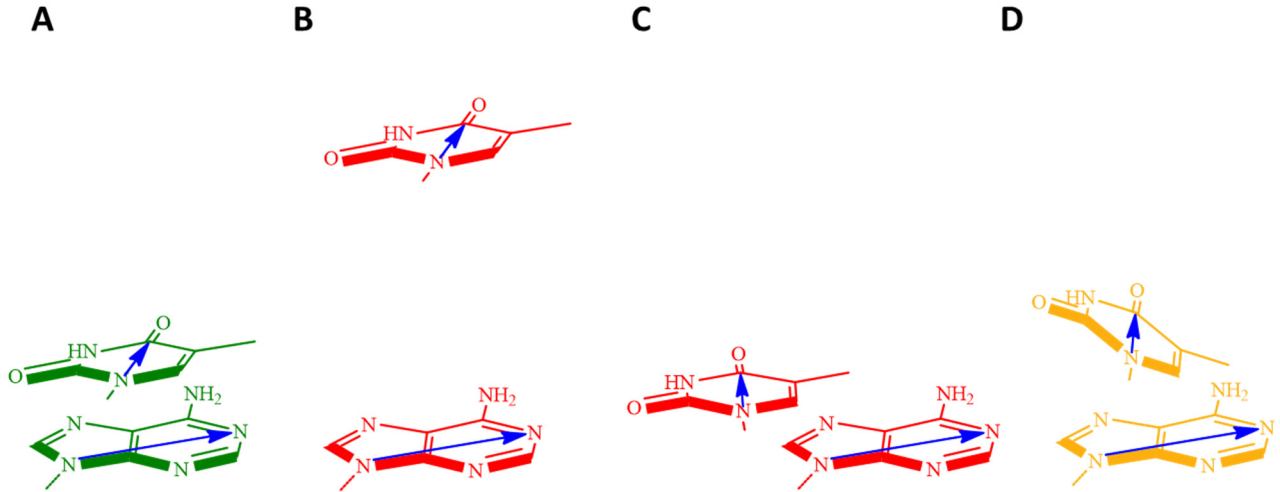


Figure 3.4: Schematic examples of stacking, not stacking and partial stacking arrangements. In blue are represented vectors used to describe local order parameter. **A)** Two bases are stacked; **B)** two bases are too distant for stacking interaction; **C)** two bases are close, but not stacked. In this case distance is favorable, but not the angle; **D)** Distance between bases is optimal, but angle not completely thus leading to partial stacking.

Going into the details, local order parameter is defined as the product of two sigmoidal and one single gaussian function:

$$f_{ij} = \frac{1}{(1+e^{a(r_{ij}-r_{cut})})} \quad (3.1)$$

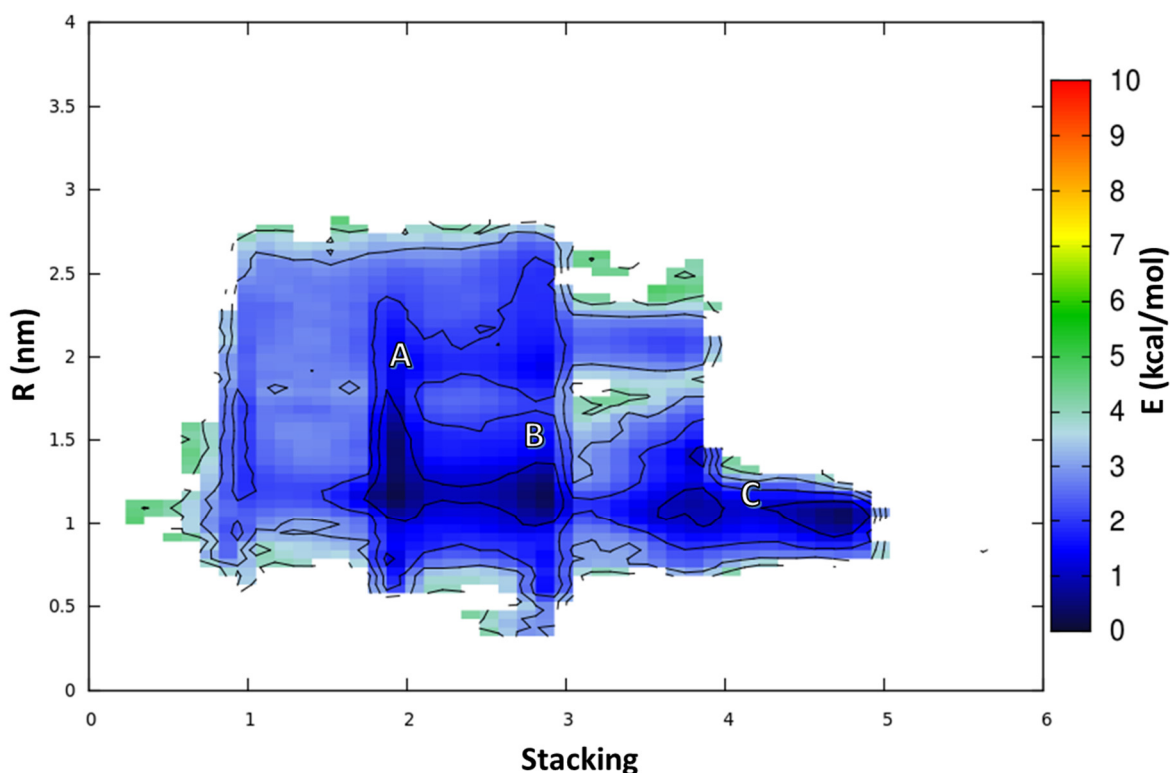
$$\rho_i = \frac{1}{(1+e^{-b(n_i-n_{cut})})} \quad (3.2)$$

$$\theta_{ij} = \sum_{k=1}^{k_{max}} e^{-((\vartheta_{ij}-\vartheta_k)^2/2\sigma_k^2)} \quad (3.3)$$

where \mathbf{r}_{ij} are the distances between the above discussed vectors, \mathbf{r}_{cut} is cutoff distance for the stacking interaction, \mathbf{n}_i is coordination number, \mathbf{n}_{cut} is cutoff for coordination number, ϑ_{ij} is the angle between the vectors, ϑ_k is a favorable angle for stacking and σ_k is the width of the gaussian applied on that angle. Lastly, \mathbf{a} and \mathbf{b} are exponential factors, determining how steep are the sigmoids.

These functions essentially monitor respectively the distance between bases (equation 3.1), the coordination number (equation 3.2) and angle between bases (equation 3.3). As discussed above, the coordination parameters were set to obtain a value of $\rho_i = 1$ ($\mathbf{n}_{cut} = 1$). The distance function \mathbf{f}_{ij} was set to have a value of 1 for distance within a range from 5.0 to 6.5 Å, depending on the couple of bases i and j considered. Above the cutoff distance, the \mathbf{f}_{ij} value rapidly decreases to 0. The angle

function θ_{ij} is a sum of functions, one for each characteristic angle chosen. For each angle θ_k a Gaussian function that exhibits maximum value of 1 for θ_k , is defined. In order to have stacking, bases should present opportune values of distances and angles. Based on MD simulation data, the coefficients (r_{cut} , θ_k , σ_k , \mathbf{a}) were tuned in order to maximize f_{ij} and θ_{ij} when bases are stacked. The functions here described are referred to a single couple of vectors, but in our system we have several possible couples and also multiple bases coupled at the same time. Description of every single couple is not meaningful alone, so to describe entirely the system we used a linear combination of these local order parameters, defined for couples of bases, and we considered this combination a measure of total stacking (this function is here after indicated with **Stacking**). Then, we constructed the free energy surface as a probability distribution²¹⁹ (equation 2.10) as a function of **R** and **Stacking**. In Graph 3.4, we report the Free Energy Surface (FES) obtained for the **ssPNA** (extracted from duplex 176D) within an energy range between 0 and 10 kcal/mol (referred to the lowest energy conformation), which can be considered wide enough at 300K.



Graph 3.4: Free energy surface (FES) as function of head to tail distance (**R**) and **Stacking** function obtained from the probability distribution of unbiased MD simulation.

The free energy surface appears quite flat: the local minima are less than 2 kcal/mol more stable than all the other conformations and a very tiny energy barrier separates them. This finding suggests that the single-stranded PNA is free to explore a multitude of conformations avoiding any pre-organized structure capable for potential RNA binding.

The structural analysis conducted on the MD trajectory highlights three principal clusters (Figure 3.5: A, B, C). All of them share the same tail-head distance ($R \sim 1.1$ nm), but with different **Stacking** contribution. The first cluster of structures (**A**) is characterized by two couples of bases stacked (**Stacking** = 2, violet and yellow, white and orange), in the second (**B**) three couples of bases (**Stacking** = 3, violet and yellow, white and orange, green and red) and in the third (**C**) more complicated structures with piles of bases leading to higher **Stacking** values (**Stacking** between 4 and 5, white and orange, piling of yellow, violet, red and green / red and green, piling of orange, white, yellow and violet). According to the calculated FES these structures differ in energy roughly by 0.5 kcal/mol (**A** and **B**) and 0.3 kcal/mol (**B** and **C**) and the barriers to be crossed are lower than 2 kcal/mol. Nonetheless, the evolution of the system is not completely random, as could be supposed by RMSD plot, but it is characterized by the passage between different stacked conformations.

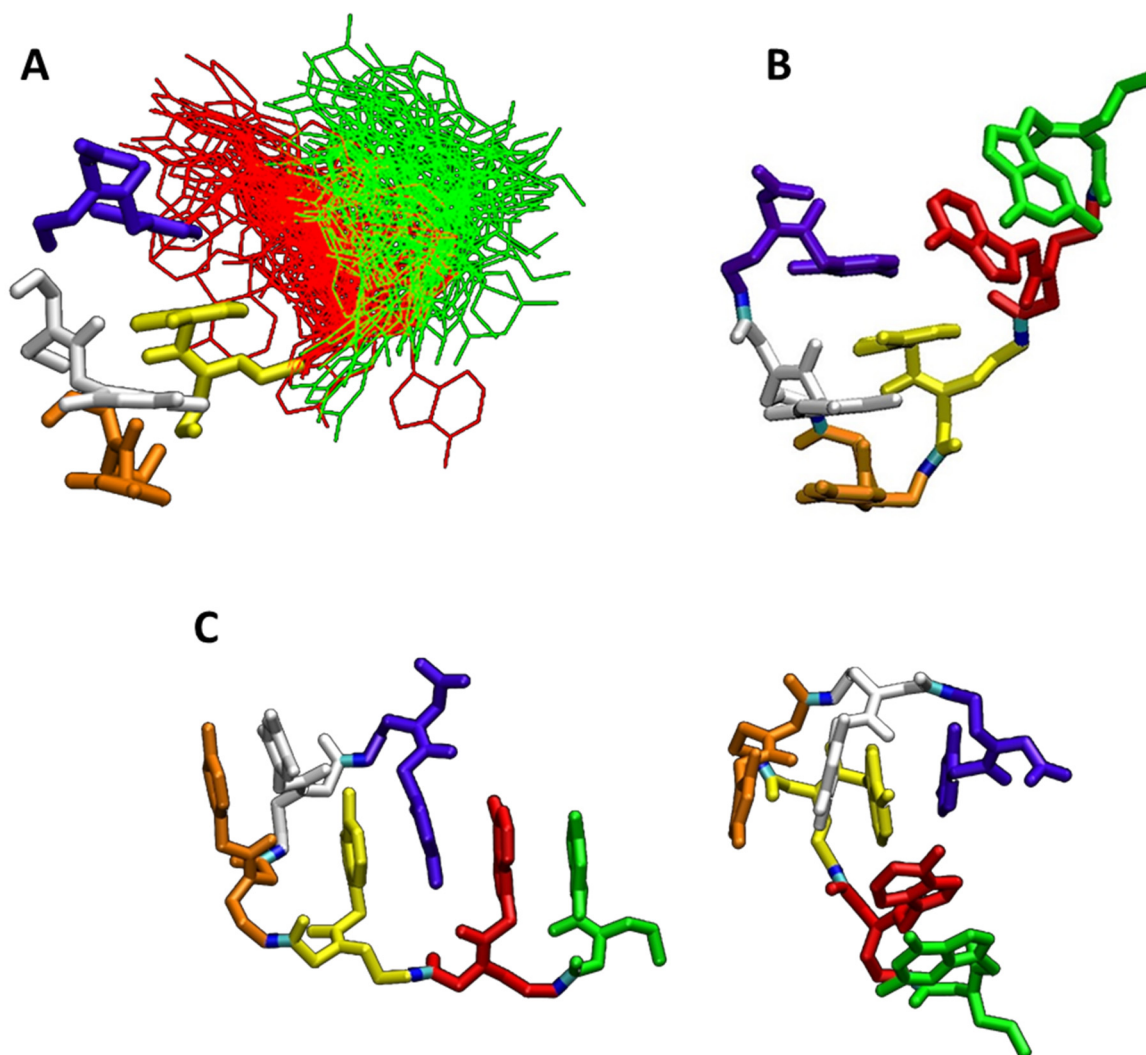
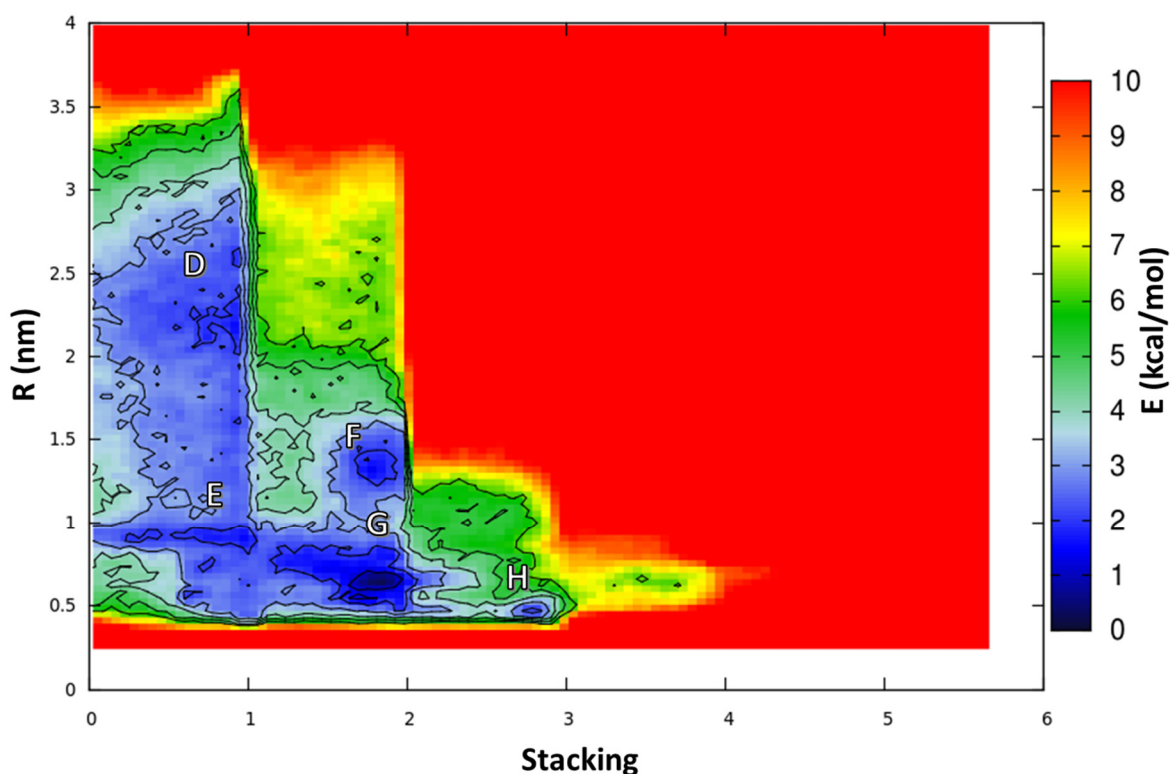


Figure 3.5: Local minima highlighted by Molecular Dynamic simulation. Color code: H-GAACTC-O⁻.

As discussed previously, free energy surface obtained from distribution of probability is ideally correct only for infinite times, when the system is at equilibrium. However, such kind of systems, with high degree of freedom, is intrinsically complicate to be modeled by standard MD simulations and many important global minima can be missed. For this reason, we aimed to better explore the conformational space of the **ssPNA** by means of Metadynamics. To build the Metadynamics bias during the simulation we deposited every picoseconds a Gaussian hill with width of 0.01 (nm for **R** and dimensionless for **Stacking**) and height of 2.5 kJ (~ 1 $k_B T$) on the same CVs described above. To ensure the convergence of the Free-energy calculation, we adopted the Well-Tempered Metadynamics scheme (WT-MDMT), as discussed in section 2.4, with gamma equal to 20. External potential on the CVs allowed the systems to populate all the possible conformations of interest. We carried out a 950 ns long WT-MDMT, and the simulation was stopped after verifying that the local minima converged.



Graph 3.5: Free energy surface (FES) reconstructed from Gaussian hills deposition of MTMD. Quantities on x- and y-axes are the same plotted in Graph 3.4.

In Graph 3.5, we report the FES obtained biasing the collective variables described before. The flexibility of the PNA single strand is confirmed by the MDMT, which allows to explore wider configurational space if compared with the classical MD. In particular, the completely unfolded configuration ($R > 2$ nm) has weaker stacking between 0 and 1. This suggests that the PNA single strand can also populate non-staked structures. On the contrary, higher levels of stacking imply an enthalpic gain and the corresponding conformations are stabilized when the PNA is significantly folded for $R < 1$ nm. **G** is the global minimum and it differs from others by less than 2.5 kcal/mol (**D** 2.2 kcal/mol, **E** 1.2 kcal/mol, **F** 1.5 kcal/mol, **H** 2.2 kcal/mol). This last FES computed through the Metadynamics expand significantly the configurational space including the one reported in Graph 3.4 and it provides an important added value to the classical MD simulation. A structural analysis of the conformations found inside the principal minima are reported in Figure 3.6. As observed, none of them shows a helical structure confirming the experimental data²⁶¹ for unmodified PNA and resulting in hardly obtainable crystal structures justified by the intrinsically high degrees of freedom. In conclusion, both classical MD and MDMT bring to similar results showing how this PNA single strand can freely assume a wide range of conformations avoiding any significant structural preorganization otherwise important for RNA binding. We believe that this finding can be reasonably extended to unmodified PNAs with other sequences.

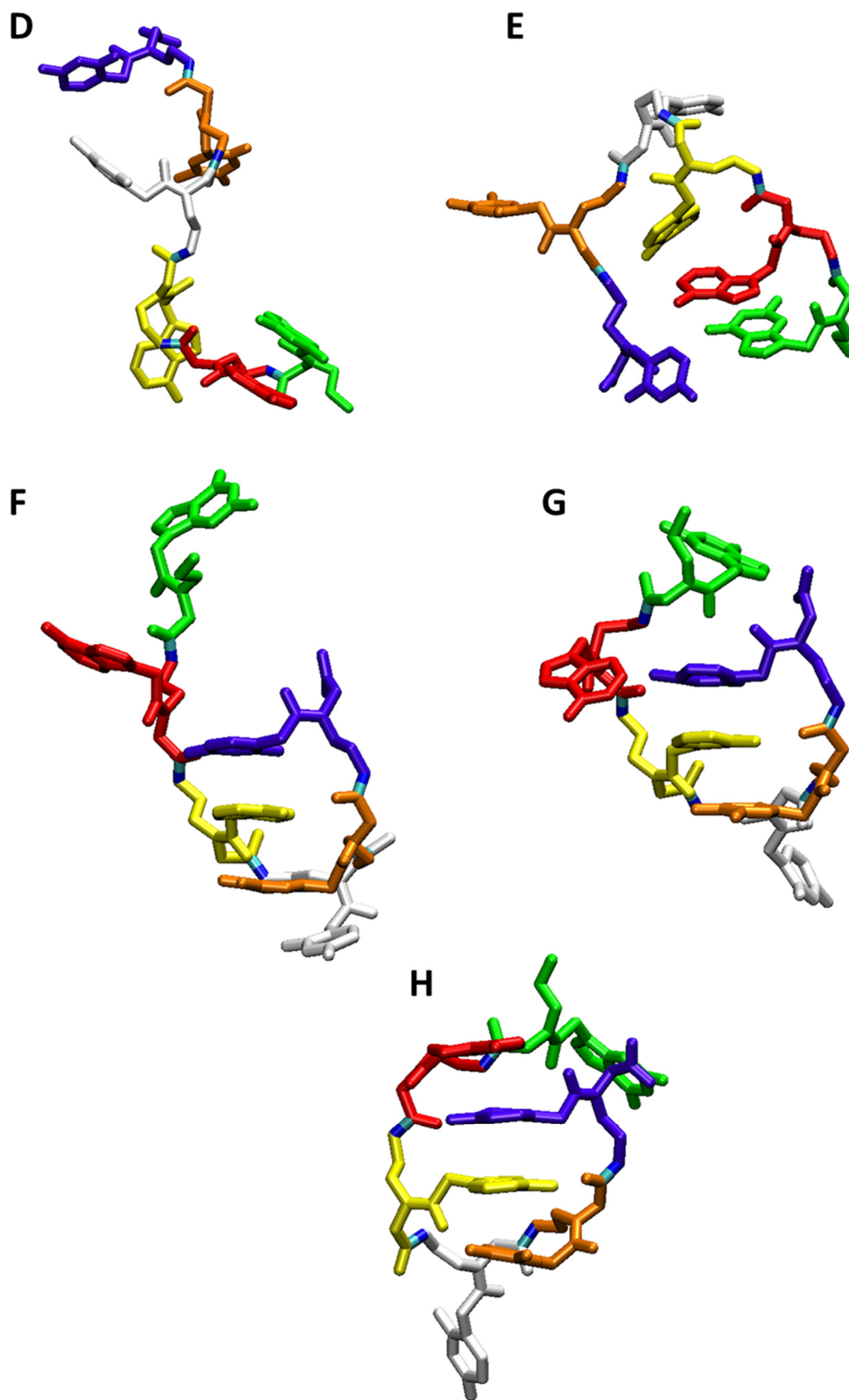


Figure 3.6: Structures of local minima from MTMD. Color code: H-GAACTC-O⁻.

3.4. Simulation of single stranded gamma-modified PNAs (ss- γ -PNA)

As discussed briefly in Chapter 1, modification of the backbone usually affects preorganization of the system. γ -modified PNAs present a side chain on the γ -position (C5) (Figure 3.7).

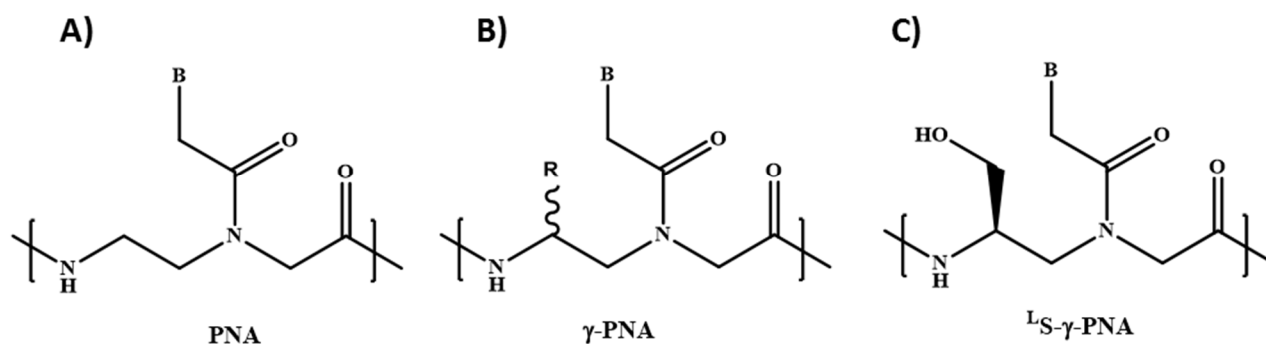


Figure 3.7: Comparison between A) standard PNAs, B) gamma modified PNAs and C) gamma serine modified PNA considered in this study.

Many examples of this type of PNA are reported in the literature.^{107,262,263} Such modified PNAs form duplexes with DNA or RNA that are more stable compared to unmodified ones. The properties of these compounds have been related to the preorganization of the backbone.¹¹⁴ Circular Dichroism spectra of γ -modified PNAs present signals in 200-230 nm range that are not present in unmodified PNAs. The CD signals in this region are due to the carbonyl chromophores of the backbone and were attributed to a helical disposition of these units. We then wanted to better study the effect of the introduction of gamma-modified PNA monomers in conformational properties of the single strand (**ss- γ -PNA**) as a test for the ability of molecular dynamics to predict this different behavior and to give molecular details accounting for the observed differences. Many groups have been attached in this position of the backbone in previous studies, but to reduce computational cost we selected a very simple serine side chain. Moreover, since experimental CD signals are proportional to the number of modified residues, we decided to modify every single monomer of the sequence.

In order to have a comparison with the unmodified PNA, we manually inserted serine groups in the helical structure used in section 3.3 (Figure 3.8). The duplex was solvated in a cubic box of 56 X 74 X 63 Å³ for a total of 21100 atoms and then we performed a 200 ns long MD simulation in the conditions reported in section 2.5. The original helix undergoes some significant structural modification within the first 60 ns of our simulation as demonstrated by the RMSD reported in Graph 3.6.

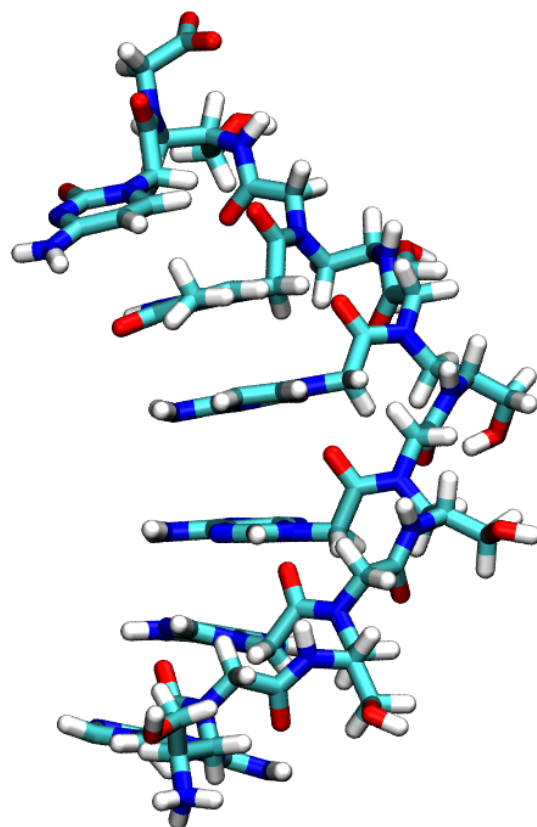
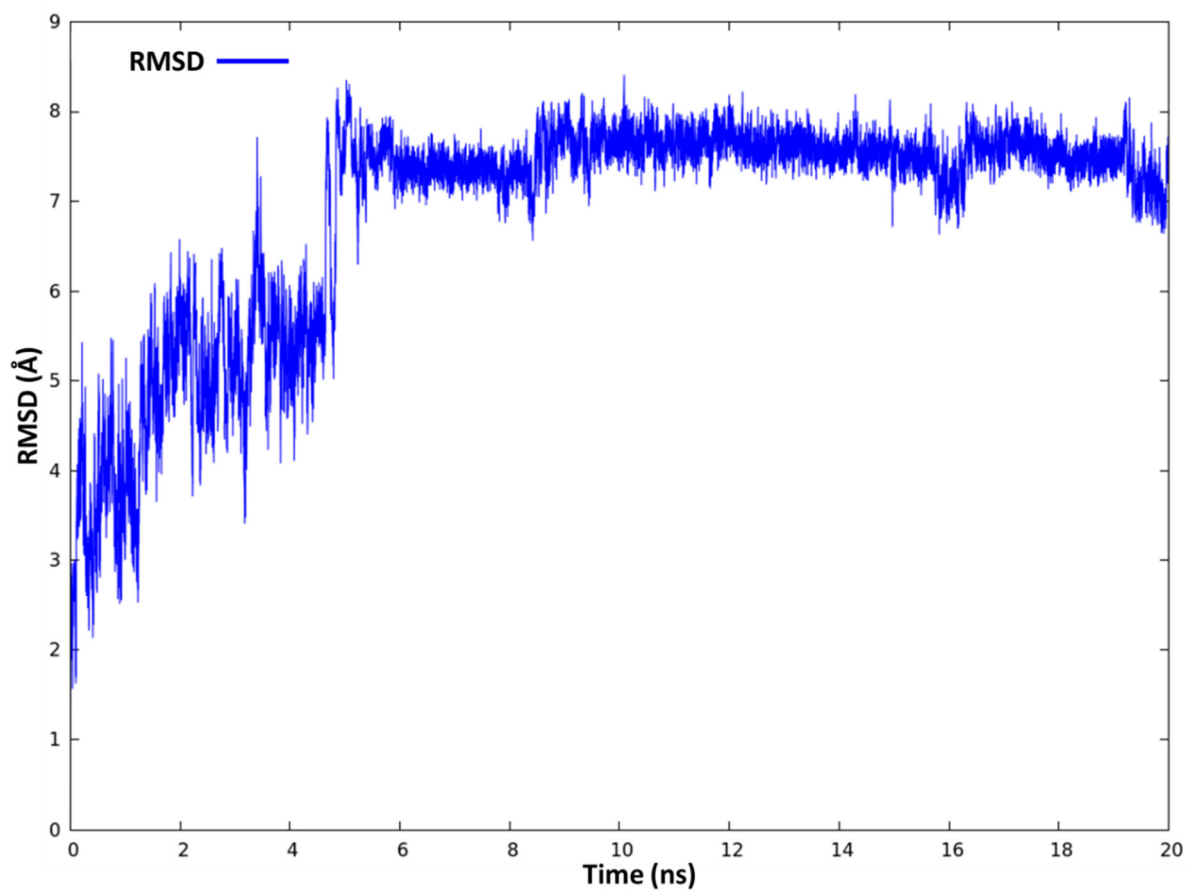
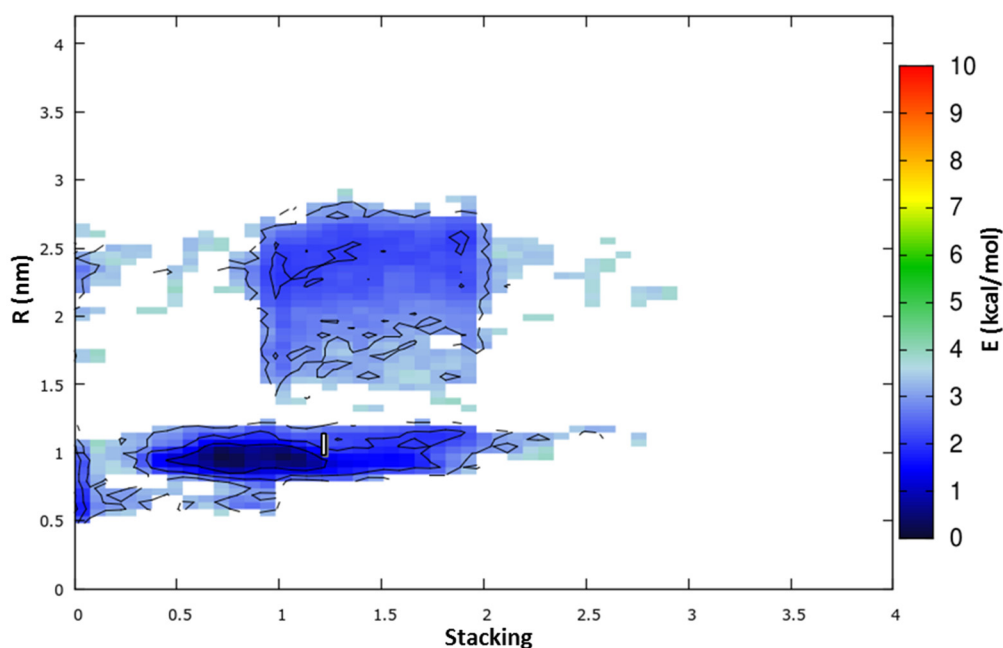


Figure 3.8: Analogue structure of ssPNA with γ - modified PNA bearing a L-serine side chain.



Graph 3.6: RMSD plot of simulated ss- γ -PNA as function of time.

However, the chemical functionalities positioned in γ induce such a lower flexibility of the backbone if compared with the **ssPNA** discussed in the previous section. The FES obtained with unbiased MD, reweighted on the two CVs discussed before for the unmodified PNA, shows one local minimum characterized by an average **R** (H-T distance) equal to 1 nm and a **Stacking** coefficient between 0.5 and 1 (Graph 3.7). From this basin we extracted several structures very close each other (Figure 3.9). Indeed, the γ -modified PNA quickly converge to a preferred conformation that last until the end of our simulation (Figure 3.9) and that is characterized by a relatively close-packed structure stabilized by the H-Bonding network established between the terminal guanine and the cytosine.



Graph 3.7: Free energy surface (FES) as function of head to tail distance (**R**) and **Stacking** function obtained from the probability distribution of unbiased MD simulation.

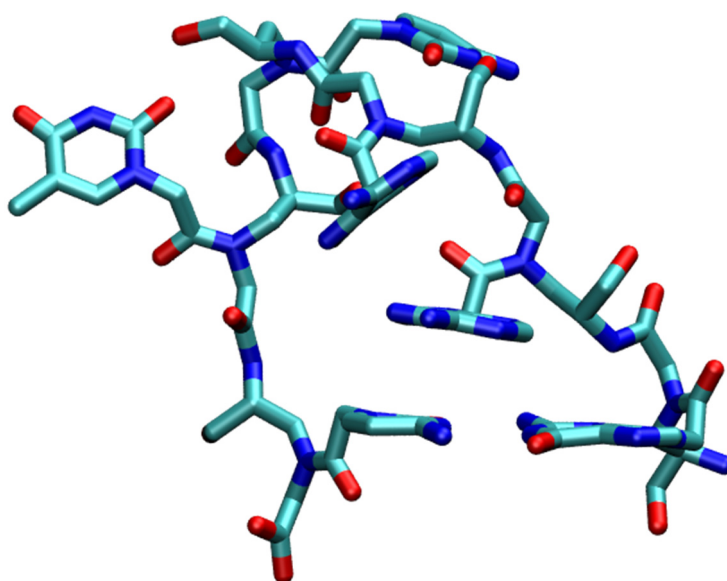
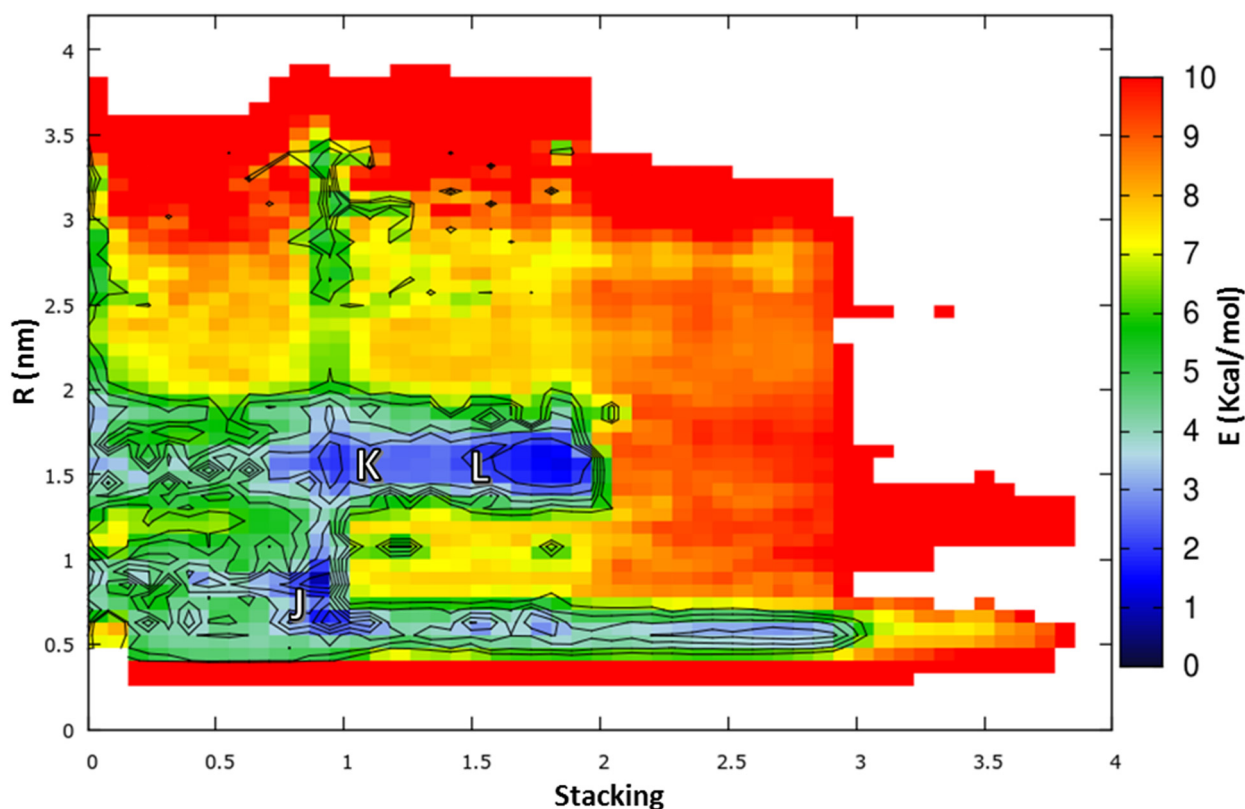


Figure 3.9: Representative structure of minimum I.

Even if conformational freedom is reduced compared to **ssPNA**, also in this case it is necessary to use Metadynamics to better explore conformational space. The PNA strand was solvated in a cubic box of 71 X 71 X 71 Å³ for a total of 34700 atoms. To build the Metadynamics bias we deposit every picoseconds a Gaussian hill with width of 0.01 (nm for **R** and dimensionless for **Stacking**) and 2.5 kJ (~ 1 k_BT) of height on the same CVs described in section 3.3. To ensure the convergence of the FES we adopt the Well-Tempered Metadynamics scheme, with gamma equal to 30. External potential on the CVs allows the systems to populate all the possible conformations of interest. We carried out a 400 ns long WT-MDMT verifying, also in this case, the local minima to be converged. In Graph 3.8, we report the FES obtained biasing the usual collective variables.



Graph 3.8: FES reconstructed from Gaussian hills deposition of WT-MTMD.

The rigidity of **ss-γ-PNA** is confirmed by the WT-MDMT and the system populates a reduced, almost isoenergetic (**L** global minimum, **J** +0.1 kcal/mol, **K** + 0.4 kcal/mol) number of states compared to **ssPNA**. Nevertheless, both from unbiased and biased simulations appear that the initial helical conformation is not preserved. However, we observed structures in the three main minima of the

FES and those in basin L are particularly interesting (Figure 3.10).

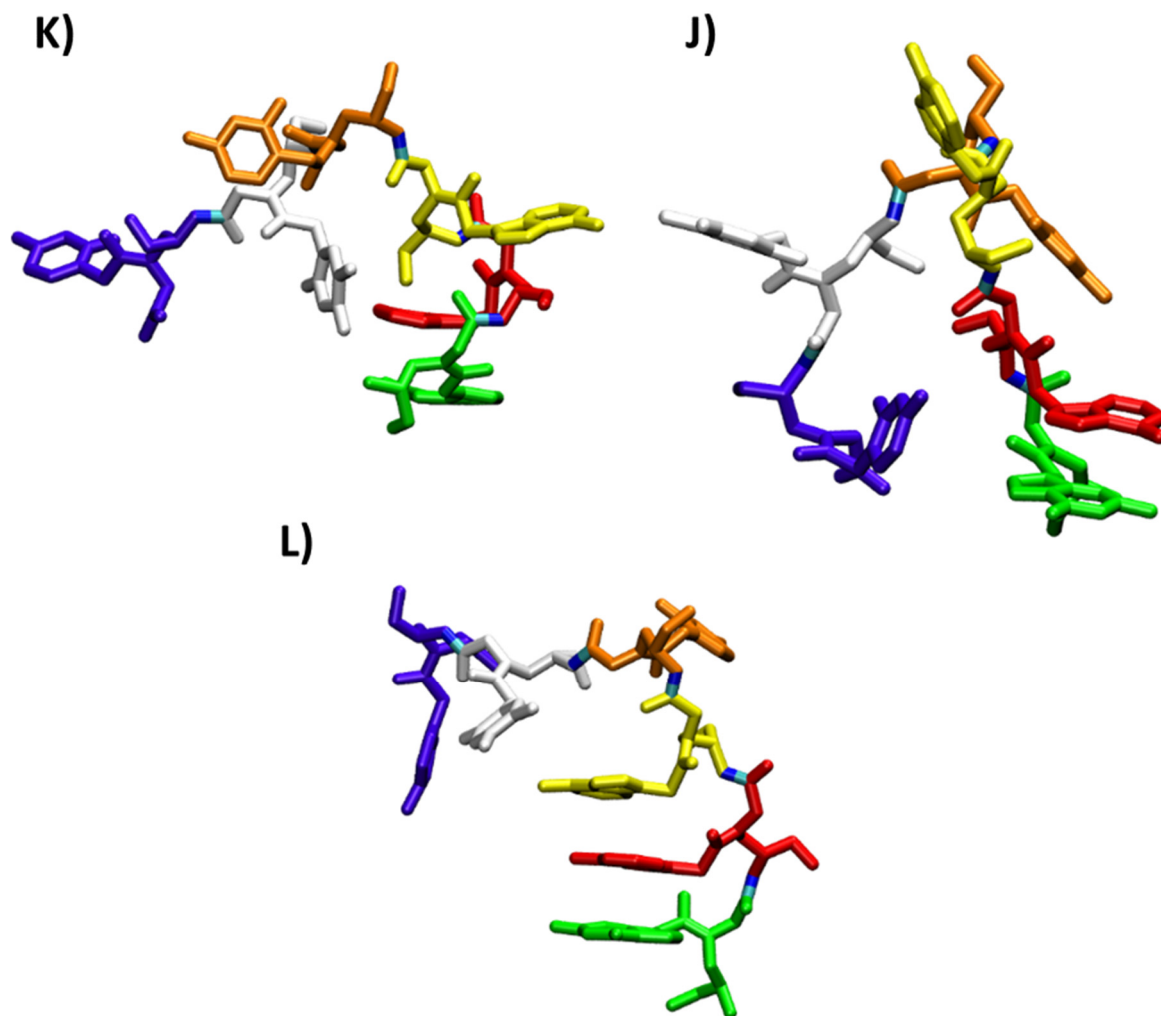


Figure 3.10: Representative structures of main minima of MTMD. Color legend is the same of unmodified PNA.

Portions of the original helical conformation are preserved in this type of structures, in particular three bases starting from the N terminus. The origin of this asymmetrical behavior is connected to the specific sequence, with a three purine on one side and a three pyrimidine on the other side. We therefore suggest that Circular Dichroism spectra reported in literature for this type of PNA¹¹⁴ does not necessarily represent a stretched helical conformation of the PNA as that present in the duplex (starting conformation in the simulation), but rather a cumulative effect of local helically shaped conformation favored by backbone rigidity.

Another experimental evidence that can be explained by an induced pre-organization of the PNA single strand is represented by the melting temperature (T_m) registered in order to quantify the relative stability of PNA:RNA duplexes. It is known that for each chiral unit an increase of $\sim 3^\circ\text{C}$ in

T_m for PNA-DNA duplex and $\sim 2^\circ\text{C}$ for PNA-RNA occurs.¹¹⁴ This can be correlated with the balance between the enthalpy and the entropy of the PNA single strand in solution: higher rigidity translates in a lower internal entropy of the system increasing the possibility of establishing a most durable H-bonding network and consequently requiring higher temperature to melt. The balance between ΔH and ΔS is important also for duplex formation: single stranded PNA is a highly disordered system whereas the resulting duplex is a well-organized complex stabilized by Watson and Crick Bonds. The formation of the duplex is therefore accompanied by a large entropy loss, which is compensated by the favorable enthalpic gain due to hydrogen bonding and stacking interactions. However, if the starting state has a less disordered structure, as in the case of the γ -PNA, due to restricted conformational freedom, the entropy variation upon duplex formation will be less negative than in the case of the flexible unmodified **ssPNA**.

For example, using melting data reported in the work of Ly and coworkers¹¹⁴ and fitting these with the model described by Marky and Breslauer for a two-state transition,²⁶⁴ it is possible to calculate the entropy loss for an unmodified PNA and for γ -modified PNA during duplex formation. For the 10 mer PNAs, considered by Ly, results a ΔS of -883.98 J/molK for unmodified PNA whereas it becomes -683.88 J/molK for γ -modified PNA. Modification in gamma position determines then a reduction in entropy loss of about 200 J/molK , in agreement with the above reported findings of MD simulation. Studies are underway to repeat these data using the same sequence used for the simulation.

3.5. MD studies on the interaction between PNA and RNA

Study on single strands permitted to evaluate interesting properties of two PNA systems, highlighting the crucial role of entropy in the behavior of the molecule, but of course, the structure and stability of the duplex and the mechanism of its formation are also crucial points. For this reason we have studied the PNA:RNA duplex in two different processes: duplex re-annealing and duplex melting. These are complementary process and in our study they lead to concordant conclusions that will be here discussed.

3.5.1. Duplex re-annealing

In order to understand possible mechanisms that lead to the formation/dissociation of PNA:RNA duplex we applied our methodology to the study of double strand re-annealing after an induced external stress. As a general strategy, we chose to use two strands (PNA and RNA) with partially base pairing, since a simulation of annealing from totally dissociated strands is computationally too demanding (even for high performance computing tools), since the two strands should a) encounter one another; b) initially pair (nucleate) with all the possible base combinations; c) anneal from different nucleation sites.

We evaluated the molecular dynamics of the system starting from five different distorted geometries (Figure 3.11) in which the two strands were placed in close contact and some of them with partial base pairing. **DIST1** and **DIST2** are spread like a fan on 5' and 3' ends, in **DIST3** strands are shifted for 5-6 Å, in **DIST4** strands are crossed and in the **DIST5** the two strands were stretched using MTMD and put close together. We thermally equilibrated each structure in a cubic box filled with TIP3P water molecules (box size and atoms: **(1)** 45 X 53 X 52 Å³, 12300 atoms; **(2)** 49 X 59 X 47 Å³, 10000; **(3)** 53 X 54 X 55 Å³, 11900; **(4)** 52 X 54 X 51 Å³, 10550; **(5)** 57 X 53 X 75 Å³, 17800) and performed a 200 ns long MD simulation, in the conditions reported in section 2.5. Within this time frame, we can investigate the evolution of these stressed configurations understanding key steps of re-annealing.

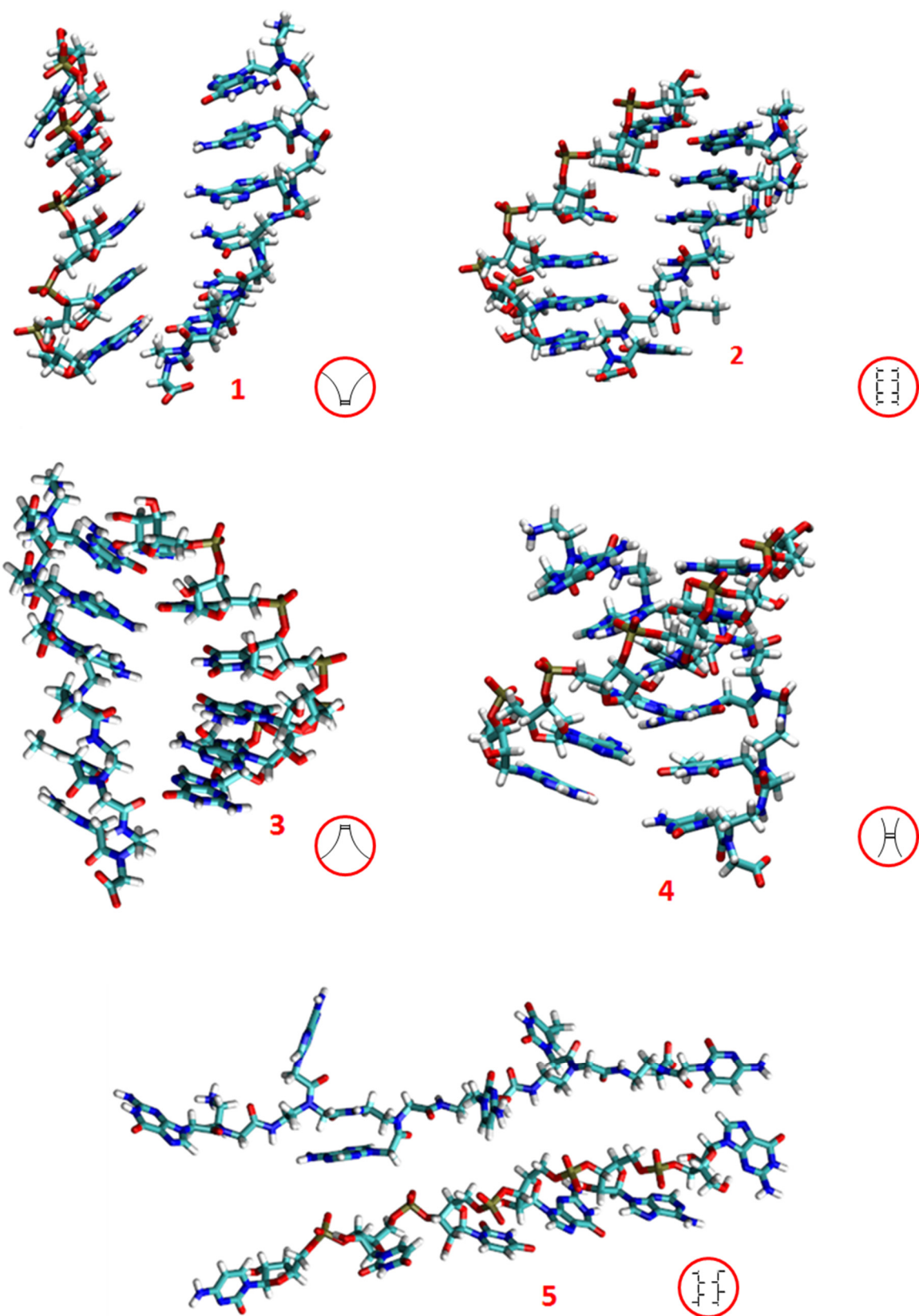
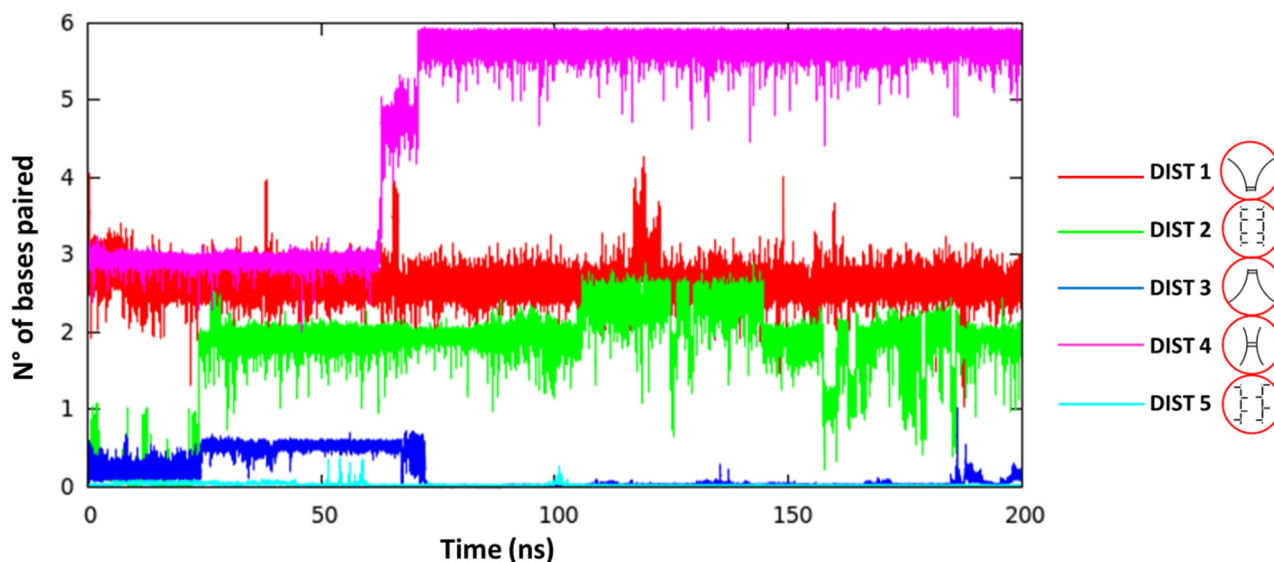


Figure 3.11: Distorted duplex structures: **1)** spread like a fan on CCP:RG5 end; **2)** strands shifted of 5-6 Å; **3)** spread like a fan on NGP:RC3 end; **4)** twisted strands **5)** stretched strands. In red circles the symbolic representation of stress induced in the system are represented.

The re-annealing kinetics is highly dependent on the kind of external stress induced on the duplex. In the ideal case of an infinitely long simulation, all these cases would converge to the same stable duplex structure as investigated in section 3.2. However, for finite simulation times the accessible time scale allows to understand some crucial dynamics only that are responsible for triggering the duplex formation. In order to track the duplex time-evolution we computed the number of bases paired. This parameter was calculated dividing the H-Bonding between base pairs, calculated using the CV “Hydrogen bond” as implemented in the Plumed 1.3 library for each couple, by the maximum possible H-Bonding number (complete base pairing). In Graph 3.9 we plot this function with respect to the simulation time.



Graph 3.9: Number of base paired as function of time for **DIST** systems described above.

Among the five different scenarios, only **DIST4** reaches complete base pairing within 200 ns. The distortion initially induced on this structure brings PNA and RNA strands to be interacting by the central bases only. This has consequences on the pairing mechanism evolving simultaneously toward the two extremes of the duplex. The kinetic of the duplex formation is driven by the H-bonding formation between base pairing and the base intra- and inter-molecular stacking. The constrain of the central bases paired in the structure of **DIST4** allows to obtain the right balance between H-Bonding formation and stacking for the next bases. **DIST1** and **DIST3** initially present the same base pairing number and interaction are in both cases located at the PNA extremity (respectively C-terminus and N-terminus). However, **DIST1** retain a constant number of bases paired during all the simulation, while **DIST3** immediately loses initial favorable interactions and turned

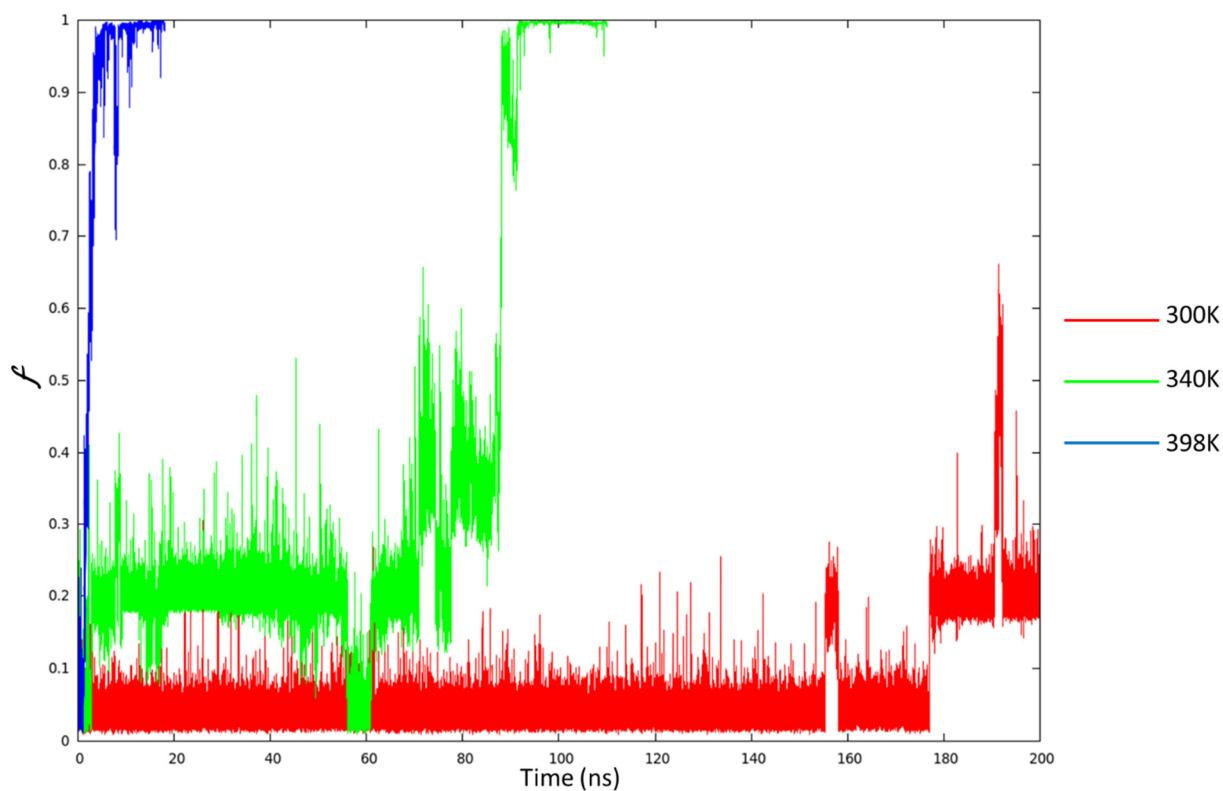
into a structure completely mispaired (Graph 3.9, red and blue traces). This asymmetric behavior is probably due to the specific sequence considered (GAACTC): the first half of the duplex presents a total of 7 hydrogen bonds while the second half 8, slightly favoring C5' edge for an initial interaction. In **DIST2** the two strands are initially shifted of about 5 – 6 Å which means that H-bonding interactions are almost zero. However, the systems readily forms inter-molecular stacking interactions that allow after about 25 ns to establish a partial H-bonding network. **DIST5** is a limit case in which the two strands start from a completely disordered situation. Without favorable interactions, the competition between H-bonding and stacking leads to a very slow formation of the duplex and within 200 ns pairing is almost zero.

The interaction models considered in these simulations are not certainly sufficient to describe all possible ways of contact between the two strands. However, this study demonstrates that the orientation of the interface between bases are crucial; central bases exhibited unique properties in favoring duplex formation, suggesting a possible mechanism of formation of the duplex starting from this position.

3.5.2. Duplex melting

The backward process of duplex re-annealing is the dissociation (i.e. *melting* process). Experimentally, this is achieved increasing the temperature up to a value that is conventionally called melting temperature T_m , that is the temperature at which half of the duplex population is still associated and the other half is not.²⁶⁵ The experimental data are normally interpreted as derived from a two-state model: duplex and dissociated single strands, without intermediate states, due to the highly cooperative effects of binding interactions. The cooperative effect is attested by the sigmoidal curves obtained by spectroscopic techniques (UV-CD) upon heating of the duplexes.²⁶⁶ T_m is generally used as a measure of duplex stability, though in principle it only describes the thermal stability of the duplex. This experimental parameter only gives comparative information on the stability of different duplex, but no information about the mechanism behind the dissociative process induced by the temperature. In order to provide a molecular description we performed several MD simulations on the same duplex PNA:RNA, described in section 3.2, at different temperatures (i.e. between 333K-398K). We define f as the ratio of broken hydrogen bonds with respect the total presented by the system after thermal equilibration. This value ranges between 0, when the duplex is optimally paired, and 1, when the two PNA and RNA single strands are non-

interacting through hydrogen bonds. In Graph 3.10, we report the f value for three representative cases simulated at 300, 340 and 398 K.



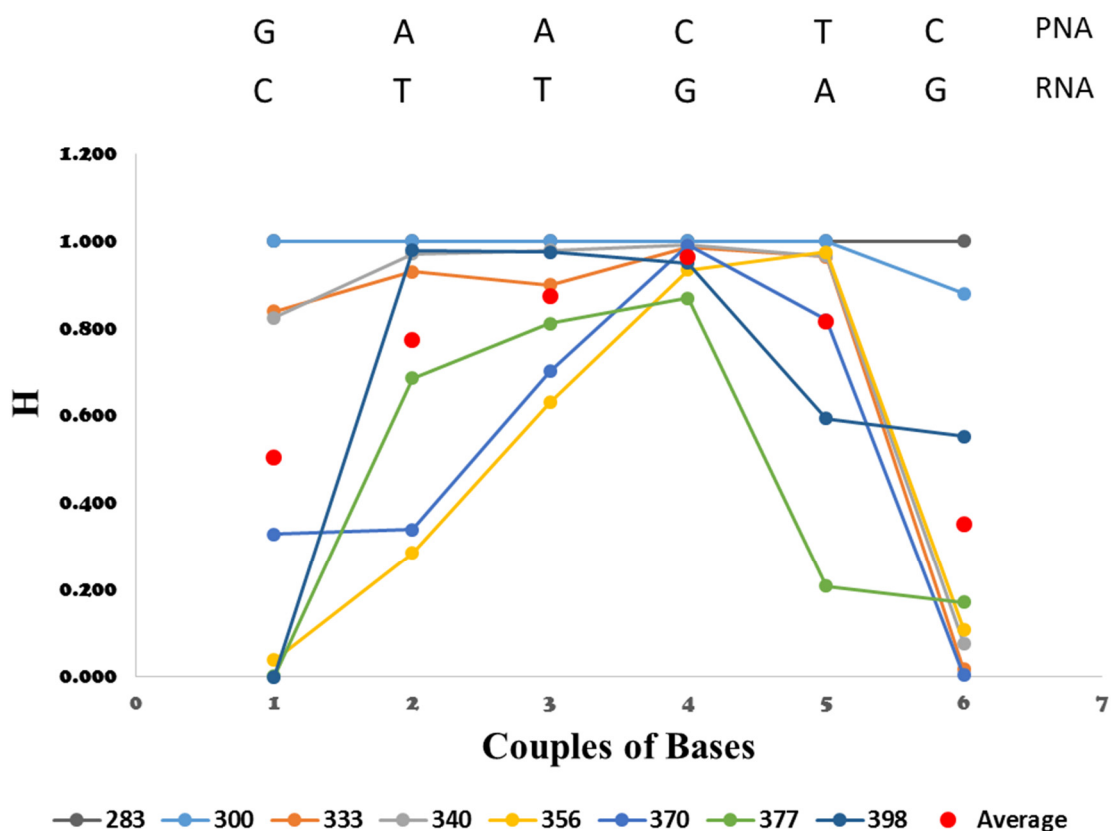
Graph 3.10: f factor as function of time at different temperatures.

This duplex at 300K is stable and only in the very last part of the simulation we observe pair of bases switching from hydrogen-bonded to stacked state, increasing mildly the f value. At 340K we observe an intermediate case in which thermal denaturation starts at the very beginning of the simulation and it remains quite ineffective until 80 ns, when the system shows a sudden increase of f due to the complete opening of the duplex. At 398 K denaturation is very quick and the dissociation occurs in the very first nanoseconds. This temperature dependence was expected and provides important insights to be analyzed in order to extract the contribution of each couple of bases into the denaturation process. Similarly to f , for each base pair we can define the hydrogen bonding parameter H as the ratio between the time being paired and the total simulation time before melting condition (i.e. $f \leq 0.5$). This value gives us the opportunity to analyze the most persistent base pairs in duplex dissociation. In Table 3.3, we report the H parameter calculated for each base pair at the different temperatures analyzed.

Table 3.3: H factor for each base pair as function of temperature.

	283	300	333	340	356	370	377	398
GC	1.000	1.000	0.839	0.825	0.039	0.328	0.002	0.000
AT	1.000	1.000	0.931	0.972	0.284	0.339	0.686	0.980
AT	1.000	1.000	0.900	0.979	0.630	0.702	0.811	0.976
CG	1.000	1.000	0.988	0.991	0.934	0.991	0.870	0.950
TA	1.000	1.000	0.965	0.966	0.975	0.819	0.208	0.593
CG	1.000	0.880	0.017	0.077	0.109	0.004	0.171	0.552

As a reference, we took duplex at 283 K, which is perfectly stable along the entire run as demonstrated by each base pair contributing with an H factor equal to one. By increasing the temperature, the kinetic energy of the system grows along with the probability for base un-pairing. This induces the duplex to dissociate due to the loss of hydrogen bonds and the increasing of entropy. In Graph 3.11 we plot data from Table 3.3 for a clear interpretation of the phenomenon. H factor is on the average lower for those bases placed at the two extreme sides of the duplex while it is higher in the middle. This suggests that the duplex dissociation occurs starting from the terminations and then favoring water invasion on central bases.



Graph 3.11: H factor at different temperatures for each couple of base. PNA sequence is written from N-terminus to C-terminus, RNA sequence is written from 3' to 5'.

These results are complementary and in agreement with the study on re-annealing presented in the

previous section. Moreover, they further suggest the importance of central bases in both formation and dissociation process. A direct consequence of this finding is that in order to improve PNA binding properties, functional modifications must be focused on stabilizing central part of the PNA. The further effect of introducing modifications in different position of the sequence has not been specifically studied, but can be deduced by many experimental data.^{106,267} A complete experimental study on base mismatched as a function of modification positioning²⁶⁸ is available in the literature. In this study, on a 17 mer PNA:DNA duplex the authors showed that a single mismatch in the central part of the PNA leads to a destabilization that causes a drop in the melting temperature up to 25.7 °C, while mismatches in terminal positions destabilize much less the system (range 0.4 – 10 °C).

3.6. Conclusions

Crystal structures represent an important point of reference for the study of PNA-related systems. However, they often offer a static point of view of the problem, while these types of compounds are extremely dynamic and can adopt different conformations. In this chapter, we presented a novel approach using Molecular Dynamics and Metadynamics that permitted to focus on molecular properties of the systems and to observe evolution and adaptability of PNAs to different conditions. Unmodified single-stranded PNA (**ss-PNA**) presented a great flexibility, populating several conformations: stretched conformations with low stacking, stabilized by entropic factors and closed conformations with high staking, stabilized by enthalpic factors. This can explain unique binding properties of these compounds that can adapt to the substrate to bind. As expected, **γ -ss-PNA** presented a reduced flexibility compared to unmodified one. However, this system is not rigid, having the possibility to populate different states and not, as previously proposed, fixed in a rigid helix disposition. Nonetheless, among these well-defined conformations we were able to identify chirally organized structures that could justify the experimental CD signals and a smaller number of populated conformations, which account for the higher binding capability of γ -PNA compared to unmodified PNA discovered experimentally.

Our simulations on PNA:RNA interactions suggested a possible mechanism of formation for the duplex which starts from central bases pairing. Simulated annealing and melting processes both highlighted the importance of these bases for duplex formation and stability. Thus functional modifications, aiming to reinforce sequence selectivity, should be designed by focusing in the central part of PNA strand and must either favoring kinetics of duplex formation or stabilizing base pairing through extra interactions.

4. Synthesis and study of “Adenine-clamp” base

4.1. Triplex structure and TAT mimicking base

As discussed in Chapter 1, usually PNA binds to DNA forming a helix similar to DNA:DNA duplex. When the target sequence is homo-purine and the PNA sequence is homopyrimidine, a triplex structure, in which two PNAs bind one DNA strand using both Watson-Crick and Hoogsteen bonds, can be formed.

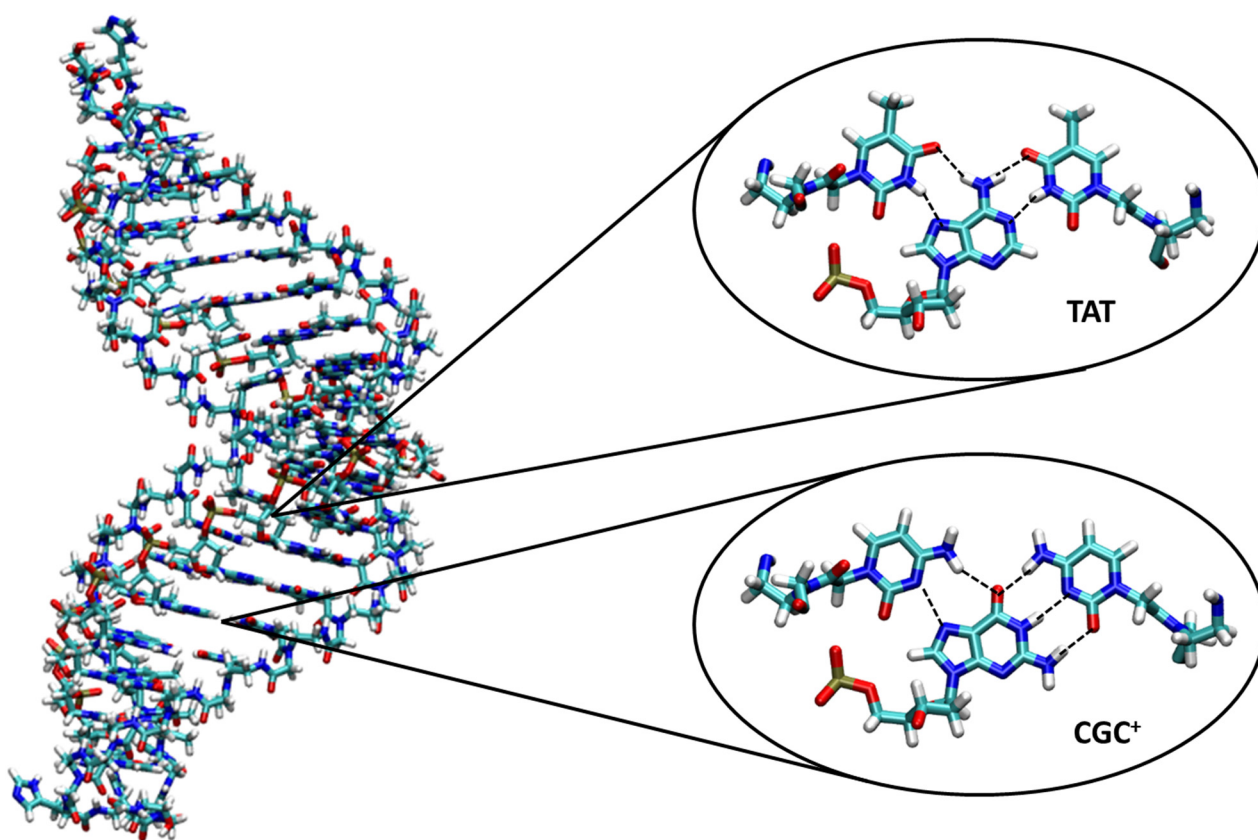


Figure 4.1: Crystal structure of a triplex taken from PDB (1PNN) and enlargement of a TAT and CGC⁺ triplet.

In Figure 4.1 an example of PNA:DNA:PNA triplex taken from the Protein Data Bank (1PNN)¹⁴⁰ is shown, displaying the typical H-bonding patterns described above. Triplexes are very stable structures showing high melting temperatures. By controlling the pH it is possible to observe, upon melting, the transition from triplex to duplex and then to single strand or directly from triplex to single strand;²⁶⁹ the pH dependence is primarily due to the protonation of cytosine needed for the CGC⁺ triplet; under optimal conditions, the binding of the two strands is cooperative, thus the

binding of one strand favors the binding of the second one by Hoogsteen base pairs. The distance between methyl groups of thymine in triplex structure is roughly the same in all TAT triplets and has a mean value of 9.5 Å. In a previous PhD thesis work,²⁷⁰ this allowed to design a dimeric base (figure 4.1) by connecting two uracil moieties with an opportune linker based on 2,7-bis(aminomethyl) naphthalene; this was chosen as a bridge because it can in principle allow to obtain a distance between the methylene groups protruding from uracils of about 10 Å, similar to the distance of the thymine methyl groups in the triplex structures. A rigid spacer like naphthalene is necessary also to avoid collapse of the uracil moieties by formation of intramolecular hydrogen bonds.

4.2. Dimeric uracil base: testing previously reported synthetic route

In Figure 4.2 the structure of the PNA monomer bearing the previously designed dimeric base is reported. Synthesis of such modified compound can be achieved by four different synthons, depicted in Figure 4.2 and represented with different colors.

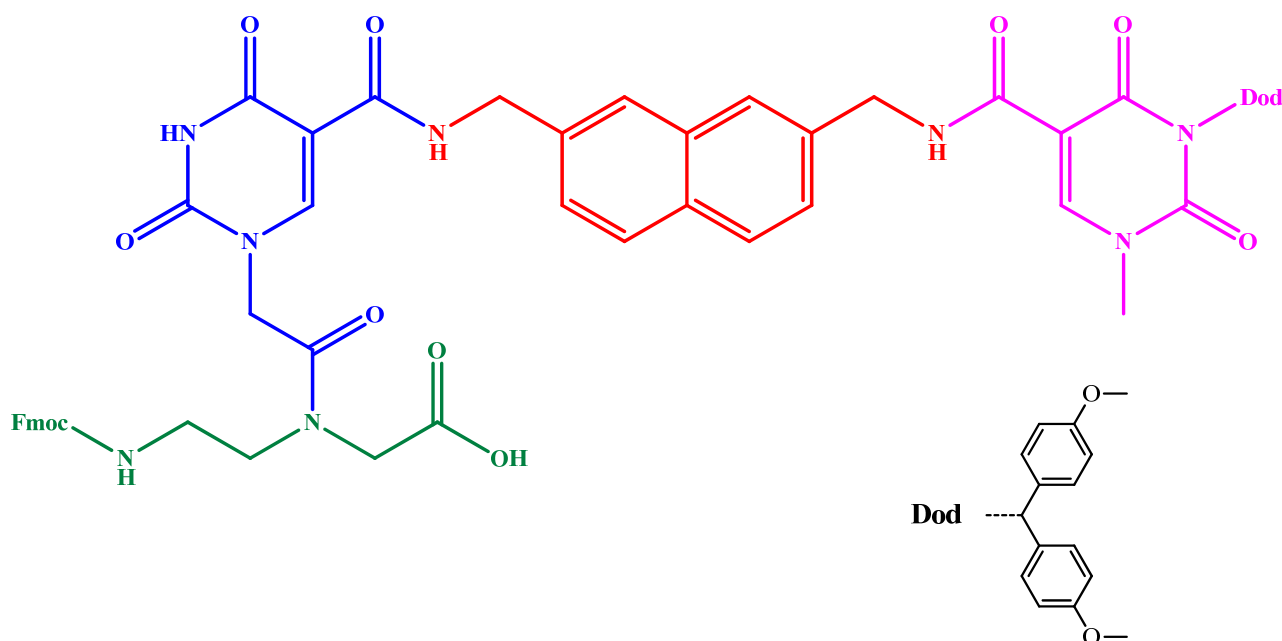
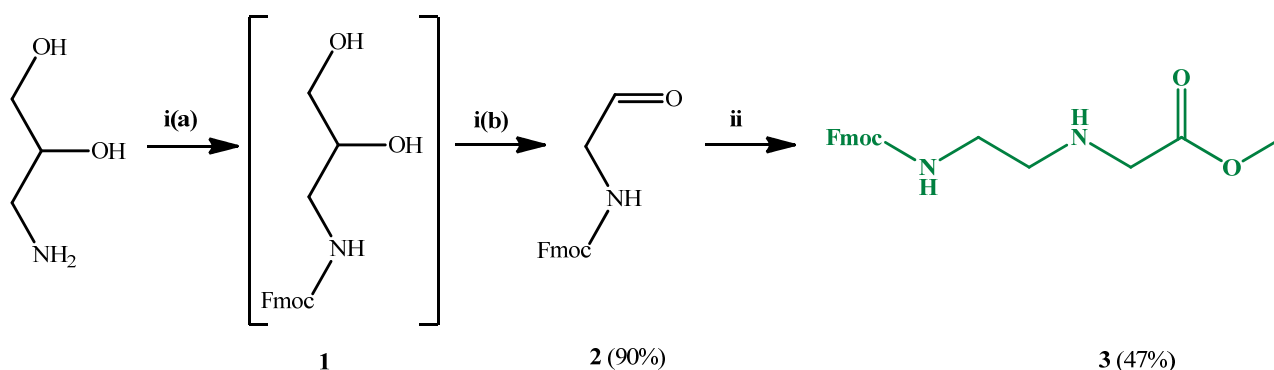


Figure 4.2: Structure of the dimeric base previously designed.

In this thesis, the steps previously used for the synthesis of constituent units²⁷⁰ were checked and optimized, where possible.

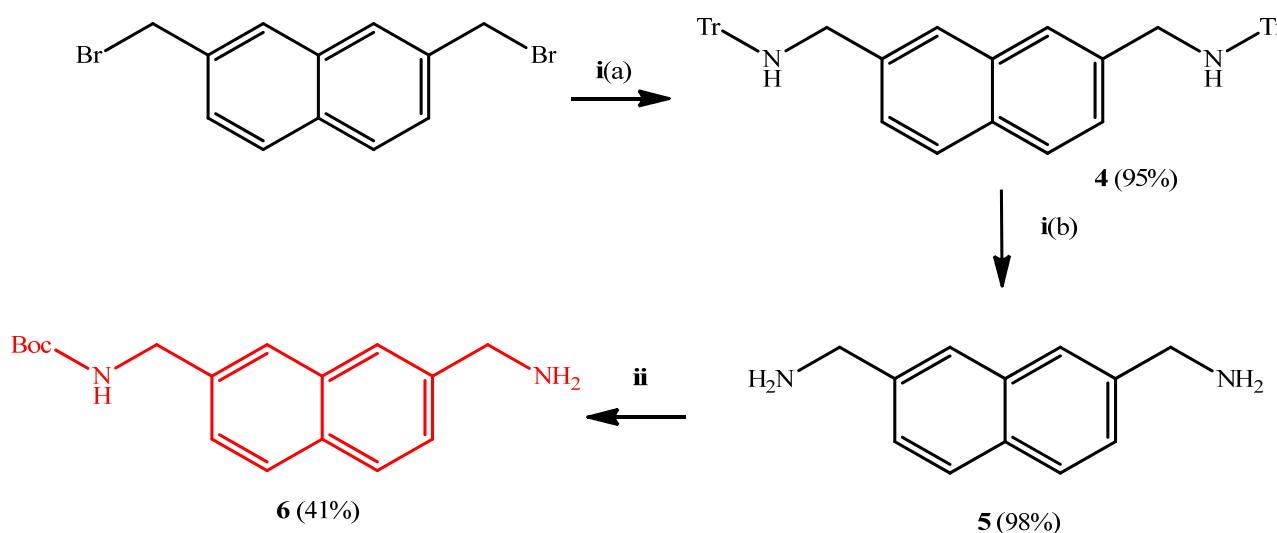
The synthesis of the PNA backbone (Scheme 4.1) was achieved starting from 3-amino-1,2-propanediol which was Fmoc-protected to give the intermediate **1**, which was readily oxidized to

the aldehyde **2** using mild conditions (KIO_4). The protected backbone **3** was obtained with reductive amination of **2** with glycine methyl ester.²⁷¹



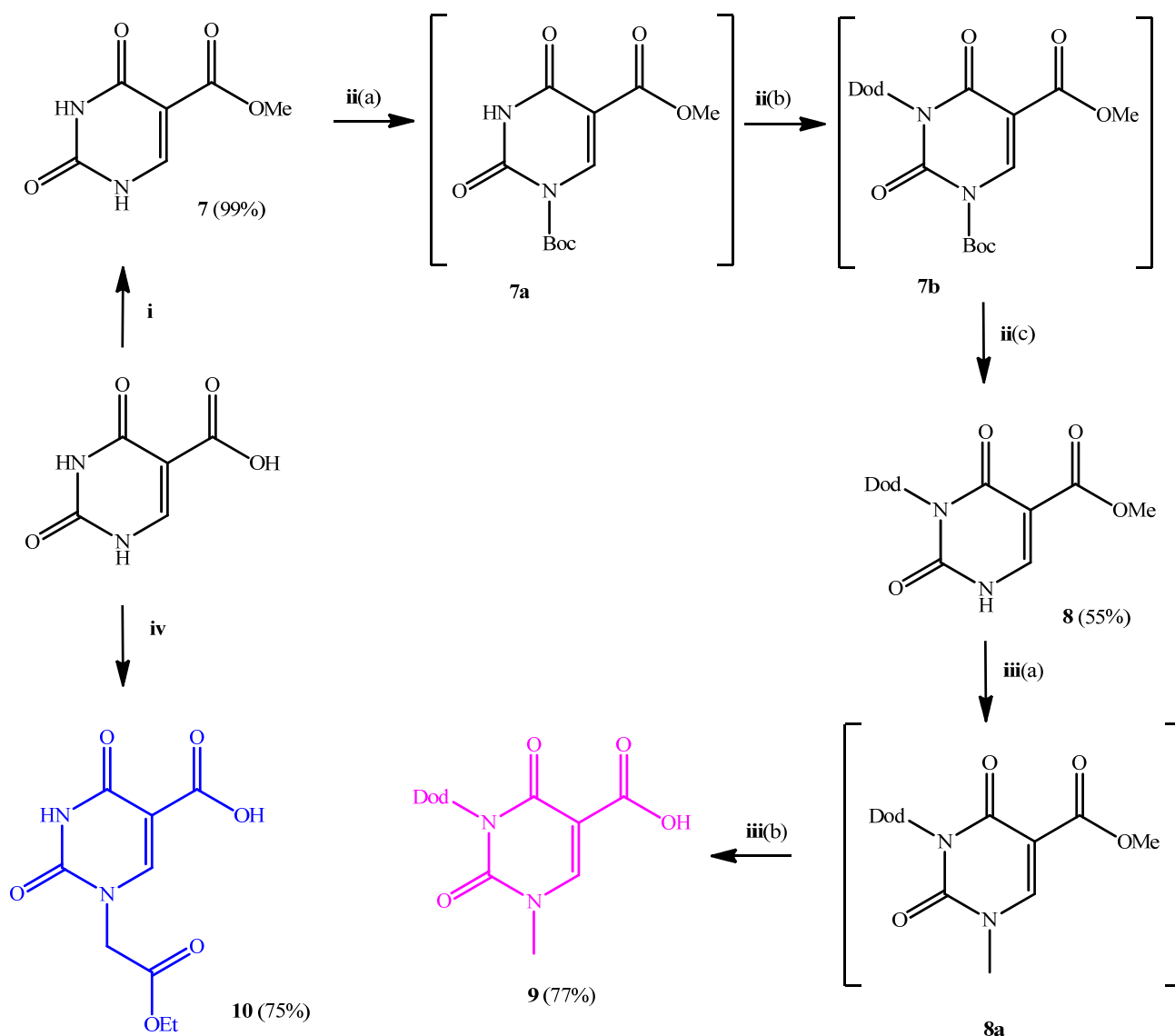
Scheme 4.1: Synthesis of the Fmoc-aeg-OMe backbone. (i) [(a) Fmoc-OSu, Na_2CO_3 , dioxane, water, 75 min, rt; (b) KIO_4 , acetone, water, 12h, rt]. (ii) [GlyOME, NaBH_3CN , MeOH, 6h, 0°C to rt].

The synthesis of the central linker (Scheme 4.2) was carried out starting from 2,7-bis(bromomethyl)naphthalene that underwent a nucleophilic substitution with tritylamine.²⁷² Amine moieties were obtained by deprotection of trityl group under acidic conditions. In order to couple two different uracil units to this spacer, unsymmetrical protection of the two amino groups was needed; this was achieved by mono protection with Boc, using a mild Boc donor phenyl-*t*-butylcarbonate (BocOPh), which guaranteed better yields compared to $(\text{Boc})_2\text{O}$.



Scheme 4.2: Synthesis of the mono protected linker. (i) [(a) Tr-NH_2 , K_2CO_3 , MeCN, 96h, 55°C; (b) TFA, DCM, MeOH, 40 min, rt]. (ii) [BocOPh, EtOH, overnight, reflux].

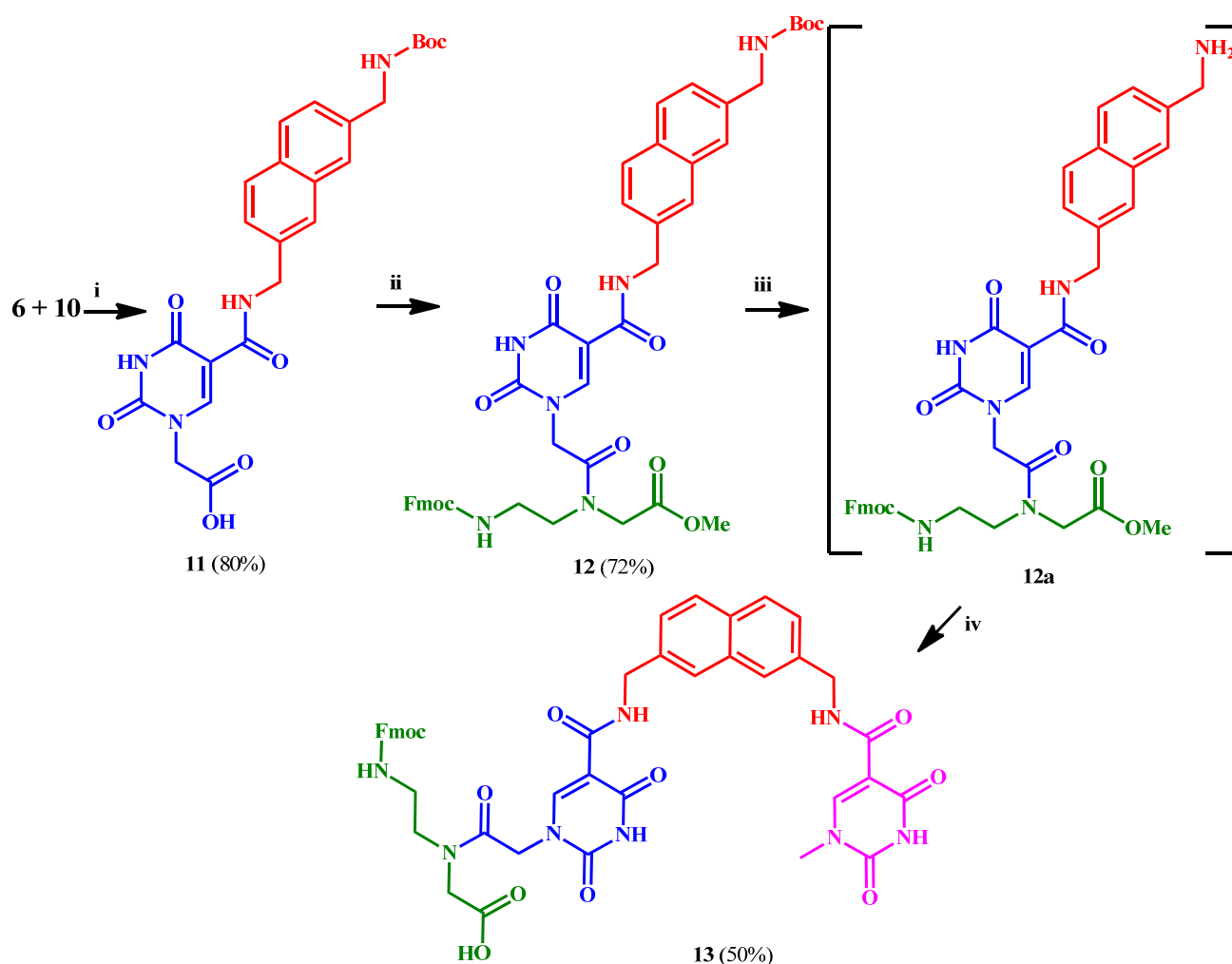
The two different uracil moieties were synthesized starting from isoorotic acid (Scheme 4.3). Selective methylation of N1 is difficult to achieve directly due to high reactivity of alkylating agents and to small steric hindrance of the methyl group. After esterification of isoorotic acid, alkylation proceeded then through Boc-protection of N1 nitrogen. The crude product **7a** was treated with Dod-Cl, thus protecting N3 position. N1 Boc showed an enhanced reactivity due to electron withdrawing effect of the uracil ring, allowing its removal under basic conditions.



Scheme 4.3: Synthesis of Uracil moieties. (i) [SOCl₂, MeOH, overnight]. (ii) [(a) Boc₂O, DMAP, MeCN, 5h, rt; (b) Dod-Cl, NaH, DMF, overnight, 0°C to rt; (c) K₂CO₃, MeOH, 2h, rt]. (iii) [(a) MeI, K₂CO₃, MeCN, 4h, rt; (b) 1M NaOH, MeOH, 2h, rt]. (iv) [(a)HMDS, TMS-Cl, 4h, reflux; (b) Br-CH₂COOEt, o.n., reflux; (c) water/AcOH 1:1, 20 min, rt].

The intermediate **8** was then methylated and final product **9** obtained for hydrolysis of methyl ester. **10** was easily obtained through direct alkylation of silylated isoorotic acid with ethyl bromo acetate.²⁷³

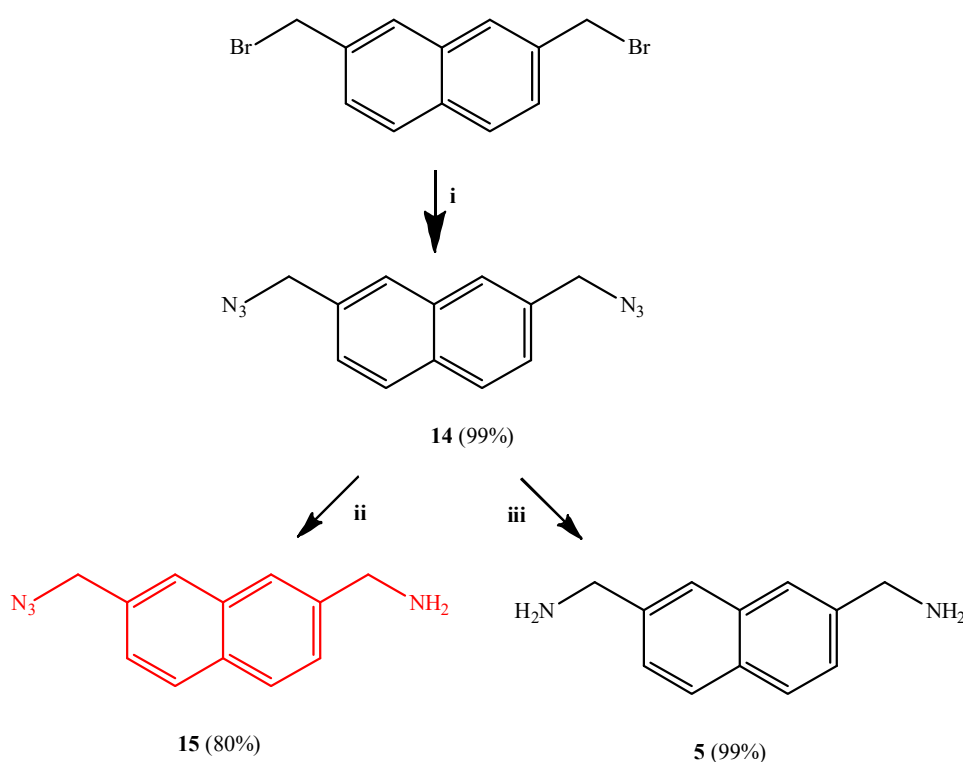
Linker **6** was coupled with uracil **10** using HBTU and then hydrolyzed to obtain **11**. This was coupled with backbone **3** using DCC/DhBtOH to afford **12**. Derivative **12** was deprotected using TFA and intermediate **12a** was coupled with **9** using HBTU. The final product **13** was obtained after hydrolysis using Ba(OH)₂ and controlling reaction time in order to avoid Fmoc deprotection (Scheme 4.4).



Scheme 4.4: Synthesis of dimeric base monomer. (i) [(a) HBTU, DIEA, DMF, 4h, rt; (b) 2M NaOH, THF, 90 min, rt]. (ii) [**3**, EDC, DhBtOH, DIEA, DMF, overnight, 0°C to rt]. (iii) [TFA/DCM 4:6, 1h, 0°C to rt]. (iv) [(a) HBTU, DIEA, DMF, 4h, rt; (b) Ba(OH)₂, THF, 1.5h, rt].

4.3. Dimeric uracil base: new synthetic strategies

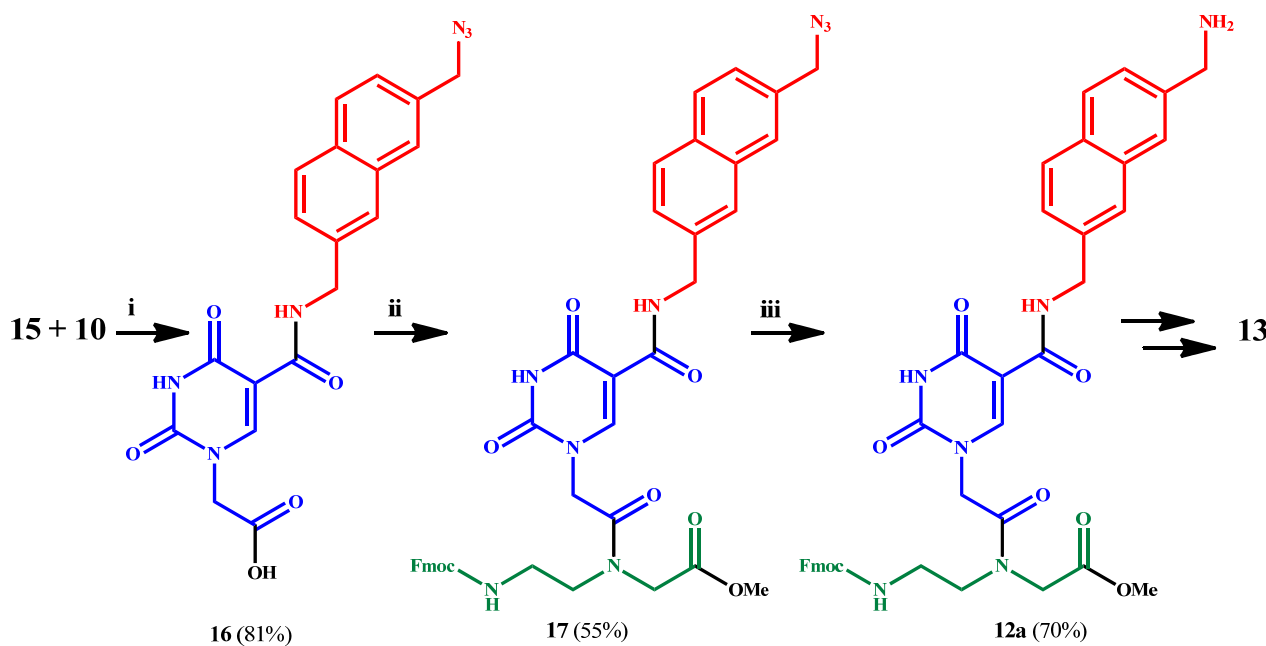
In the schemes of reaction described above, desymmetrization of central linker was made by mono protection of an amino group with Boc. Despite of the reduced activity of BocOPh, desired product was achieved in slightly more than statistical distribution; thus, we revised the synthetic strategy to allow an improved overall yield, using 2-methylazide-7-methylammino naphthalene as linker for obtaining final monomer **13** or for new solid phase strategy. These new synthetic routes were developed under the supervision of Dr. Alex Manicardi.



Scheme 4.5: synthesis of azido-amino linker. (i) [NaN₃, DMF, r.t., 30']; (ii) [PPh₃ in Et₂O, 1M HCl/AcOEt 1:1 (then THF/H₂O 1:1), r.t., 5h + 3h + o.n.]; (iii) [PPh₃, H₂O, THF, r.t., o.n.].

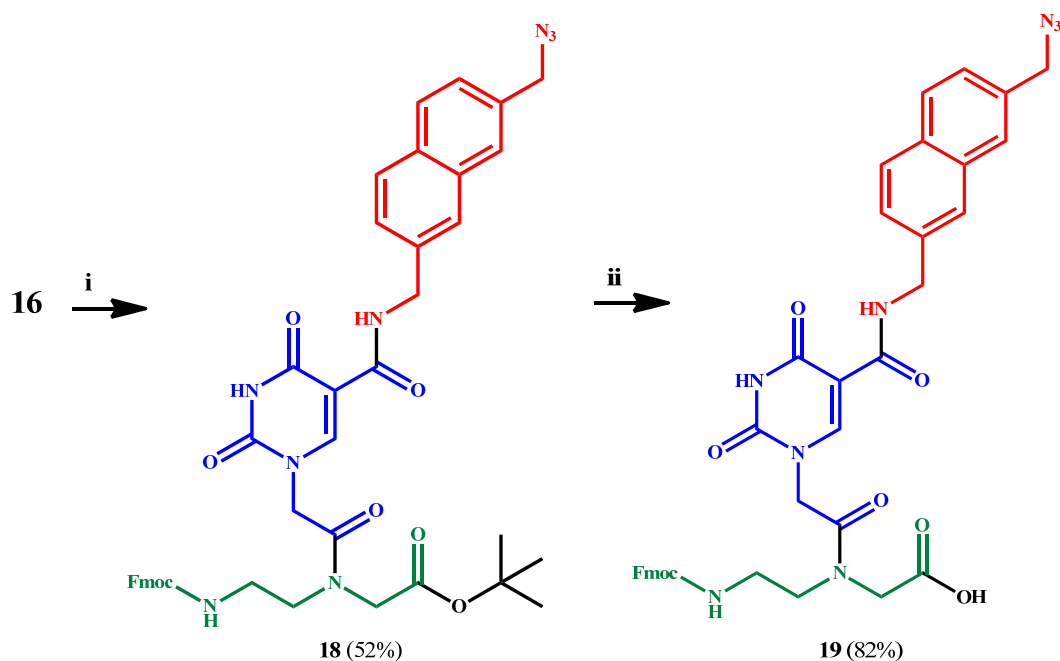
2,7-bis(bromomethyl)naphthalene readily reacted with sodium azide to give the product **14**. Sodium azide is much cheaper than tritylamine, leading in the same way to quantitative reaction. The key point is also in this case desymmetrization of intermediate (Scheme 4.5). Azide can be efficiently reduced to amine using Staudinger reaction.²⁷⁴ The reaction proceeds through formation of phosphazide, which decomposes to an imminophosphorane (phosphazene), generating N₂. Reaction of the imminophosphorane with water leads then to desired amine product and to formation of phosphine oxide²⁷⁵. In order to obtain reduction of only one azide group, reaction was performed

in a biphasic system using a 1M water solution of HCl and ethyl acetate. Under vigorous stirring, intermediate **14** was dispersed in the biphasic solution and triphenylphosphine, dissolved in Et₂O, was slowly added. Reaction between azide and phosphine occurred in organic phase, and as soon as the imminophosphorane was reduced to amine the latter was protonated, due to acidic conditions, subtracting product from organic phase and avoiding the further reaction leading to the double reduction byproduct. In this way it was possible to obtain the product **15** with very high yield (80%). Following this reaction scheme was possible to achieve linker synthesis with a yield double of that obtained by the previous pathway (from 38% to 79%). Intermediate **14** can be used also in alternative synthesis of substrate **5**, through reduction of both azide groups. Compound **15** is useful since it presents a masked amino group that can be used for the synthesis of **13** using a modified version of the previous synthesis (Scheme 4.6).



Scheme 4.6: Synthesis of methyl/tert butyl intermediates. (i) [(a) HBTU, DIEA, DMF, 4h, rt; (b) 2M NaOH, THF, 90 min, rt]. (ii) [3, DCC, DhBtOH, DIEA, DMF, overnight, 0°C to rt]. (iii) [PPh₃, H₂O, THF, r.t, o.n.].

Indeed product **15** was coupled with **10** using HBTU to obtain **16**, after ester hydrolysis. Coupling with backbone led to **17**, a product analogue to **12** shown in Scheme 4.4. The azido group can then be reduced by Staudinger reaction obtaining the intermediate **12a**. Reactions to obtain final monomer are then the same discussed above.



Scheme 4.7: (i) [Fmoc-aeg-O^tBu, EDC, DhBtOH, DIEA, DMF, overnight, 0°C to rt]. (ii) [TFA, DCM, 0 °C to r.t., 30 min].

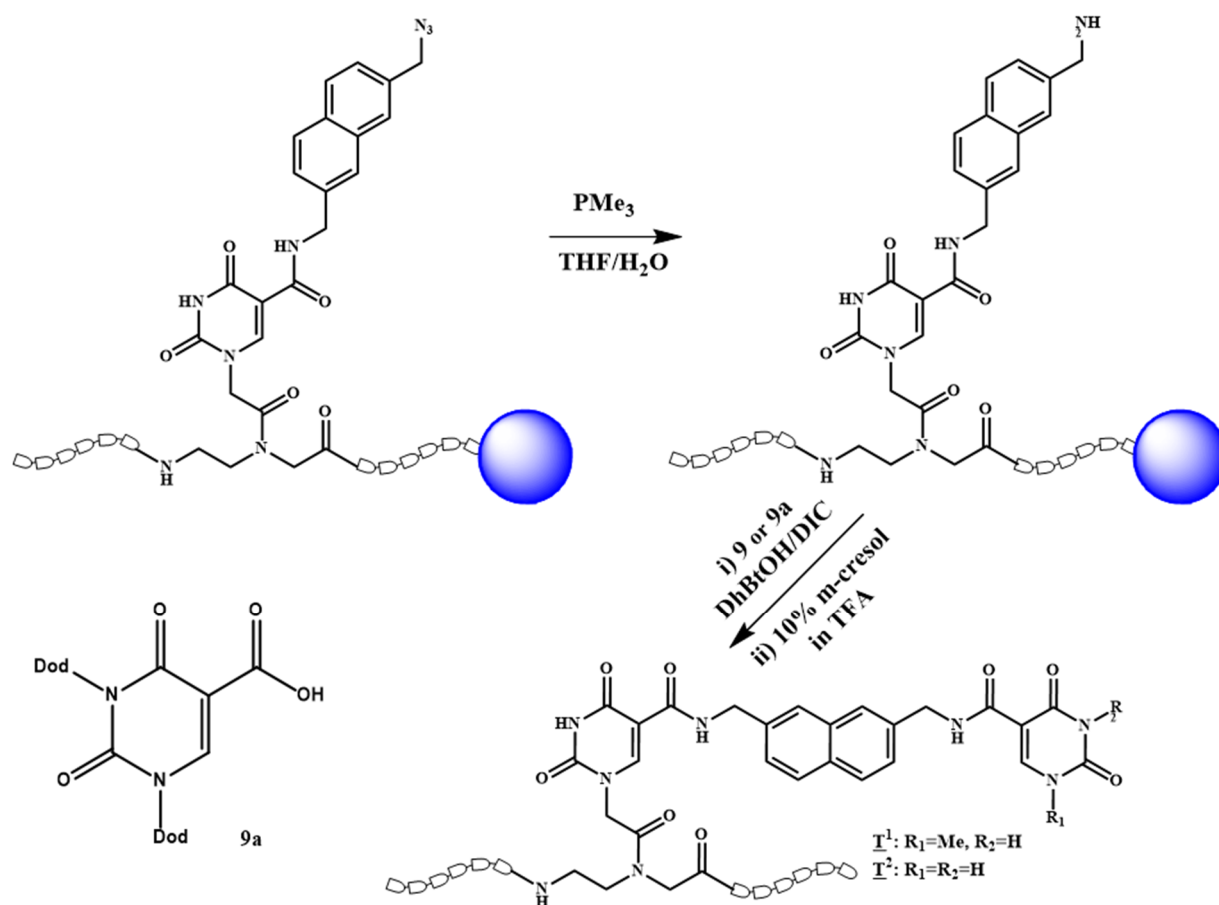
The intermediate **16** was coupled also with commercially available *N*-[2-(Fmoc-amino)-ethyl]glycine tert-butyl ester to give **18** (Scheme 4.7) that was then hydrolyzed to obtain compound **19**. This monomer can be used for a solid phase approach, in which second uracil is coupled directly on resin after PNA chain completion. Details about this synthetic strategy are reported in the following section (Scheme 4.8).

4.3.1. On resin assembly of the dimeric base.

The use of the monomer **13** in PNA solid-phase synthesis give rise to poor yields, due to high steric hindrance of the modified monomer; an alternative route can be to use the precursor **19** for the PNA synthesis, and then to obtain the dimeric base by insertion of the second uracil unit after the completion of the synthesis of the entire PNA sequence. Since we wanted initially to determine if self-association could be induced only by presence of A_T doublet, we specifically designed a sequence containing this doublet without possibility of any other complementarity: H-TTA_TCC-Gly-NH₂. The synthesis of this PNA oligomer was performed using Fmoc strategy. Monomer **19** was inserted in the growing sequence of PNA using a longer coupling time (overnight instead of 30'). After insertion of modified monomer the sequence was completed using standard protocols. Fmoc was not removed at the end of the synthesis in order to allow the attack the second uracil moiety

on the solid phase (Scheme 4.8) without competing reactions. The azide group was reduced performing a Staudinger reaction on the solid phase, in which a crucial point was the use of trimethylphosphine in THF/water. Trimethylphosphine was chosen in place of commonly used triphenylphosphine due to its reduced steric hindrance and thus favoring access to reactive sites. Despite these measures, reduction of azide resulted difficult due to a formation of a biphasic system in the presence of water. Further studies suggested the possibility to reduce this problem by vigorously stirring the solution before adding it into reaction vessel. The reduced PNA was coupled with second uracil moiety using DhBtOH/DIC. We used two different uracils, compound **9** and compound **9a** (Scheme 4.8), which was obtained as byproduct of **8**.

Cleavage from the resin with 10% m-cresol in TFA allowed to recover PNA and, at the same time, to remove Dod protective group. The final products were obtained by two purification cycles with HPLC. First, the Fmoc-protected PNA was purified on RP-HPLC, then the Fmoc group was removed and a second purification was carried out, using RP-HPLC with different conditions.²⁷⁶ Details are reported in experimental section.



Scheme 4.8: Solid phase synthesis of dimeric bases. Symbols in the chain represent PNA monomers. \underline{T}^1 derives from coupling with **9** (PNA1, PNA3) and \underline{T}^2 with **9a** (PNA2).

Due to problems in the reduction step described above, the two PNAs were obtained with low yields (1.1% in case of $\underline{\mathbf{T}}^1$ and 0.5% in case of $\underline{\mathbf{T}}^2$). Modifications of this solid phase protocol, in particular refining Staudinger reaction conditions, must be introduced in future in order to achieve better yields. We synthesized also two unmodified PNA as control (**NM1**, Yield = 12%; **NM2**, Yield = 20%).

4.4. Solution properties of PNAs bearing modified dimeric base

PNAs containing a modified monomer with the dimeric uracil base (**13**) were tested in a previous work in our laboratory.²⁷⁰ The sequences studied were H-GTAGAT $\underline{\mathbf{T}}^1$ CACT-NH₂ (**PR210**), which is a sequence widely used in the literature as a reference for comparing PNA properties as a function of structure,^{106,113} and H-TCCT $\underline{\mathbf{T}}^1$ CACT-NH₂ (**PR212**), which was used in the development of sensing approaches in order to recognize a single point mutation (W1282X) of the cystic fibrosis (CF) gene.^{277,278} The central thymine was substituted with modified monomer $\underline{\mathbf{T}}^1$ in both cases. Preliminary studies revealed tendency to self-association in the case of single strand **PR210**, but not in the case of **PR212** (Figure 4.3 top).

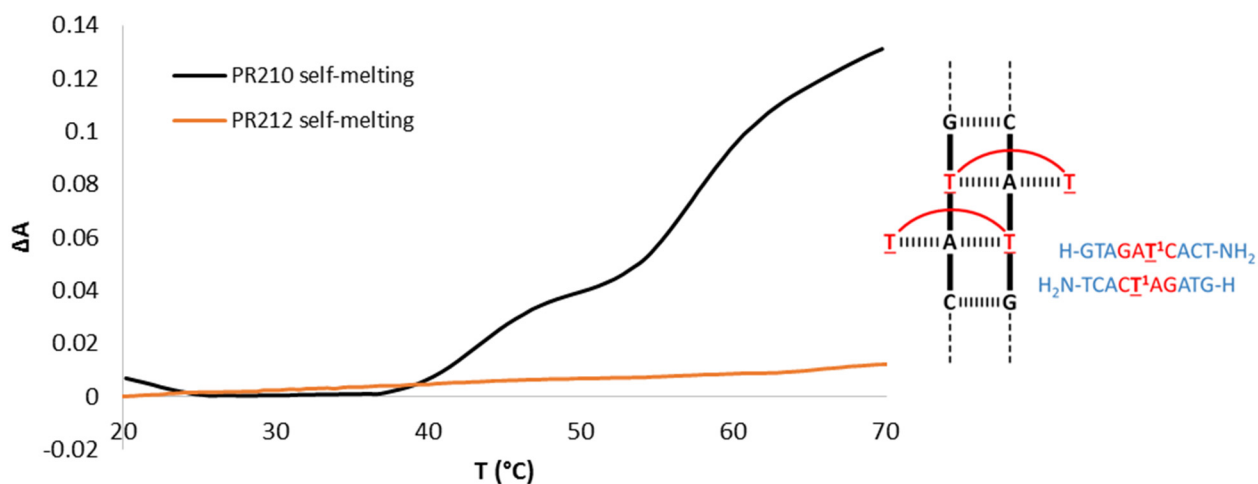


Figure 4.3: Top) UV melting profile expressed as variation of absorbance as function of temperature; Bottom) schematic representation of PR210 self-aggregation. $\underline{\mathbf{T}}^1$ is the modified monomer $\underline{\mathbf{T}}^1$.

Sequence of **PR210** presented actually four complementary bases in the middle of the sequence, but so high melting temperatures are not typical of a 4 mer duplex and were probably due to the presence of dimeric base (Figure 4.3). **PR212** is not affected to this problem of partial complementarity, but contrary to what was expected, it didn't lead to great duplex stabilization (T_m +2 °C compared to unmodified PNA). This system exhibited anyway interesting properties of

selectivity toward single mismatched base.²⁷⁰ We then wanted to better explore properties of PNA bearing this modified monomer using both experimental and computational approach (see next chapter).

To evaluate the presence of auto-association of PNA single strands we performed UV melting experiments on 5 μ M **PNA1** and **PNA2** solutions in PBS buffer (Figure 4.4). Samples were annealed at 90 °C and then cooled at 10 °C.

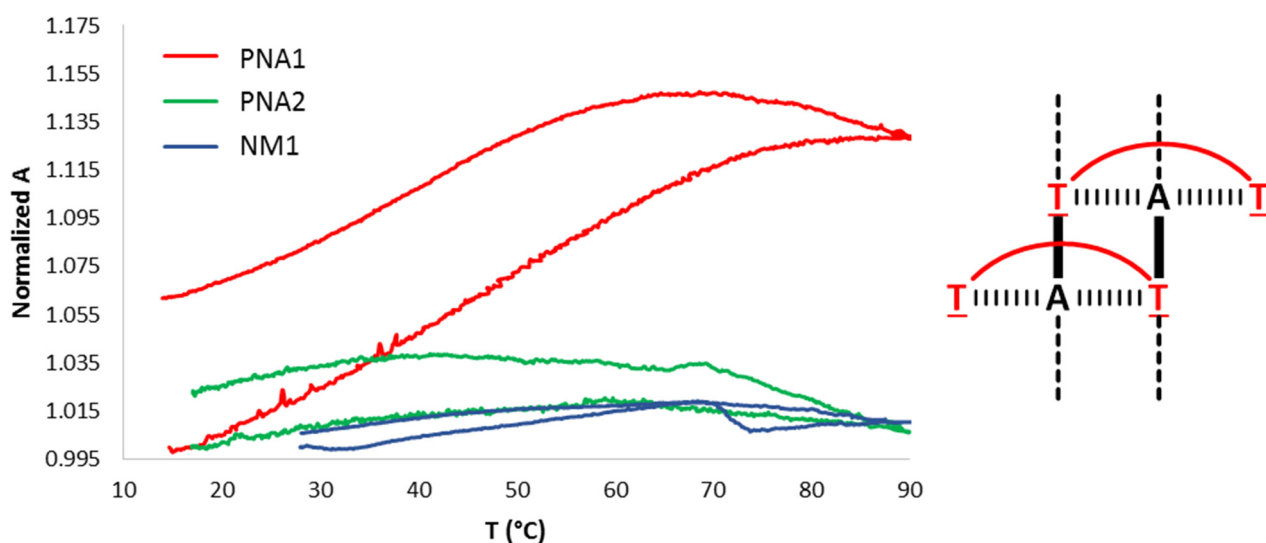


Figure 4.4: Left) UV melting curves of non-modified PNA (**NM1**) and PNAs containing modified base obtained from coupling with uracil **9** (**PNA1**) and **9a** (**PNA2**); right) schematic representation of **PNA1** self-aggregation.

PNA2 and **NM1** presented low absorbance variations as function of the temperature, not correlated to a melting. Irregularity in the shape of the curves was addressable to instrumental problems and not to the samples, as verified by several tests on different samples. **PNA1** melting temperatures were comparable in both heating and cooling cycles ($T_m^{up} = 46.8$ °C, $T_m^{down} = 39.2$ °C) and they resulted quite high considering that, in the case of self-association, were due to only two couples of bases (Figure 4.4 right). **PNA2** presented completely different properties compared to **PNA1**, even if the only difference was in the methyl group in N1 position. We supposed that interaction of N1 with solvent molecules could disturb Hoogsteen bonds of the second uracil and steric hindrance of methyl group could reduce water access. A 3D model of this difference is shown in Figure 4.5.

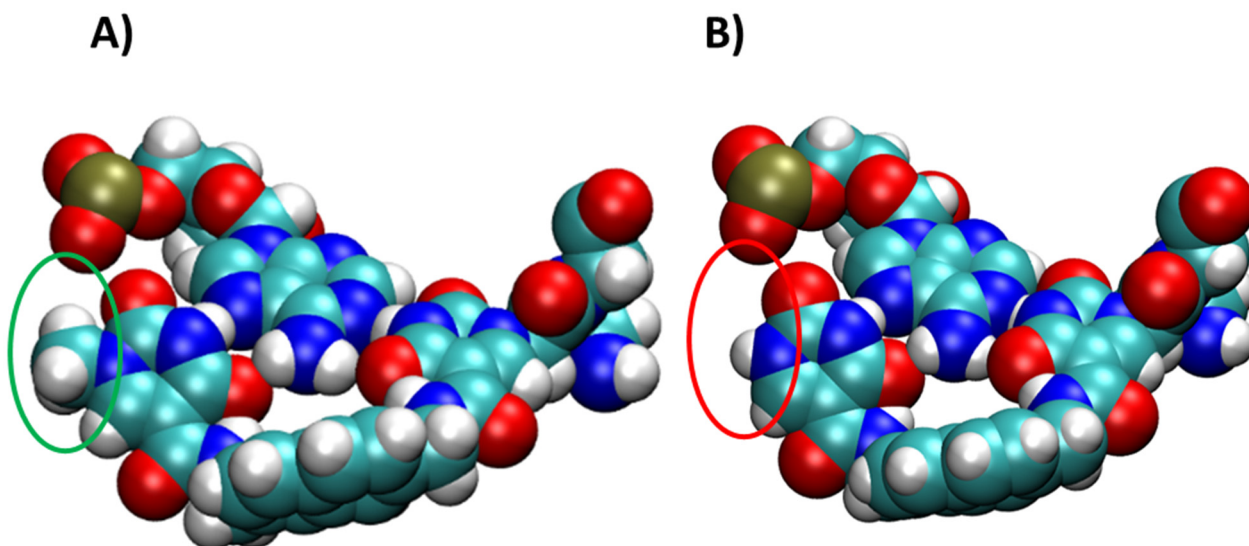
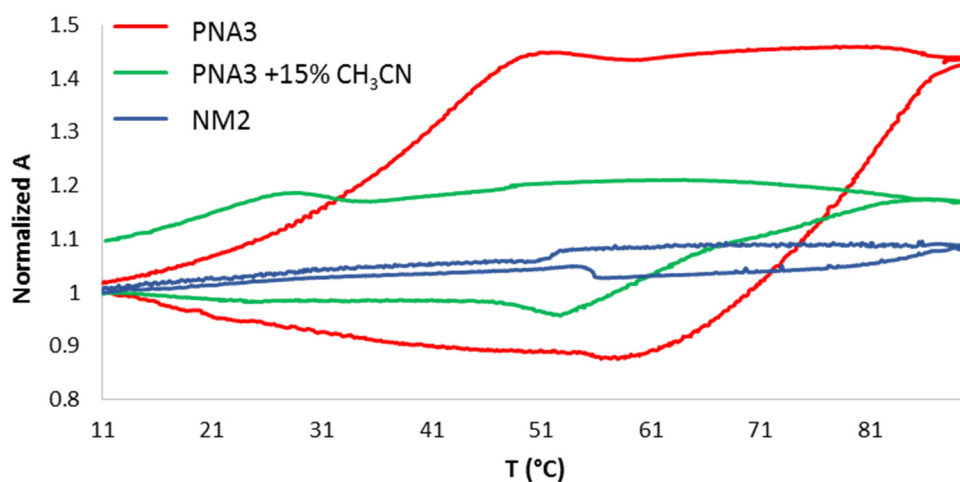


Figure 4.5: Dimeric base bound to adenine. **A)** monomer with uracil N1 protected; **B)** monomer with N1 not protected. Van der Waals radius representation of the atoms allows to recognize a protective function of methyl group attached to N1 compared to simple hydrogen.

We then wanted to observe if the same behavior was presented in a sequence with reversed doublet order (TA). In this case we have studied a sequence complementary to seed region of miR 221 and 222 (H-ATGT¹AGC-Gly-NH₂, **PNA3**). This target has a biological relevance since has been classified as onco-miR^{279,280} in many different pathologies. This system allowed us to study both possible association effects and a real interesting target.

PNA3 was synthesized with the same strategy described above for **PNA1** and **PNA2**, using uracil **9**. The yield was in this case particularly low (0.2%) due to difficulties in purification with HPLC.

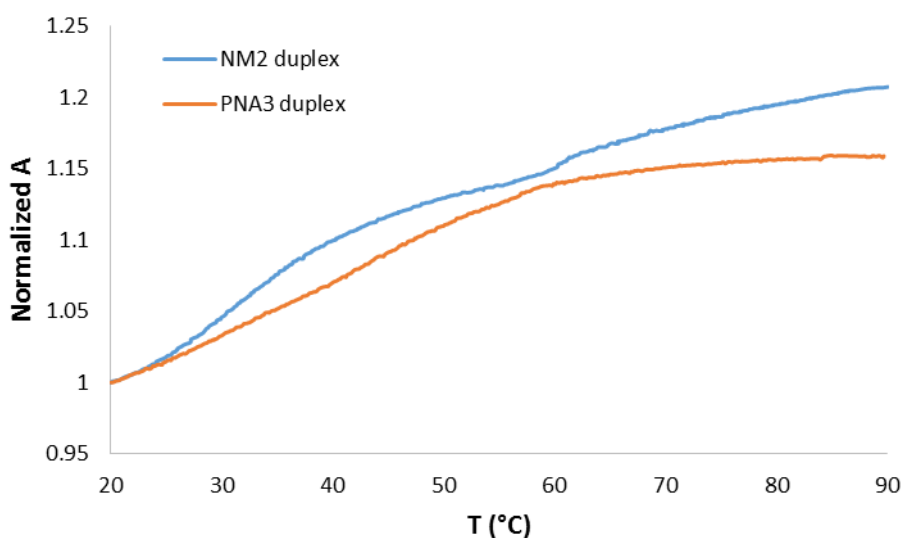
Unmodified PNA (**NM2**), chosen as reference, didn't show any transition and irregular shapes were due, also in this case to instrumental problems (Graph 4.1). **PNA3**, bearing modified base, presented a wide hysteresis compared to **PNA1** with melting temperatures far apart ($T_m^{up} = 82.1$ °C, $T_m^{down} = 42.7$ °C). After the heating-cooling cycle, there was the formation of a precipitate; heating again the sample to 90 °C afforded a complete transparent solution. We therefore believe that in this case there was a sum of effects, solubilization of the dimer and then melting.



Graph 4.1: UV melting curves of non-modified PNA (**NM2**) and PNA containing modified base (**PNA3**) in water and in a 15% solution of acetonitrile.

In order to better solubilize the complex we added 15% of acetonitrile to the sample. In this case solution appeared completely transparent. Melting curve presented a hysteresis similar to that one observed in only water, but of course shifted to lower values due to denaturizing effect of acetonitrile. The first tract of the curve is quite flat compared to the other. We can suppose that, without acetonitrile, dimer precipitates on the walls of the cuvette inducing a local increased absorbance that falls down with solubilization of the complex. After 60 °C, the acetonitrile easily evaporates, probably determining a reduction of this solvent content. Indeed, at the end of the experiment precipitate was found in the cuvette. Anyway, the presence of the hysteresis in the curve showed a low kinetics of aggregate formation.

For these PNAs (**NM2** and **PNA3**) we have studied also association with DNA. In this case, melting temperature of modified PNA is about 5 °C higher compared to unmodified one, confirming a limited, but reproducible, stabilizing effect of the dimeric base (Graph 4.2).



Graph 4.2: UV melting curves of **NM2** and **PNA3** duplexes with complementary DNA.

4.5. Conclusions

We proposed an alternative pathway for the synthesis of compound **13** allowing to achieve better yields for central linker unit. We proposed also a new synthetic strategy for the solid phase synthesis of PNAs containing dimeric base. This protocol must be improved in order to obtain better yields, in particular changing Staudinger reaction conditions, but is more flexible and can allow to introduce multiple variations in the structure starting from the same PNA precursor.

PNAs containing this type of modified monomer confirmed their tendency to self-associate in presence of $A\bar{T}$ doublet in the sequence. Melting temperatures of self-associated species were found to be high if one considers that pairing occurred only on this two couple of bases. Also the PNA bearing reversed doublet order ($\bar{T}A$, **PNA 3**) showed evidences for self-association, though accompanied with low-solubility. This behavior must be considered in target selection, avoiding sequences that presents adenine close to modified thymine or choosing longer sequences in order to have competition between DNA:PNA duplex and PNA:PNA aggregate. However, it is possible to imagine a possible use of this dimeric base as non-covalent, self-assembly unit (Figure 4.6).

Study on PNA:DNA heteroduplex showed a modest stabilization induced by dimeric base compared to unmodified PNA, thus confirming the results previously obtained with a different sequence (**PR212**).

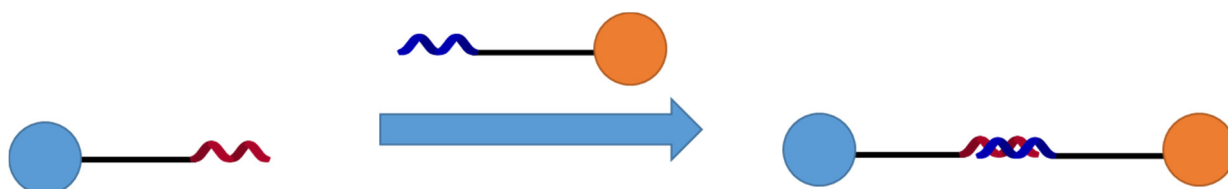


Figure 4.6: Schematic representation of self-assembly induced by $A\bar{T}$ doublet.

Though this stabilizing effect (and even more, the high sequence selectivity) of this dimeric base can be important in biological applications, it was not as large as expected. For this reason, in the next chapter, an attempt to address the design of this system by molecular modelling is reported.

4.6. Experimental section

General. Reagents were purchased from Sigma-Aldrich, Fluka, Merck, Carlo Erba, TCI Europe, LINK and used without further purifications. TLC were run on Supelco 56524-25EA silica gel on aluminium foils with fluorescence indicator 254 nm. Column chromatography were performed as flash chromatography on Merck silica 60 (0.040 – 0.063) under 0.1-0.2 bar of air overpressure. NMR spectra were obtained on Bruker Avance 300 MHz or 400 MHz spectrometers. δ values are expressed in ppm. FT-IR were recorded on Thermo Nicolet 5700. ESI-MS spectra were recorded on a Acquity Ultra Performance LC. Melting points were measured with GALLENKAMP melting point apparatus.

Synthesis and characterization of products from **2** to **13** were carried out using the conditions reported in previous PhD thesis work.²⁷⁰ Yields were reevaluated and were found to be those reported in the results and discussion part.

1,3-bis (bis (4- methoxyphenyl) methyl)-2,4- dioxo-1,2,3,4- tetrahydropyrimidine-5- carboxylic acid (9a). Ester of compound **9a** was obtained as a byproduct of compound **8** synthesis as a part of Boc deprotection of intermediate **7b** (Yield 35%, 417 mg). Hydrolysis was performed by action of 1 M NaOH (aq) and THF 1:1 during 2 h reaction. Final product was precipitated by adjusting pH of solution to 3 with 1 M HCl. White solid was filtrated and dried under vacuum (88%, 408.4 mg).

(9a). ¹H-NMR (300 MHz, 25°C, d₆-DMSO) δ (ppm): 7.87 (s, 1H, CH uracil), 7.16 (s, 1H, CH Dod), 7.14 (d, 4H, J = 8.8 Hz, CH aromatic Dod), 7.07 (d, 4H, J = 8.8 Hz, CH aromatic Dod), 6.97 (d, 4H, J = 8.9 Hz, CH aromatic Dod), 6.84 (d, 4H, J = 8.8 Hz, CH aromatic Dod), 6.80 (s, 1H, CH Dod), 3.77 (s, 6H, OMe), 3.73 (s, 6H, OMe). ¹³C-NMR (100 MHz, 25°C, d₆-DMSO) δ (ppm): 164.2, 161.7, 159.4, 158.6, 150.7, 145.3, 130.8, 130.3, 130.0, 127.8, 114.8, 114.2, 114.1, 113.8, 109.7, 62.9, 58.0, 55.6, 55.5. FT-IR ν (cm⁻¹) 2930, 2837, 1749, 1718, 1627, 1610, 1585, 1511, 1462, 1441, 1419, 1391, 1304, 1248, 1175, 1113, 1031. m.p.: decomposes at 190 – 200 °C.

2,7-bis (azido methyl) naphthalene (14). 2,7-bis(bromomethyl)naphthalene (800 mg, 2.55 mmol) was dissolved in dry DMF (7 ml) and then NaN₃ (347.8 mg, 5.60 mmol) was added to the solution.

After 2.5 h the reaction was completed (TLC on silica gel, Hex-AcOEt 9:1, R_f = 0.43), reaction mixture was diluted with AcOEt (250 ml) and washed with brine (3 X 225 ml). Organic layer was dried over Na_2SO_4 and evaporated under vacuum to afford pure product (light brown solid, 99%, 607 mg).

(14) $^1\text{H-NMR}$ (400 MHz, 25°C , d_6 -DMSO) δ (ppm): 7.98 (d, 2H, J = 8.5 Hz, CH naphth.), 7.94 (s, 2H, CH naphth.), 7.53 (dd, 2H, J_1 = 8.5 Hz, J_2 = 1.7 Hz, CH naphth.), 4.64 (s, 4H, CH_2); $^{13}\text{C-NMR}$ (100 MHz, 25°C , d_6 -DMSO) δ (ppm): 134.29, 133.07, 132.57, 128.81, 127.63, 127.19, 54.16. Compound decomposes in mass; MS (ESI+) m/z : found 406.11, 235.26, 211.6. FT-IR ν (cm^{-1}) 2908, 2852, 2101, 1635, 1510, 1441, 1429, 1370, 1325, 1267, 1175, 1145. m.p. $84 - 86^\circ\text{C}$.

2-azidomethyl-7-aminomethyl naphthalene (15). Compound **14** (300 mg, 1.21 mmol) was dissolved in a mixture of 1 M HCl (150 ml) and AcOEt (75 ml). Solution was vigorously stirred and triphenyl phosphine (313.8 mg, 1.15 mmol), dissolved in diethyl ether (75 ml), was slowly added over a period of 5 h. Mixture was kept stirring for additional 3 h followed by organic solvent removal under vacuum. THF (150 ml) was added and reaction proceeded overnight. Organic solvent was removed and water phase was washed with ether (3 x 150 ml). Water phase was removed and residue dissolved in the minimum amount of 1 M HCl (\sim 100 ml). pH was adjusted to about 12 with NaOH pellets and mixture kept in refrigerator for 2 h. Precipitate (white solid) was collected to afford pure product (138 mg). The remaining product (65.4 mg) was obtained by flash chromatography on silica gel (AcOEt + 1% NH_3 , R_f = 0.2 in AcOEt-MeOH 9:1). (total yield: 80%, 203.4 mg).

(15) $^1\text{H-NMR}$ (300 MHz, 25°C , MeOD) δ (ppm): 7.94 (dd, 1H, J = 1.8, CH naphth.), 7.92 (s, 1H, CH naphth.), 7.87 (bs, 1H, CH naphth.), 7.56 (dd, 1H, J_1 = 8.4, J_2 = 1.8, CH naphth.), 7.51 (dd, 1H, J_1 = 8.4, J_2 = 1.8, CH naphth.), 4.56 (s, 2H, $\text{CH}_2\text{-N}_3$), 4.15 (s, 2H, $\text{CH}_2\text{-NH}_2$). $^{13}\text{C-NMR}$ (75 MHz, 25°C , MeOD) δ (ppm): 128.2, 128.1, 126.8, 126.7, 126.2, 126.0, 54.3, 44.1. MS (ESI+) m/z : $[\text{M} + \text{H}]^+$ calcd. 213.11, found 213.18. FT-IR ν (cm^{-1}) 3348, 3270, 2908, 2853, 2107, 1632, 1560, 1511, 1473, 1440, 1430, 1388, 1352, 1326, 1278, 1171, 1144. m.p. $93 - 95^\circ\text{C}$.

2,7-bis(aminomethyl) naphthalene (5) (alternative synthesis). Compound **14** (301.8 mg, 1.27 mmol) was dissolved in THF (20 ml) in presence of triphenyl phosphine (697.8 mg, 1.267 mmol) and water (114 μl , 6.34 mmol). To complete the reaction, after 4 h, additional amount of triphenyl phosphine (348.9mg, 0.63 mmol) and H_2O (228 μl , 12.67 mmol) were added and mixture stirred overnight. THF

was removed under vacuum, crude dissolved in 2 M HCl (150 ml) and washed with DCM (3 X 100 ml) to eliminate phosphine. Then, pH was adjusted to about 12 and product extracted with DCM (7 x 200 ml). Organic layers were combined, dried over Na₂SO₄ and evacuated under vacuum to afford pure product (99%, 236 mg).

(5). ¹H-NMR (300 MHz, D₂O, 25 °C) δ (ppm) 4.33 (s, 4H), 7.59 (dd, 2H, J₁ = 8.5 Hz, J₂ = 1.7 Hz), 7.98 (d, 2H, J = 1.7 Hz), 8.02 (d, 2H, J = 8.4 Hz). ¹³C-NMR (75 MHz, D₂O, 25°C) δ (ppm) 46.5, 130.2, 131.6, 132.3, 134.5, 136.1, 136.2. Full characterization of this compound is reported in pervious thesis work.²⁷⁰

2-(5-(((7-(azidomethyl)naphthalen-2-yl)methyl)carbamoyl)-2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl) acetic acid (16). Compound **10** (171.2 mg, 0.71 mmol) was dissolved in dry DMF at 0°C, under nitrogen atmosphere. HBTU (268.1 mg, 0.71 mmol) and DIPEA (246.1 μl, 1.41 mmol) were added and stirred at 0 °C for 15 min and at room temperature for further 15 min. Compound **15** (100 mg, 0.471 mmol) was then added and mixture stirred overnight at room temperature. At the end of the reaction, solution was partitioned between saturated KHSO₄ (200 ml) and AcOEt (200 ml), and then organic phase washed with NaHCO₃ and brine. Solvent was evaporated under vacuum and crude dissolved in a mixture of methanol (6 ml) and 1 M NaOH (6 ml). Solution was stirred for 4 h and then pH adjusted to 1 using 1 M HCl. Precipitate was separated on a Büchner funnel and dried under vacuum to afford pure product (81 %, 140 mg).

(16) ¹H-NMR (400 MHz, 25°C, d₆-DMSO) δ(ppm): 13.31 (bs, 1H, COOH), 12.05 (s, 1H, NH uracil), 9.24 (t, 1H, J = 6.1 Hz, NH amide), 8.64 (s, 1H, CH uracil), 7.94 (d, 1H, J = 8.5 Hz, CH naphth.), 7.91 (d, 1H, J = 8.7 Hz, CH napht.), 7.89 (s, 1H, CH napht.), 7.80 (s, 1H, CH napht.), 7.49 (dd, J = 8.0, 1.6 Hz, 1H, CH napht.), 7.47 (dd, J = 8.2, 1H, 1.6 Hz, CH napht.), 4.68 (d, 2H, J = 6 Hz, CH₂NH), 4.63 (s, 2H, CH₂N₃), 4.61 (s, 2H, CH₂COOH). ¹³C-NMR (100 MHz, 25°C, d₆-DMSO) δ(ppm): 169.7, 164.1, 162.2, 152.3, 150.5, 138.0, 133.9, 133.2, 132.1, 128.7, 128.4, 127.5, 126.7, 126.6, 125.8, 105.2, 54.2, 49.8, 42.7. FT-IR ν (cm⁻¹) 3011, 2818, 2125, 1706, 1632, 1605, 1555, 1460, 1420, 1381, 1331, 1268, 1234. Decomposes at 270 – 280 °C.

Methyl 2-(N-(2-(((9H-fluoren-9-yl) methoxy) carbonyl) amino) ethyl)-2-(5-(((7-(azidomethyl) naphthalen-2-yl)methyl)carbamoyl)-2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl)acetamido)acetate (17). Compound **16** (70 mg, 0.17 mmol) was dissolved in dry DMF under nitrogen atmosphere and stirred at 0 °C with EDC (32.9 mg, 0.17 mmol) and DhBtOH (27.9 mg, 0.17 mmol) for 15 min and at r.t. for additional 15 min. Backbone **3** (72.9 mg, 1.2 eq) was then added together with DIPEA (34 μ l, 1.2 eq) and mixture stirred overnight. Solution was diluted with AcOEt (150 ml) and washed with saturated KHSO₄ (2 x 150 ml), saturated K₂CO₃ (2 x 150 ml) and brine (1 x 150 ml). Organic layer was dried under vacuum and crude purified by flash chromatography on silica gel (AcOEt-Hexane 8:2) to afford pure product (55 %, 70 mg).

(17) ¹H-NMR (400 MHz, 25°C, d₆-DMSO) δ (ppm): 12.00 (s, 1H, NH uracil), 9.25 (t, 1H, J = 6 Hz, NH amide), 8.48 (M) and 8.39 (m) (s, 1H, CH uracil), 7.92 (m, 2H, CH naph), 7.89 (d, 2H, J = 7.1 Hz, CH aromatic Fmoc), 7.88 (s, 1H, CH naph.), 7.80 (s, 1H, CH naph.), 7.69 (bd, 2H, J = 7.3 Hz, CH aromatic naph.), 7.49 (dd, 1H, J₁ = 6.5 Hz, J₂ = 1.5 Hz, CH naph.), 7.47 (dd, 1H, J₁ = 6.5 Hz, J₂ = 1.6 Hz, CH naph.), 7.41 (t, 2H, J = 7.4 Hz, CH aromatic Fmoc), 7.32 (td, 2H, J₁ = 7.4 Hz, J₂ = 0.7 Hz, CH aromatic Fmoc), 7.27 (t, 1H, J = 5.9 Hz, NH Fmoc), 7.95 (M) and 4.76 (m) (s, 2H, CH₂ linker), 4.68 (d, 2H, J = 5.8 Hz, Naphth.-CH₂-NH), 4.61 (s, 2H, CH₂N₃), 4.35 (M) and 4.29 (m) (d, 2H, J_M = 6.8 Hz, J_m = 3.6 Hz, CH₂ Fmoc), 4.27 – 4.18 (m, 1H, CH Fmoc), 4.32 (m) and 4.08 (M) (s, 2H, CH₂COOMe), 3.73 (m) and 3.63 (M) (s, 3H, Me), 3.45 (M) and 3.37 (m) (t, 2H, J_M = 6.4 Hz, J_m = 6.8 Hz, CH₂ backbone), 3.27 (M) and 3.09 (m) (q, 2H, J_M = 6.1 Hz, J_m = 6.1 Hz, CH₂ Fmoc). ¹³C-NMR (100 MHz, 25°C, d₆-DMSO) δ (ppm): 170.2 (m) and 169.9 (M), 167.7 (m) and 167.5 (M), 164.1, 162.3, 156.8 (M) and 156.6 (m), 152.6 (M) and 152.4 (m), 150.5, 144.4, 141.2, 138.0, 133.9, 133.2, 132.1, 128.7, 128.4, 128.1, 127.5, 126.7, 126.6, 125.8, 125.6, 120.6, 105.0, 66.0, 54.2, 52.8 (m) and 52.3 (M), 49.2, 49.0 (M) and 48.7 (m), 48.2, 47.4 (m) and 47.2 (M), 42.7, 38.7 (M) and 38.1 (m). MS (ESI+) m/z: [M + H]⁺ calcd. 745.27, found 745.31. FT-IR ν (cm⁻¹) 2818, 2097, 1718, 1687, 1683, 1611, 1538, 1464, 1450, 1375, 1341, 1239, 1213, 1145, 1108, 1003. m.p. 155 – 157 °C.

Tert-butyl 2-(N-(2-(((9H-fluoren-9-yl) methoxy) carbonyl) amino) ethyl)-2-(5-(((7-(azidomethyl) naphthalen-2-yl) methyl) carbamoyl)-2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl) acetamido) acetate (18).^{317,318} Compound **18** was prepared analogously to product **17** but using commercial backbone with tert-butyl ester. Purified with flash chromatography on silica gel (DCM-MeOH 98:2) to afford pure product (52%, 185.4 mg).

(18) $^1\text{H-NMR}$ (300 MHz, 25°C, CDCl_3) $\delta(\text{ppm})$: 9.04 (t, 1H, $J = 9$ Hz, NH amide uracil), 8.46 (s, 1H, NH uracil), 8.38 (s, 1H, CH uracil), 7.90-7.70 (m, 6H, CH naphth. and CH aromatic Fmoc), 7.62 (t, 2H, $J = 7.3$ Hz, CH aromatic Fmoc), 7.50-7.37 (m, 4H, CH naphth. and CH aromatic Fmoc), 7.32 (d, 2H, $J = 7.5$ Hz, CH aromatic Fmoc), 5.90 (s, 1H, NH Fmoc), 4.77 (d, 2H, $J = 5.6$ Hz, CH_2 naphth.), 4.65 (s, 1H, CH_2 linker), 4.50 (s, 2H, CH_2N_3), 4.39 (d, 1H, $J = 7.2$ Hz, CH Fmoc), 4.30-4.18 (m, 2H, CH_2 Fmoc), 3.96 (s, 1H, $\text{CH}_2\text{COO}^t\text{Bu}$), 3.64-3.33 (m, 4H, $\text{NHCH}_2\text{CH}_2\text{NH}$), 1.48 (s, 9H, ^tBu); ^{13}C (100 MHz, 25°C, CDCl_3) $\delta(\text{ppm})$: 157.0, 143.8, 141.3, 127.7, 127.0, 125.0, 120.0, 66.8, 53.46, 47.2, 42.1, 40.6, 40.3, 27.8, 27.7, 23.4. NMR signals are relative to major rotamer. MS (ESI+) m/z : $[\text{M} + \text{Na}]^+$ calcd 809.3, found 809.3.

Methyl 2-(N-(2-(((9H-fluoren-9-yl) methoxy) carbonyl) amino) ethyl)-2-(5-(((7-(aminomethyl) naphthalen-2-yl) methyl) carbamoyl)-2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl) acetamido) acetate (12a). Compound **17** (30 mg, 0.04 mmol) was dissolved in THF (1 ml) in a tube and stirred with PPh_3 (11.6 mg, 0.04 mmol) and water (36 μl , 2.00 mmol) at room temperature overnight. Then TFA (1 ml) was added and solvent removed under vacuum. Residue was dissolved in the minimum amount of TFA and then the tube was filled with Et_2O . Precipitate was centrifuged and washed two time with ether. White solid was dried under vacuum to afford pure product (70%, 20 mg).

(12a) $^1\text{H-NMR}$ (400 MHz, 25 °C, d_6 -DMSO) $\delta(\text{ppm})$: 12.00 (s, 1H, NH uracil), 9.22 (t, 1H, $J = 6.1$ Hz, NH amide uracil), 8.49 (s, 1H, CH uracil), 8.00 – 7.80 (m, 4H, CH naphth. and CH aromatic Fmoc), 7.72 – 7.64 (m, 4H, CH naphth. and CH aromatic Fmoc), 6.61 (1H, t, $J = 5.5$ Hz, NH amide Fmoc), 4.95 (s, 2H, CH_2 linker), 4.66 (d, 2H, $J = 5.5$ Hz, NHCH_2 naphth.), 4.41 (d, 2H, $J = 4.6$ Hz, CH_2NH_2), 4.35 (d, 2H, $J = 6.7$ Hz, CH_2 Fmoc), 4.24 (t, 1H, $J = 6.6$ Hz, CH Fmoc), 4.08 (s, 2H, CH_2COOMe), 3.84 (bs, 2H, NH_2), 3.62 (3H, s, OMe), 3.44 (t, 2H, $J = 5.9$ Hz, $\text{CH}_2\text{CH}_2\text{N}$), 3.32 – 3.23 (m, 2H, $\text{CH}_2\text{CH}_2\text{N}$). $^{13}\text{C-NMR}$ (100 MHz, 25°C, d_6 -DMSO) $\delta(\text{ppm})$: 169.9, 167.5, 164.1, 162.3, 158.7, 156.8, 152.6, 150.5, 144.3, 141.2, 139.3, 137.5, 133.3, 131.6, 128.3, 128.1, 127.5, 126.1, 125.9, 125.6, 125.2, 120.6, 105.0, 66.0, 52.3, 49.2, 49.0, 48.3, 47.2, 43.6, 42.7, 38.8. NMR signals are relative to major rotamer. MS (ESI+) m/z : $[\text{M} + \text{H}]^+$ calcd. 719.3, found 719.3. FT-IR ν (cm^{-1}) 3314, 1686, 1613, 1543, 1466, 1450, 1380, 1340, 1237, 1203, 1180, 1144, 1006. Decomposes at 235 – 236 °C.

2-(N-(2-((((9H-fluoren-9-yl) methoxy) carbonyl) amino) ethyl)-2-(5-(((7-(azidomethyl) naphthalen-2-yl)methyl) carbamoyl)-2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl) acetamido) acetic acid (19).

Compound **18** (185.4 mg, 0.24 mmol) was dissolved in DCM (6 ml) and then TFA (2 ml) was added to the solution at 0 °C. Solution was stirred for 30 min and for further 150 min at room temperature. Reaction was quenched with MeOH (10 ml) and solvent evaporated under vacuum to afford pure product product (82%, 172.2 mg).

(19) ¹H-NMR (400 MHz, 25 °C, d₆-DMSO) δ(ppm): 11.96 (s, 1H, OH), 9.20 (s, 1H, NH), 8.45 (s, 1H, CH uracil), 7.90 (d, 2H, J = 8.00 Hz, CH napht.), 7.87 (s, 2H, CH Fmoc), 7.68 (s, 2H, CH naphth.), 7.64 (s, 2H, CH Fmoc), 7.38 (d, 2H, J = 1.65 Hz, CH naphth.), 7.37 (s, 2H, CH Fmoc), 7.29 (s, 2H, CH Fmoc), 4.90 (s, 2H, NCOOH), 4.71 (s, 2H, CH₂ Fmoc), 4.64 (t, 1H, J = 4.00 Hz, CH Fmoc), 4.56 (d, 2H, J = 5.9 Hz, NHCH₂), 4.16 (s, 2H, NCH₂), 3.38 (t, 2H, J = 4.00 Hz, NCH₂CH₂NH), 3.23 (q, 2H, J = 4.00 Hz, NCH₂CH₂NH), 2.92 (s, 2H, NCH₂CO), 2.28 (s, 2H, CH₂N₃); ¹³C-NMR (100 MHz, 25 °C, d₆-DMSO) δ(ppm): 170.7, 167.2, 164.1, 156.8, 150.5, 144.3, 141.2, 138.0, 133.9, 133.1, 128.7, 128.4, 128.0, 127.5, 126.7, 125.8, 125.6, 120.5, 79.6, 65.9, 54.2, 49.1, 48.9, 48.2, 47.3, 47.1, 29.4. NMR signals are relative to major rotamer. MS (ESI+/-) m/z: [M - H]⁻ calcd. 729.25, found 729.04, [M + H]⁺ calcd. 731.26, found 731.23, [M + Na]⁺ calcd. 753.24, found 753.16, [M + K]⁺ calcd. 769.21, found 769.13. FT-IR ν (cm⁻¹) 2948, 2360, 1682, 1611, 1540, 1465, 1450, 1416, 1381, 1339, 1180, 1136, 1007. m.p. 208 – 210 °C.

PNA oligomers synthesis. PNA oligomers were synthesized on solid phase using Fmoc strategy. The synthesis was performed manually using plastic vessel equipped with polyethylene frit, cap and valve. Rink amide MBHA resin was downloaded before use to 0.2 mmol/g of active sites using Fmoc-Gly-OH. Unreacted active sites were capped with acetic anhydride (DMF/Ac₂O 1:1) and exact loading determined by measuring UV absorbance at 290 nm of small amounts of resin dispersed in a 20% piperidine in DMF solution. Loading, expressed in mmol/g, was given from following formula:

$$\text{Loading} = \frac{1.68}{\text{Abs}(290\text{nm}) \times \text{mg}(\text{resin})}$$

Chain elongation occurred with the following protocol: 1) Fmoc deprotection with 20% piperidine solution in DMF for 8 min 2 times; 2) activation of 5 eq of PNA monomer (respect resin active sites) with 4.9 eq HBTU, 10eq DIEA for 2 min; 3) Coupling 30 min for standard monomers and overnight for monomer **20**; 4) Capping with DMF/DIEA/Ac₂O 89/6/5 1 min for 2 times; 5% DIEA in DMF washings 2 min for 2 times. PNA was cleaved with 10% m-cresol in TFA, 1.5 h (twice). The PNA was precipitated from the TFA solution with ethyl ether (10 times TFA volume) in freezer for 2 h and washed three times. Terminal Fmoc group was removed in 20% piperidine solution after first cycle of purification.

Oligomers purification. Crude PNA were purified using HPLC (Agilent Technologies 1260 Infinity, G1311C 1260 Quat Pump VL, G1316A 1260 TCC thermostat, G1314B 1260 VWD VL UV detector). Purification protocol and column changed depending on the PNA. Water +0.1% TFA (A) and acetonitrile +0.1% TFA (B) were used as eluent. Flow was set to 4 ml/min.

Unmodified PNA (NM1, NM2) were purified on reversed phase semi preparative C18 column (Xterra® Prep RP18, 10 μm, 7.8 x 300 mm, 300Å), at 35 °C using the gradient reported in Appendix 8.4.

PNA bearing modifications were separated on reversed phase semi preparative C18 column (Phenomenex® Jupiter C18 300Å, 5 μm, 10 x 250 mm), at 50 °C using gradients described in Appendix 8.4.

Characterization of pure products occurred through UPLC -ESI-MS (Acquity Ultra Performance LC) on reversed phase column (Waters UPLC BEH 300 C18, 1.7 μm, 50 x 2.1 mm), flow 0.25 ml/min, water +0.2% HCOOH (A) and acetonitrile +0.2% HCOOH (B) as eluents (gradient is reported in Appendix 8.5).

NM1. (1650.63) MS (ESI+) m/z: [M+2H]²⁺ Calcd. 826.32 Found 825.92; [M+3H]³⁺ Calcd. 551.21 Found 551.05; [M+4H]⁴⁺ Calcd. 413.66 Found 413.68. ε = 52700. Yield = 12%.

NM2. (1990.93) MS (ESI+) m/z: [M+3H]³⁺ Calcd. 664.64 Found 664.50; [M+4H]⁴⁺ Calcd. 498.73 Found 498.70; [M+5H]⁵⁺ Calcd. 399.19 Found 399.20. ε = 74600. Yield = 20%.

PNA1. (2000.92) MS (ESI+) m/z: [M+2H]²⁺ Calcd. 994.46 Found 995.02; [M+3H]³⁺ Calcd. 663.31 Found 663.69; [M+4H]⁴⁺ Calcd. 497.73 Found 497.80. ε = 63774. Yield = 0.5%

PNA2. (1986.92) MS (ESI+) m/z: $[M+2H]^{2+}$ Calcd. 1001.46 Found 1001.42; $[M+3H]^{3+}$ Calcd. 667.97 Found 667.76; $[M+4H]^{4+}$ Calcd. 501.23 Found 501.16. $\epsilon = 63774$. Yield = 1.1 %

PNA3. (2341.22) MS (ESI+) m/z: $[M+2H]^{2+}$ Calcd. 1171.61 Found 1172.18; $[M+3H]^{3+}$ Calcd. 781.41 Found 781.57; $[M+4H]^{4+}$ Calcd. 586.31 Found 586.57. $\epsilon = 85674$. Yield = 0.2%

Melting experiments. Concentration of stock PNA solutions were determined by UV measurement at 260 nm using extinction coefficients reported above for each PNA. Sample were prepared in order to have a final PNA concentration of 5 μ M, in PBS buffer, for a total volume of 1 ml. Thermal denaturation profiles (Abs vs T) for single strand were recorded using UV/Vis Thermo Scientific Evolution 260 BIO spectrophotometer equipped with a Peltier temperature controlling system (Thermo Fisher Scientific SPE8W). Heating rate was set to 1°C/min. Melting curves were smoothed using Savitsky Golay algorithm. T_m values were obtained from minima of first derivative of curves.

5. Virtual screening of dimeric base monomers and building blocks synthesis

5.1. Introduction

As discussed in chapter 1, classic approach for finding new modified PNA structures with improved RNA binding properties is based on the information inferred from solid-state structures available in literature and then synthesis of new compounds in a trial-and-error strategy. Synthesis of new modified compounds is time and money consuming, generally requiring many reaction steps and conditions, and each of these must be optimized in order to obtain acceptable yields. However, the crystal structures, though very useful for getting insights into the molecular properties, represent a static model, and do not account for all possible conformations of the molecules involved; furthermore, the solid-state structure can be different from the preferred conformation in solution. Therefore, a computational approach able to predict effects of introduction of modifications on systems would be extremely useful in order to select more promising compounds and focus synthesis on these. In chapter 3 we have presented a robust methodology to study properties of PNA both as single strand and duplex. We wanted then to apply Molecular Dynamics for a virtual screening of modified PNA monomers, in order to take into account the possible evolution of the system upon thermal motion, and all possible conformations of modified monomers and their effect on duplex stability. Moreover such approach, which was made simpler by the availability of the modified NAB developed by us, allowed to directly study the effects on a specific sequence of applicative interest of the PNA:RNA duplex. In particular for this study we have chosen a PNA complementary to the seed region of miR 221 and 222 (H-ATG**I**AGC-Gly-NH₂, **I** is the modified dimeric base). Simulations described in this chapter were carried out in collaboration with the group of Prof. Michele Parrinello, at the Department of Chemistry and Applied Biosciences ETH Zurich, USI Campus, Lugano, using the computing facilities there available.

5.2. Molecular Dynamics for base-modified PNA Model 1 (Mod-1)

In Chapter 4 were presented some interesting properties concerning a modified PNA monomer containing two uracil moieties. We wanted to simulate a PNA containing this dimeric base (Figure

4.2), which will here be called Mod-1, in order to better understand experimental data. Using the modified NAB (section 2.7) and manual refinements, where needed, we constructed duplexes of the PNA with both DNA and RNA (Figure 5.1). These systems were energy-minimized and 200 ns long simulations were performed on both systems (DNA: cubic box 55 X 51 X 64 Å³, 17700 atoms; RNA cubic box 55 X 51 X 62 Å³, 17300), using the same conditions reported in section 2.5.

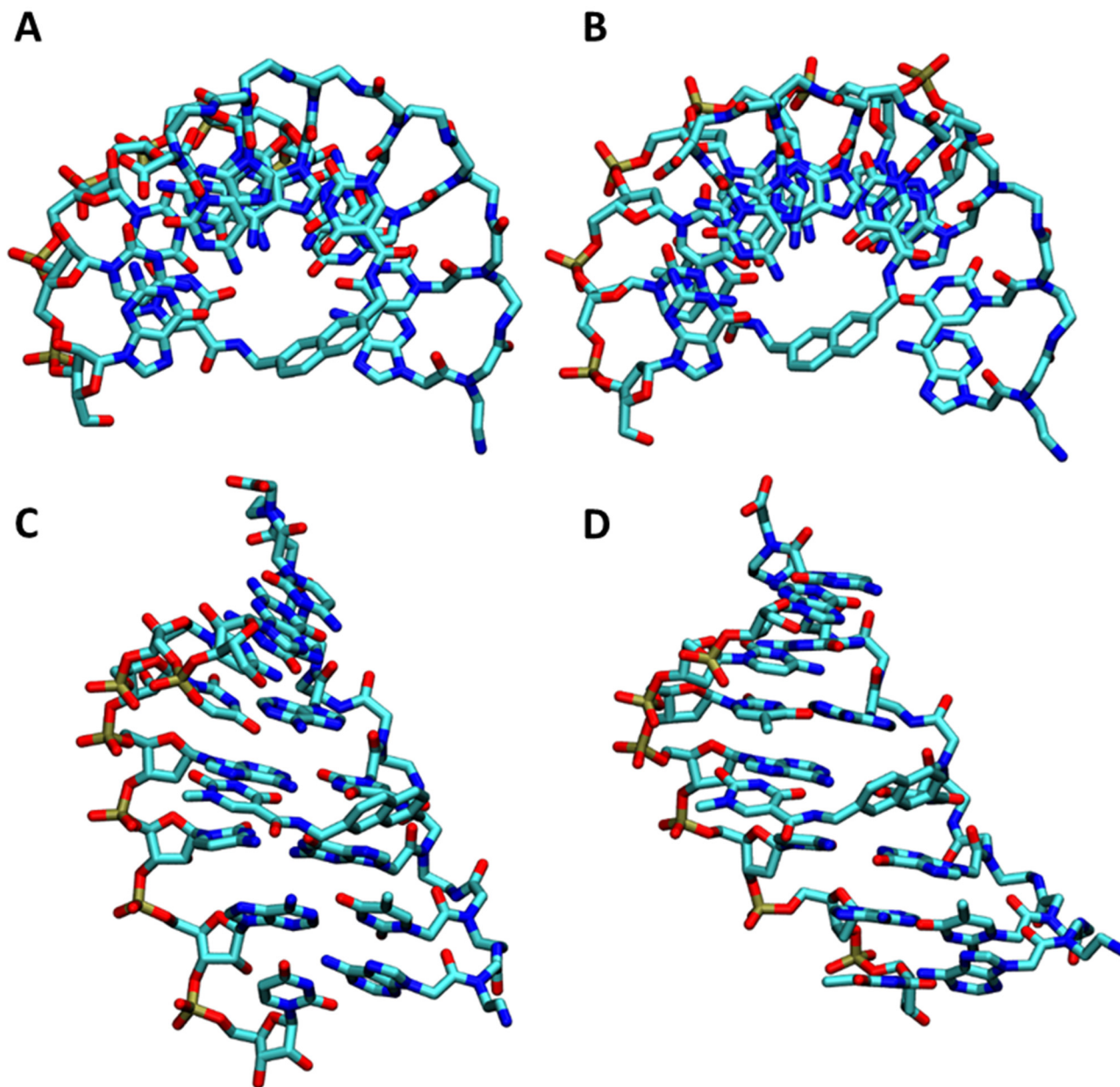


Figure 5.1: Structures of the systems containing Mod-1 monomer. A) top view and C) side view of PNA:RNA duplex. B) top view and D) side view of PNA:DNA duplex.

The first uracil (U1) was perfectly inserted in duplex and second uracil (U2) was able to interact with target adenine. However, U2 was not perfectly paired via Hoogsteen hydrogen bonds, due to sterically tensioned conformation, thus inducing only a limited interaction. It had also the possibility

to pair with another adenine present in the sequence and it was actually less tensioned in this conformation. U2 was also free to move in solvent and then to interact with other bases through stacking interactions. These two limit conformations are shown in Figure 5.2 in the case of PNA:RNA duplex.

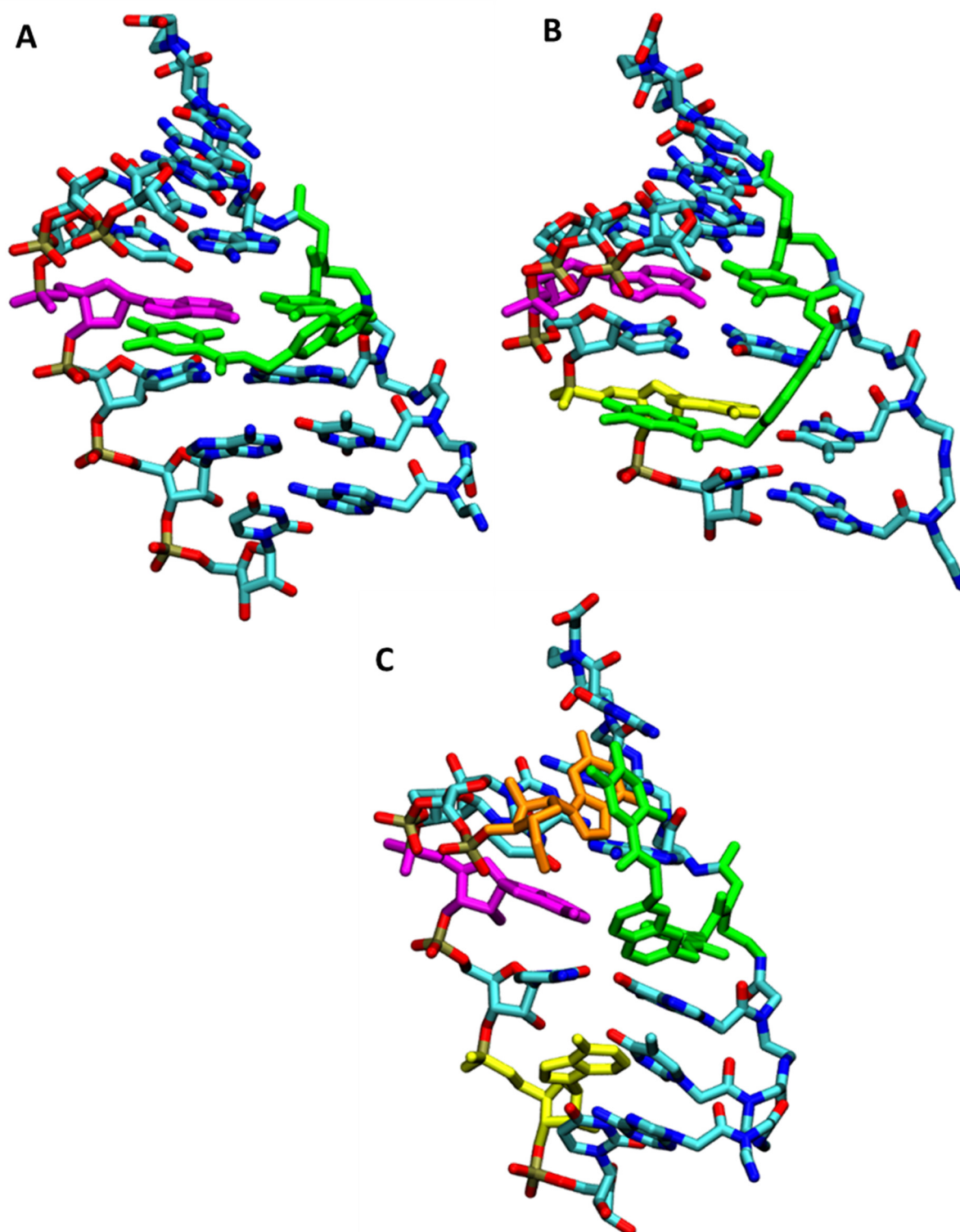


Figure 5.2: Snapshots of the MD simulation for Mod-1 pairing with RNA: **A)** interaction of modified base (green) with target adenine (magenta); **B)** interaction with another adenine of the sequence (yellow); **C)** stacking interaction with other base (orange).

These results are in accordance with experimental data. As seen in previous chapter, study on PNA:DNA heteroduplex revealed that insertion of modified base only slightly increased stability of the duplex and previous study on **PR212**²⁷⁰ pointed out more interesting properties of selectivity. When target adenine is present in the sequence, partial interaction of second uracil provides some stabilizing effect, while when a mismatch occurs, pedant uracil is completely free to move in solvent, interfering with other bases pairing and thus inducing destabilization. However, for adenine, though some attractive interactions are observed, the effect of the second uracil is less than that expected, because of the relative mobility of this moiety. Furthermore, if the PNA:DNA and PNA:RNA duplex stability is not strongly enhanced, competition of PNA:PNA self-association can be relevant. In fact, experimental melting of PNA:DNA duplexes suggested also a preferred self-aggregation of the system instead of duplex formation in presence of TA or AT doublet in the PNA sequence (section 4.4). Though we have not considered the stability of PNA:PNA duplex at the computational level, the self-pairing of the AT pair would be favored by the flexibility of the two PNA involved, which allows the nucleobases to maximize their mutual interactions.

5.3. Molecular Dynamics for base-modified PNA Model 2 (Mod-2)

Since model described above presented some geometrical constraints which hampered the pairing of the second uracil moiety with the target adenine, we focused our attention on the presence of the rigid amide bonds. We then substituted the amide group of second uracil with an amine, which should presents a greater mobility and consequently adaptability (Figure 5.3).

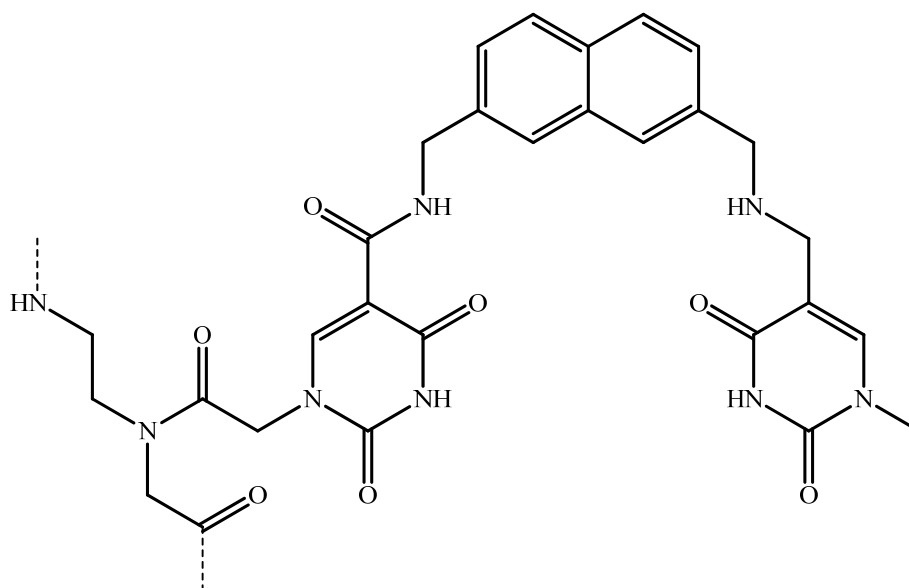


Figure 5.3: Structure of Mod-2 modified base.

Since simulations were time demanding and considering that desired final targets were micro RNAs, here after we performed simulations only on PNA:RNA duplexes. The starting duplex structure was generated using modified NAB, and the new dimeric base was introduced manually by modifying a thymine base (Figure 5.4). We performed 200 ns of standard MD (cubic box of 60 X 55 X 67 Å³, 17300 atoms) in the conditions reported in section 2.5.

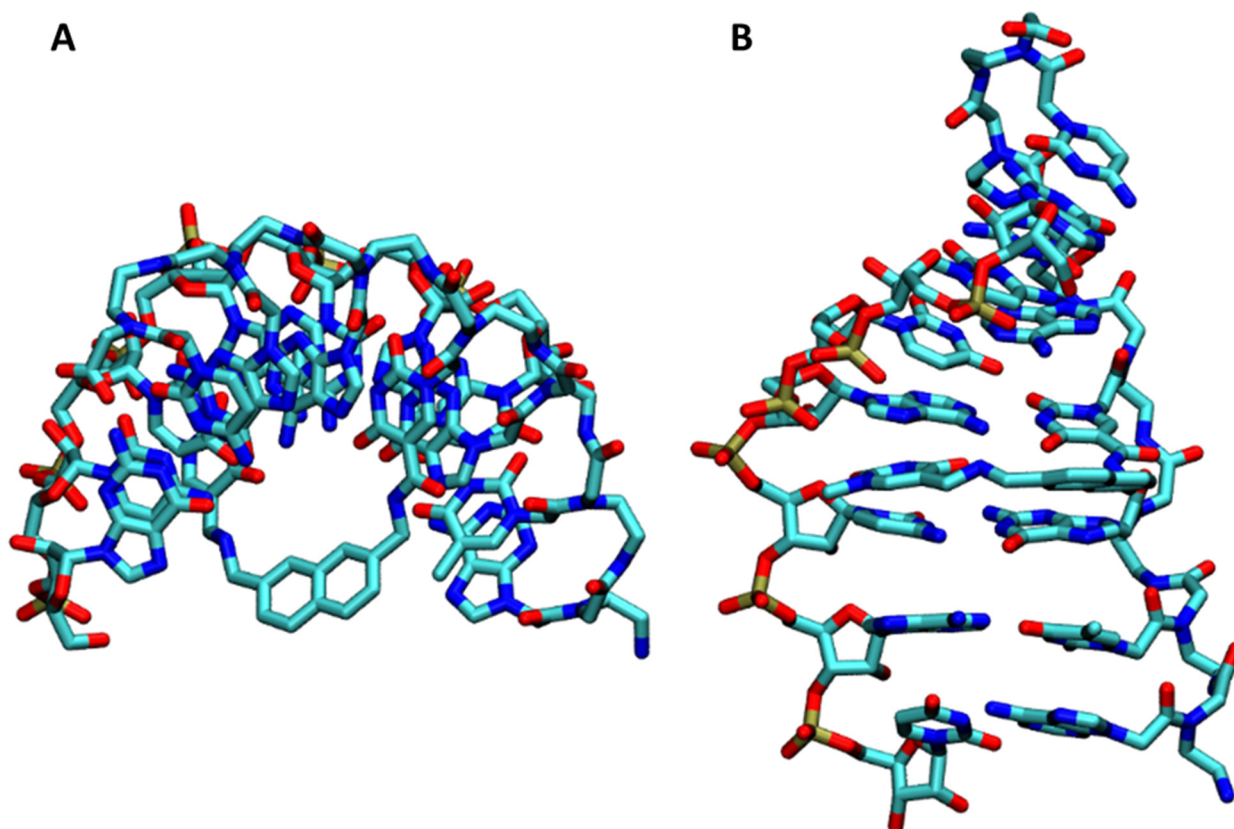


Figure 5.4: Starting structure, containing Mod-2 monomer, used for the MD simulation. **A)** top view **B)** side view.

The simulation showed that the presence of the amino group did not allow the second uracil to reach a stable interaction with the target adenine; thus also this base is not correct for a cooperative interaction with target. Moreover, the increased flexibility of the bridge linking the second uracil unit, determines a favored collocation of U2 away from the target, stabilized by interactions with solvent (Figure 5.5).

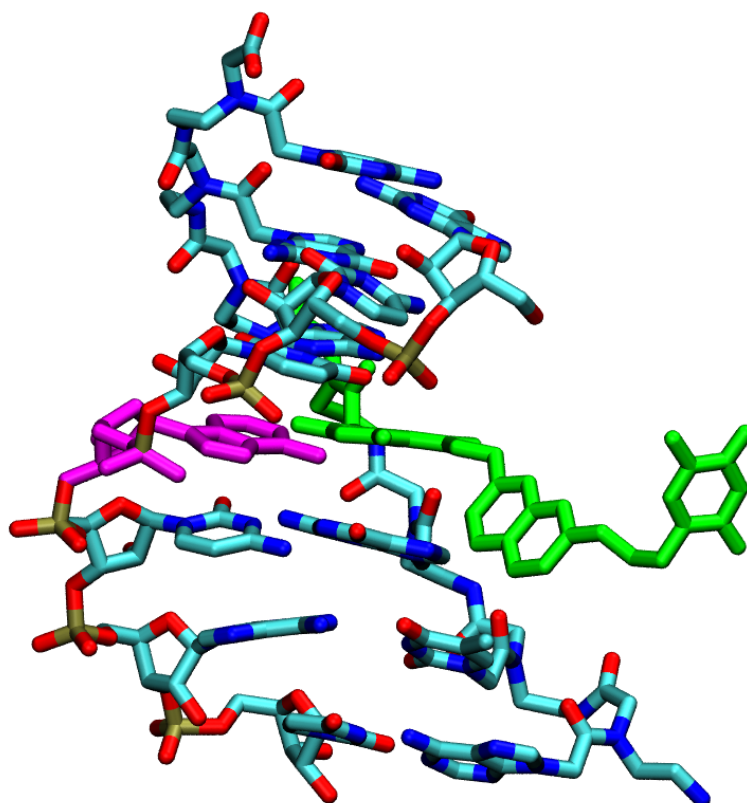


Figure 5.5: Favored disposition of Mod-2 (green). Pedant uracil freely move in the solvent while the first uracil is involved in standard Watson and Crick bonds with target adenine (magenta).

5.4. Molecular Dynamics for base-modified PNA Model 3 (Mod-3)

Opposite to the previous model, we tested if a rigid linker could allow cooperative interactions with the target adenine still avoiding the collapse of the two uracils. We thus kept the naphthalene linker, and looked for synthetically accessible structures. Therefore, we proposed Model 3 as possible new candidate for binding adenine (Figure 5.6).

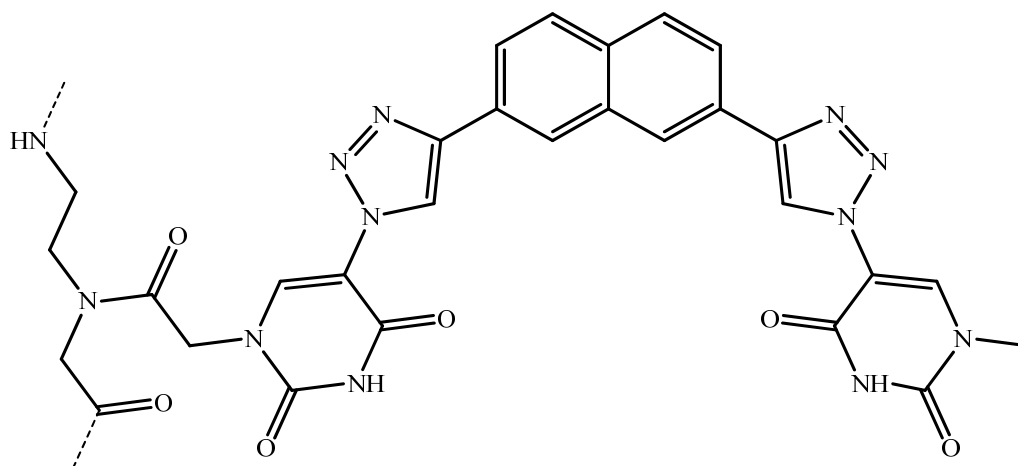


Figure 5.6: Structure of Mod-3 central fragment.

The triazole ring in this monomer can be obtained by a copper (I) catalyzed click reaction between 2,7-diethynyl-naphthalene and a 5-azido uracil derivative. This compound presents a major distance between two uracils compared to Mod-1 previously described (10 Å compared to 9.5 Å) and then should reduce steric constraints. The starting PNA:RNA duplex structure was generated using same procedure used for the previous model. We performed a 40 ns simulation in a cubic box filled with TIP3P water molecules (58 X 54 X 64 Å³, 17750 atoms), under the same conditions used before (Figure 5.7).

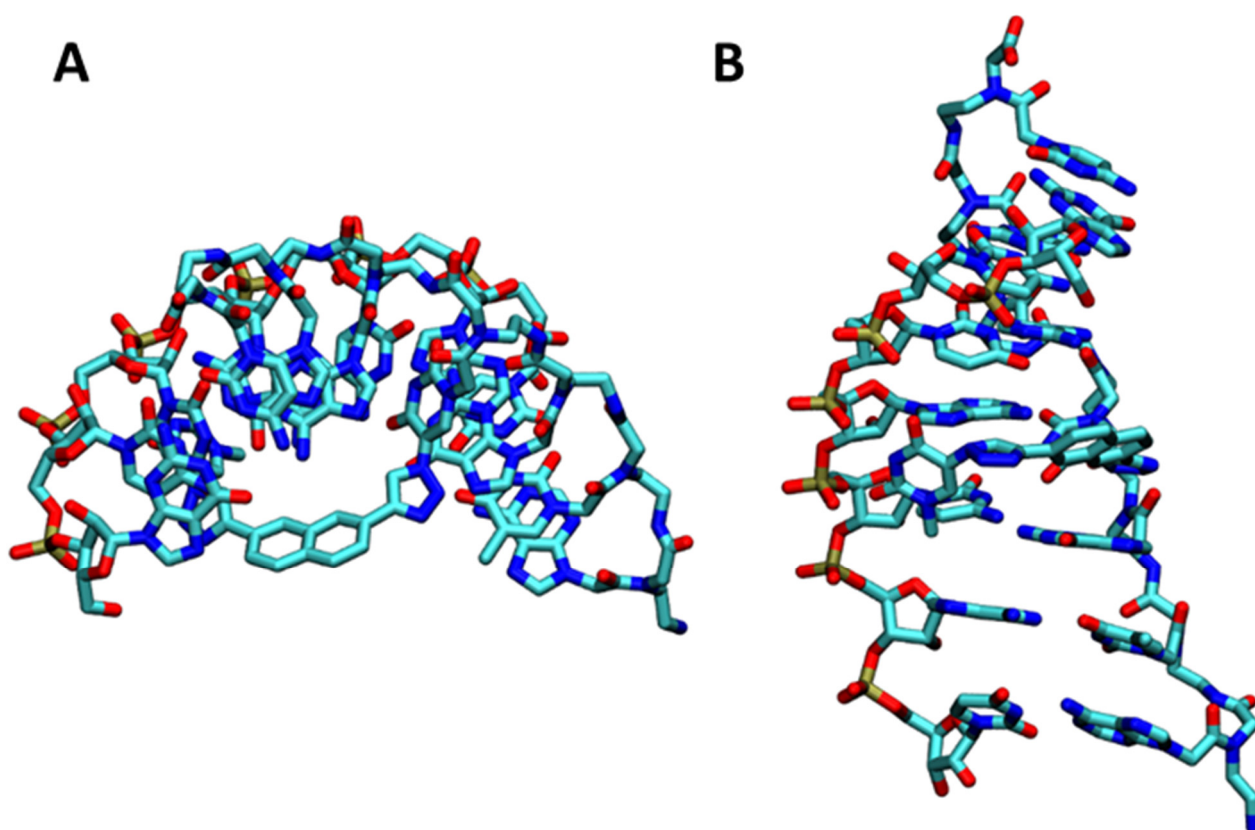


Figure 5.7: Starting structure, containing Mod-3 monomer, used for the MD simulation. **A)** Top view of the system; **B)** Side view of the system.

The simulation showed that also for Mod-3 the second uracil didn't present the correct geometry to interact with adenine. However, in this case, unlike with Mod-2, the second uracil, being part of a rigid moiety, gave rise to destabilizing steric interactions which interfered also with base pairing of the first (Watson-Crick) uracil, causing weakening in the central part of the duplex and consequent duplex disruption. Indeed a little breach was generated during MD simulation, and this allowed water entering destabilizing the systems until complete opening (Figure 5.8).

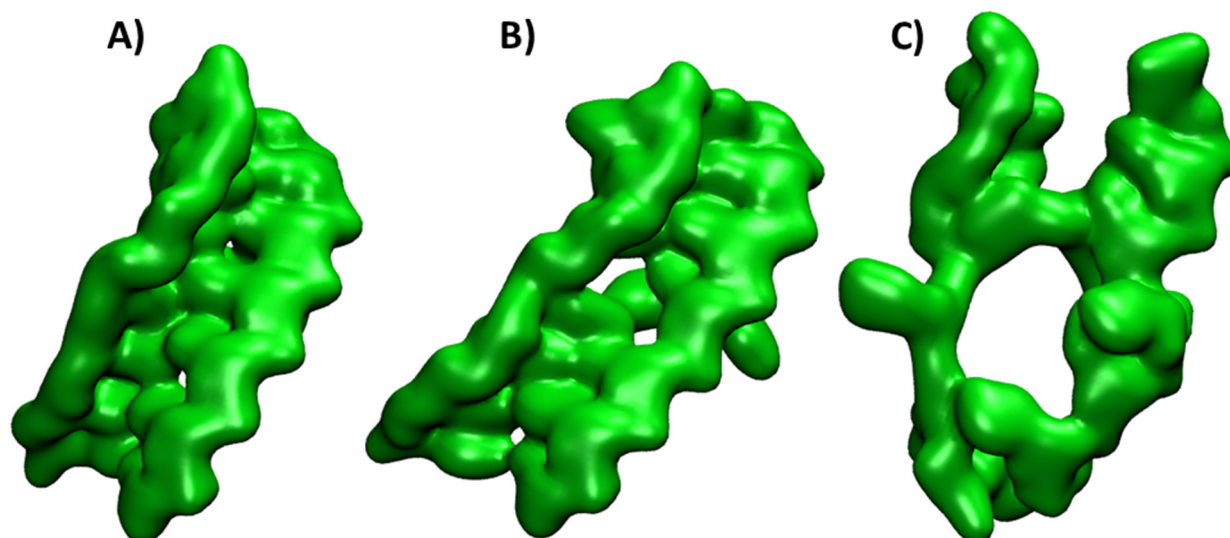


Figure 5.8: Snapshots of MD simulation for Mod-3 PNA:RNA. Duplex surface representation. **A)** view of duplex minor groove at the beginning of the simulation; **B)** in the central part of the duplex was possible to recognize a breach that allowed water entering destabilizing the system; **C)** duplex was completely opened in the central part.

Behavior of Mod-3 is a good example to show difficulty in finding new effective modified systems. Compounds with similar geometries exhibited completely different properties and what was expected to be an advantage, greater uracil distance, was indeed found to be a drawback.

5.5. *Ab initio* calculations and Molecular Dynamics for base-modified PNA

Model 4 (Mod-4)

Finally, we studied another modified base, analogue to those shown before (Figure 5.9), but with removal of any sp^3 atoms from the bridge, thus allowing a completely flat geometry to be obtained. Contrary to Mod-3, the additional sp^2 atoms were not forced to be part of a cycle, thus allowing a different geometry.

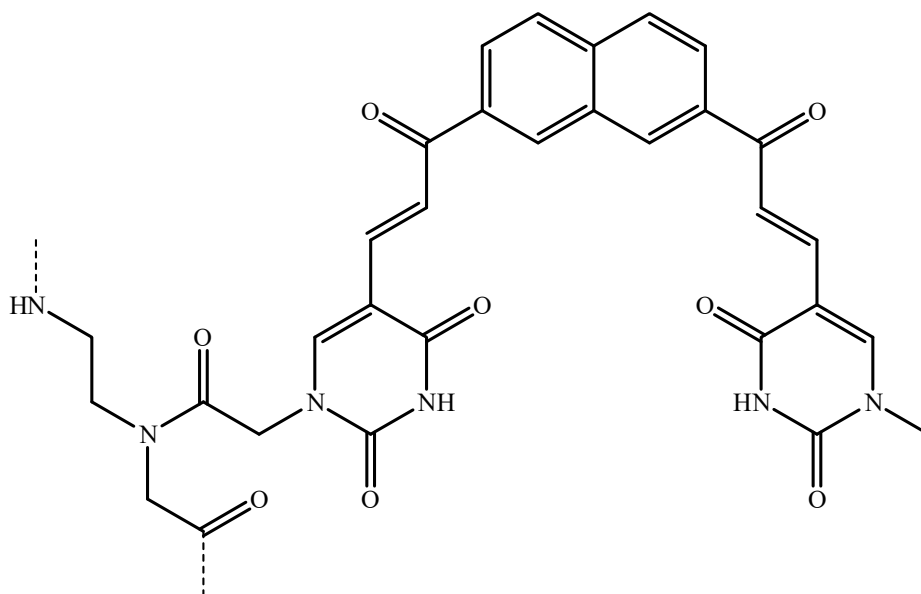


Figure 5.9: Structure of Mod-4 central fragment.

This compound presented an α,β -unsaturated ketone as linker between naphthalene and uracil moieties. This modification was designed taking account of the excessive mobility of pedant uracil of previous models. Conjugation between bonds should favor rigidity and planarity of the molecule, maintaining the Hoogsteen interactions once they are formed. The synthesis of this monomers is more challenging compared to those reported before. Double bond requires *E* geometry and also carbonyl should have the correct orientation in respect to the naphthalene. We then evaluated which could be the preferred orientation by calculating the difference in energy between model compounds reported in Figure 5.10 by *ab initio* energy calculation.

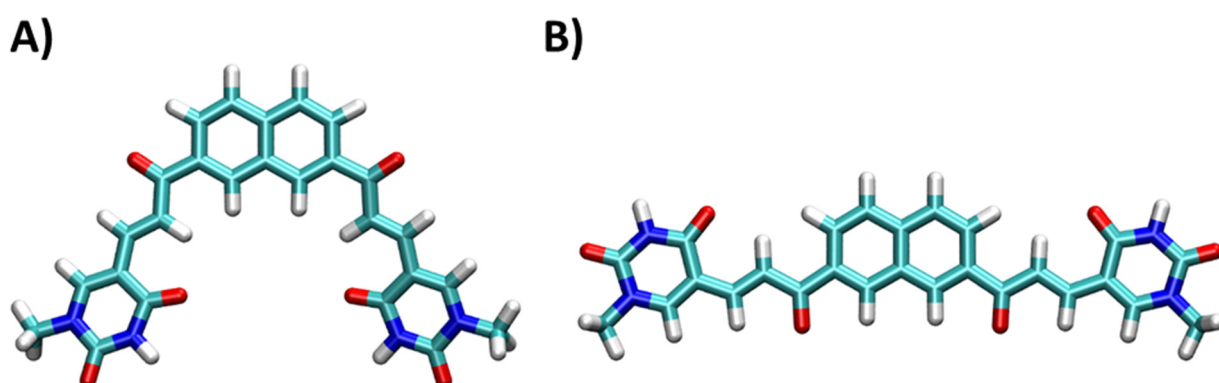


Figure 5.10: Optimized geometry for: **A)** monomer with convergent disposition of uracils which allows adenine clamping; **B)** monomer with conformation with rotated uracils (180° flip of the naphthalene-C=O bonds).

Since this molecule has at least two flat conformations, we performed an optimization of geometry of the cleft-like (Figure 5.10 A) and extended (Figure 5.10 B) structures with Gaussian 09 D1²⁴⁶ using B3LYP²⁸¹ DFT theory and 6-31G* as basis set and then we calculated frequencies in the same way. The most stable conformation was the cleft-like one (Figure 5.10 A), but energy differences of these two compounds in vacuum were small, about 0.5 kcal/mol, and so both are expected to be populated at r.t.. However, once the duplex is formed, it is reasonable to expect that equilibrium can be shifted toward the cleft-like conformation by favorable Hoogsteen interactions. Moreover, the interconversion between these two structures is possible only if the rotational energy barrier is sufficiently low. We then scanned the energy variation as function of the dihedral angle (Figure 5.11) and we found an energy barrier of 6.35 kcal/mol, which should allow the interconversion between these two forms to occur. Optimized structures and PES scan are reported in Appendix 8.7.

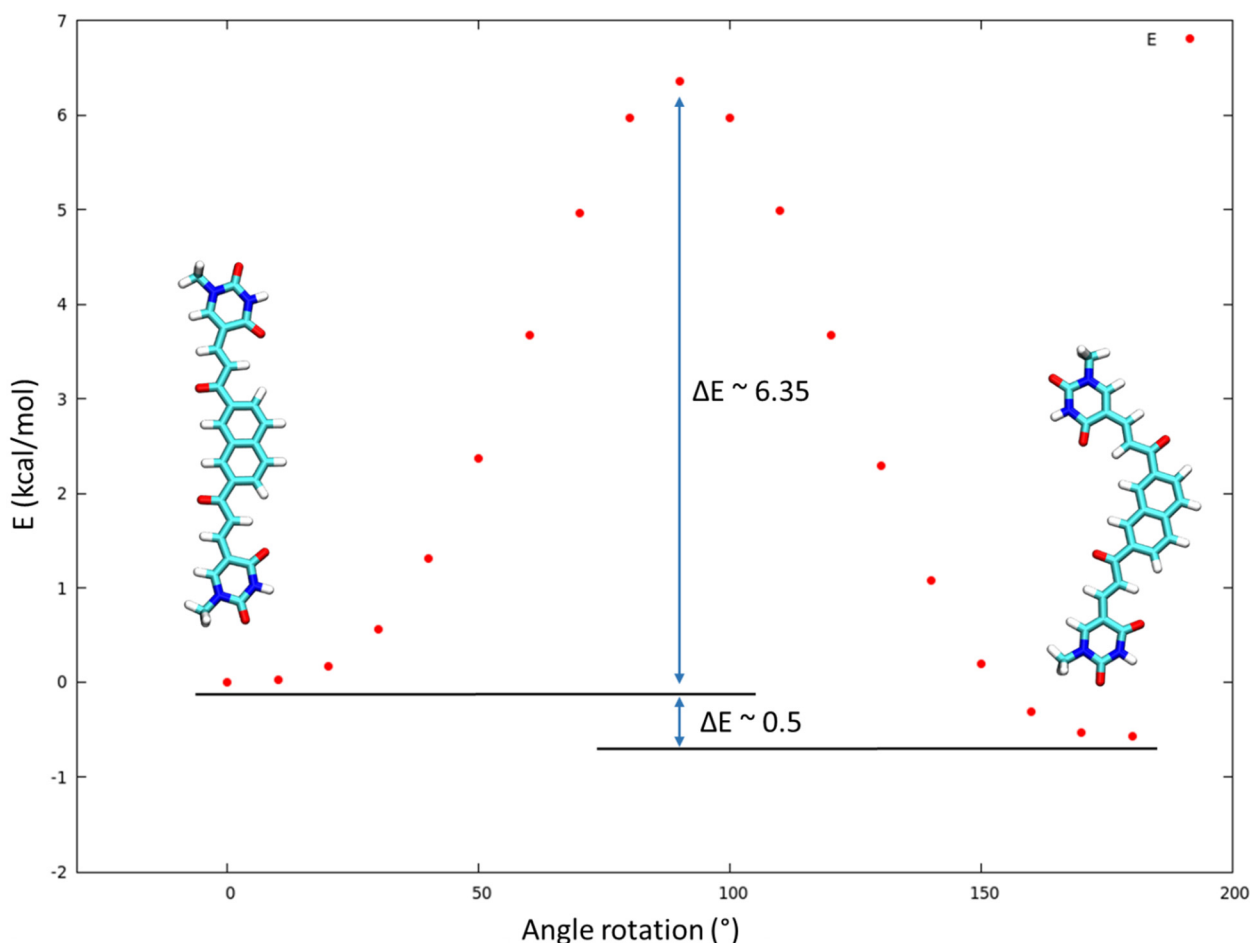


Figure 5.11: Energy plot as function of rotation on naphthalene – CO dihedral angle.

Using this nucleobase, in its cleft-like conformation, we built a PNA:RNA duplex (Figure 5.12) and then performed 200 ns Molecular Dynamics simulation in the condition reported in section 2.5 in a cubic box of 56 X 51 X 62 Å³ filled with TIP3P water molecules for a total of 17350 atoms.

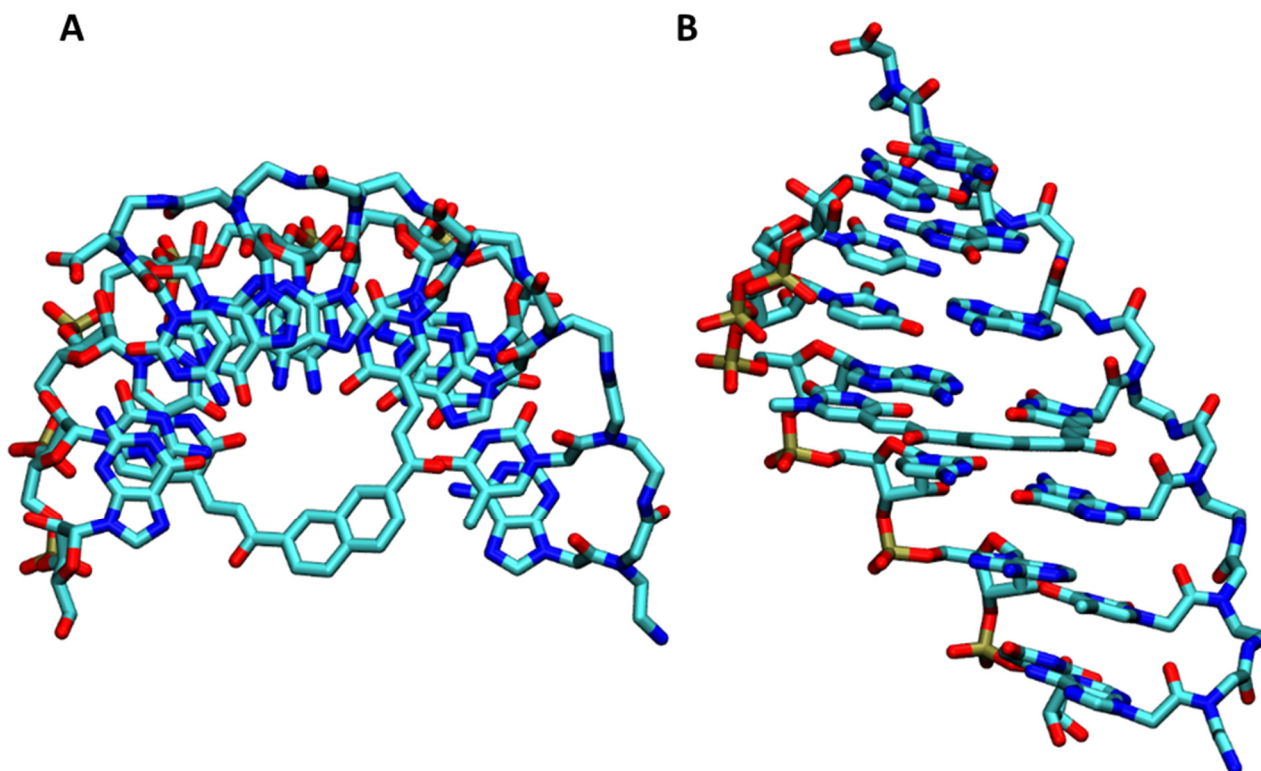


Figure 5.12: PNA containing Mod-4 modified base. **A)** Top view of the system; **B)** Side view of the system.

In this case the duplex remained intact, and the second uracil unit persistently interacted with the target adenine. Checking the average distance between atoms doing H-Bonding of second uracil and target adenine was found that interactions were more consistent than in the previous cases. Base still preserved a certain mobility, as proven by region between 20-50 ns and 60-90 ns, but it retained pairing for about 70% of the simulation (Figure 5.13). Therefore, this compound seemed a more promising candidate compared to other models and we decided to proceed with its synthesis.

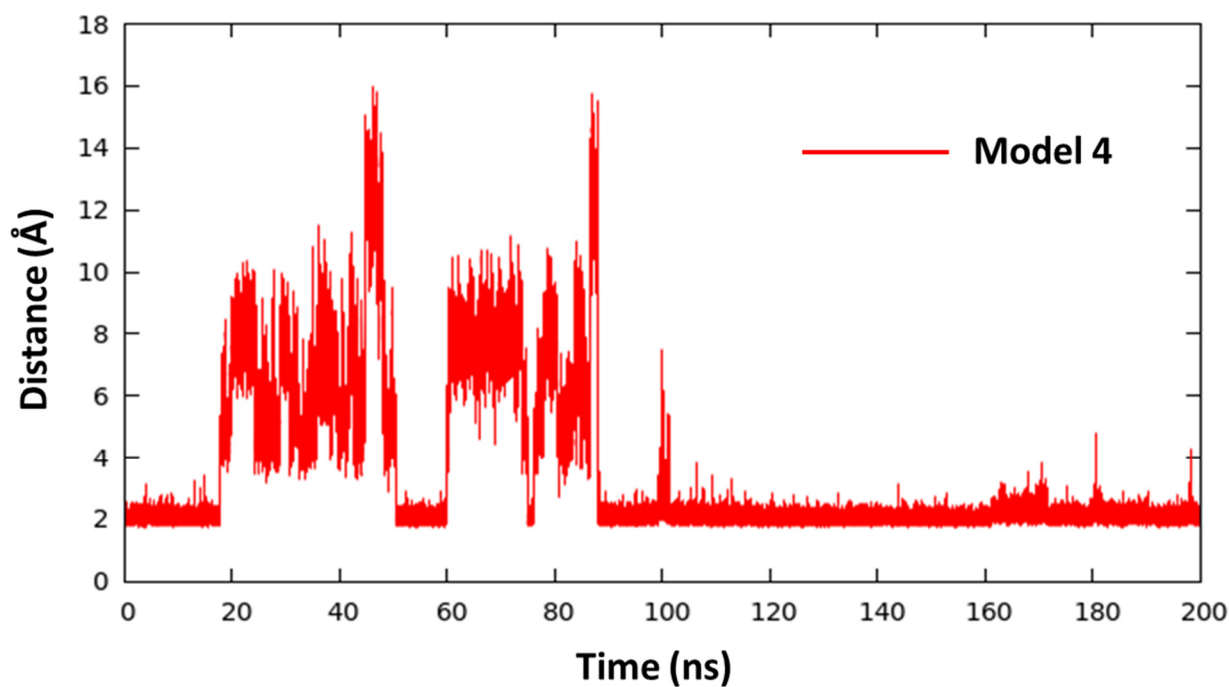


Figure 5.13: Average distance between atoms involved in Hoogsteen bonds (H3 and O4 of modified base and respectively H6 and N7 of target adenine) as function of simulation time.

5.6. Building blocks synthesis

5.6.1. Retrosynthesis

The different synthons composing the Mod-4 nucleobase are depicted in Figure 5.14 with different colors.

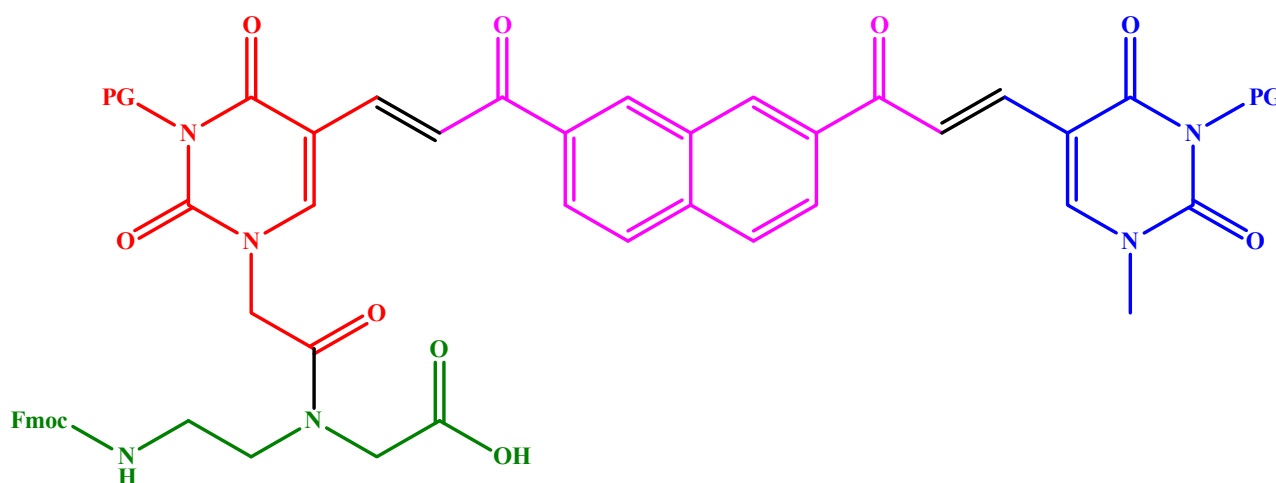
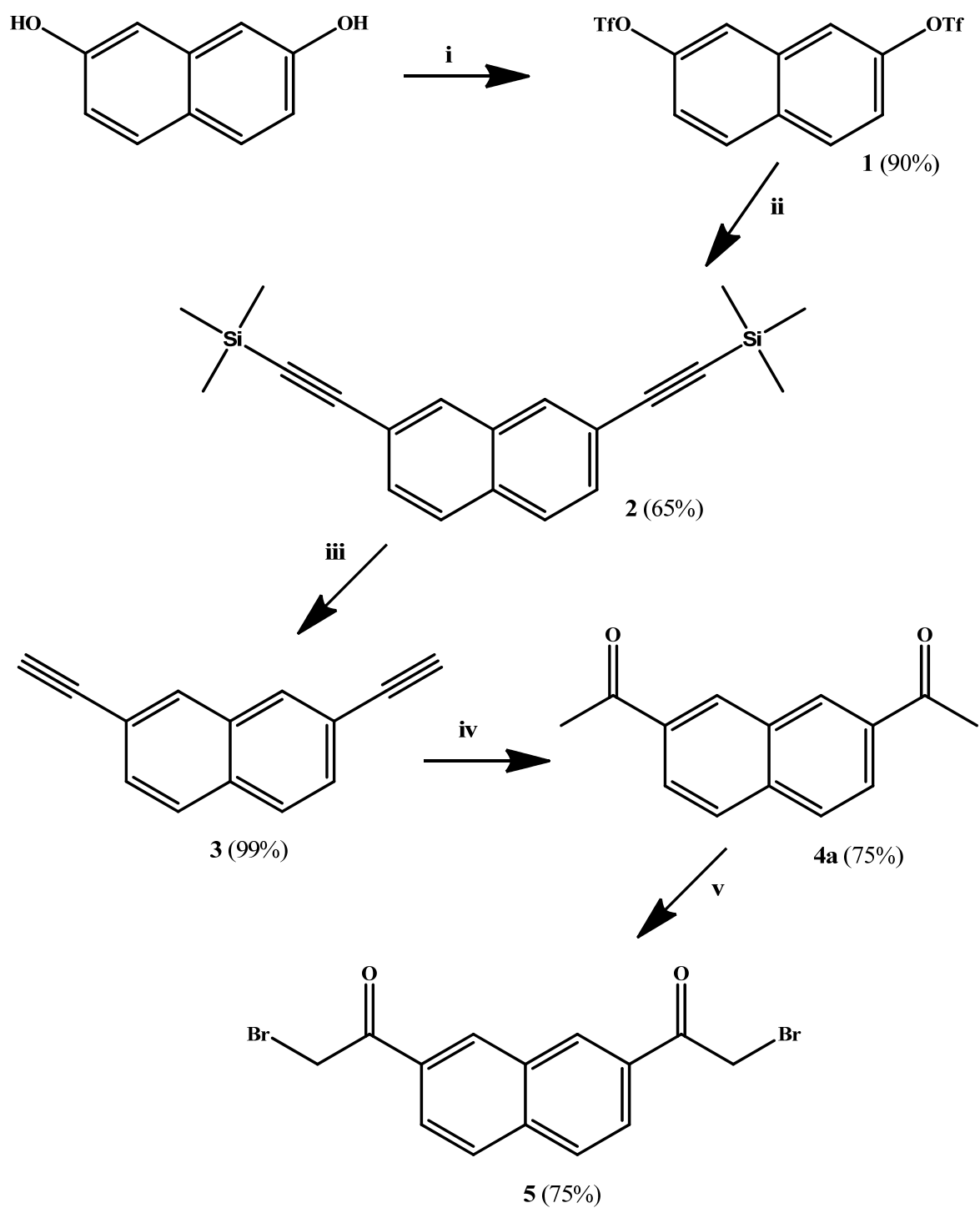


Figure 5.14: Structure of the final monomer derived from Mod-4. Constituent units are marked with different colors. PG is a protective group.

The PNA backbone (green) is the same of the previous monomer and can be synthesized as reported for compound **3** in Chapter 4. The α,β -unsaturated carbonyl unit can be obtained using different strategies such as aldol condensation, carbonylation of the alkene or Wittig reaction.

5.6.1. Synthesis of central linker precursors

The central linker was synthesized starting from 2,7-dihydroxynaphthalene as cheap starting material (Scheme 5.1); this was triflated using $(\text{CF}_3\text{SO}_2)_2\text{NPh}$. Product **2** was obtained from **1** and ethynyltrimethylsilane using a Sonogashira coupling in presence of $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$ and CuI . Silyl groups were then removed under basic conditions to yield **3**. Hydration of alkyne was achieved in glacial acetic acid using iron sulfate as catalyst.²⁸² By controlling the reaction time was possible to obtain the mono-hydrated product **4b** (Figure 5.15) or force the reaction toward compound **4a**. After 2 h the main product was **4b**, but still with a large amount of unreacted **3**. After 5 h, reagent conversion was almost complete and mono (**4b**) and di-hydrated (**4a**) products were in about 1:1 ratio. Leaving the reaction overnight, was possible to obtain an almost complete conversion to product **4a**. We tried both base- and acid-catalyzed aldol condensation with 5-formyluracil derivative **7** (Scheme 5.2) and the diketone **4a**, but the condensation reaction didn't occur under the conditions tested (basic: LDA -78 °C; acid: H_2SO_4 40 °C). This was probably due to a reduced reactivity of 5-formyluracil derivative compared to other aldehydes. Thus, a Wittig-type reaction seems to be more suitable for obtaining the final product (Scheme 5.5). In order to obtain the synthon for this reaction, the diketone was then selectively halogenated in alpha position with NBS to obtain product **5**. Since compound **5** is symmetric, introduction of two different uracil moieties could result difficult. One possible pathway is to form ylide on both side and then slowly add strictly one equivalent of aldehyde followed by one equivalent of the second aldehyde. Another possibility is to convert one bromine atom into iodine using Finkelstein reaction²⁸³. Iodine should form more readily the ylide allowing to perform sequential Wittig reaction. Another possibility is to use compound **4b** allowing introduction of a first uracil moiety and then of a second unit after hydration of alkyne. All these strategies will be tested in future in order to find best conditions for synthesis.



Scheme 5.1: Synthesis of 2,7-bis(bromoaceto)naphthalene. (i) $[(\text{CF}_3\text{SO}_2)_2\text{Nph}, \text{NEt}_3, \text{DCM}, 3\text{h}, 0^\circ\text{C to r.t.}]$; (ii) $[\text{Me}_3\text{SiCCH}, \text{Pd}(\text{PPh}_3)_2\text{Cl}_2, \text{CuI}, \text{NEt}_3, \text{dry DMF}, \text{N}_2, \text{o.n.}, 40^\circ\text{C}]$; (iii) $[1\text{M NaOH/THF } 1:1, 2\text{ h, r.t.}]$; (iv) $[\text{Fe}_2(\text{SO}_4)_3, \text{AcOH}, \text{o.n.}, 95^\circ\text{C}]$; (v) $[\text{NBS}, \text{DCM}, 2\text{ h, reflux}]$.

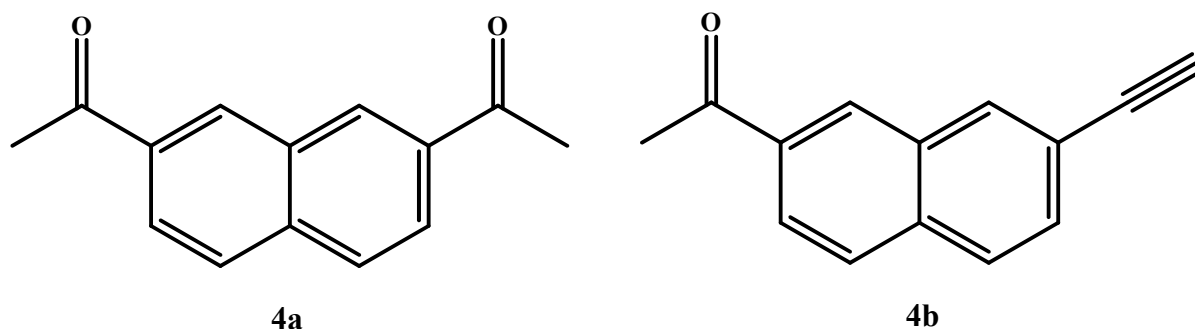
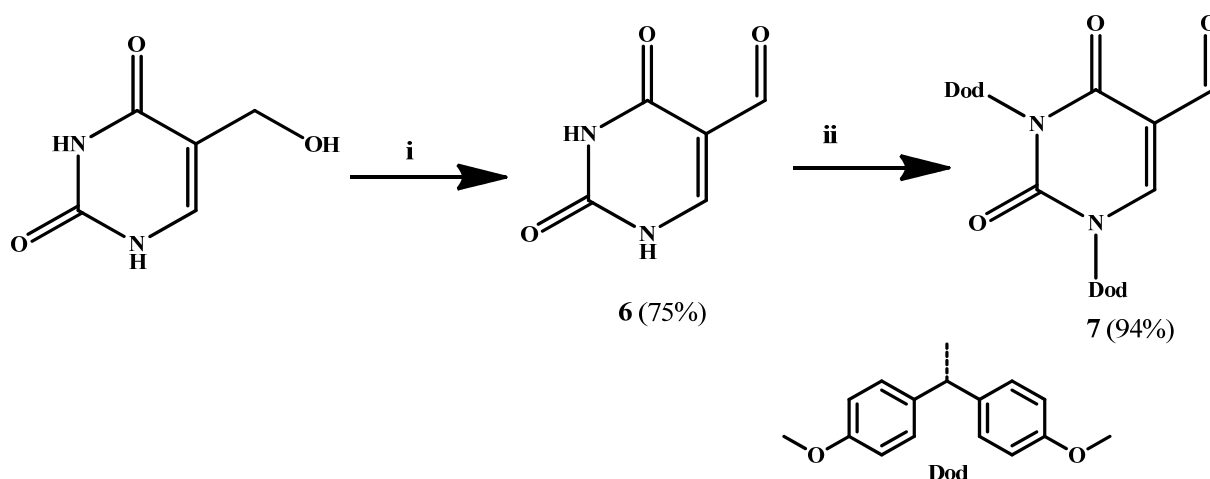


Figure 5.15: Products obtained from hydration of compound **3**.

5.6.2. Uracil derivatives

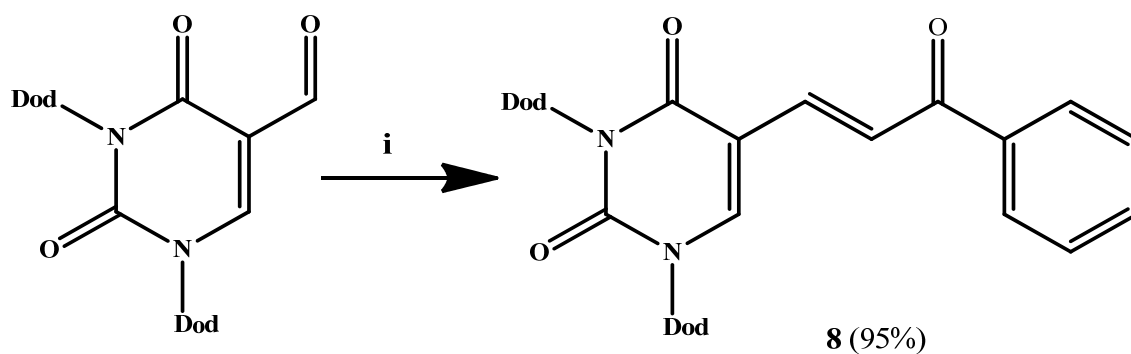
The Wittig reaction involves the use of two differently substituted uracils. In particular we designed a single precursor for both. In order to test the Wittig reaction conditions, we initially synthesized a protected derivative of the aldehyde (Scheme 5.2).



Scheme 5.2: Schematic synthesis of 1,3- di Dod- 5- formyl uracil. (i) [CAN, H₂O, 1h, 90 °C]; (ii) [Dod – Cl, K₂CO₃, dry DMF, 4 h, r.t.].

5 – hydroxymethyl uracil was synthesized according procedure reported in literature.²⁸⁴ Oxidation to the aldehyde **6** occurred in presence of mild oxidizing condition using cerium ammonium nitrate (CAN) [(NH₄)₂Ce(NO₃)₆].²⁸⁵ Aldehyde was then protected with Dod chloride in both N(1) and N(3) position.

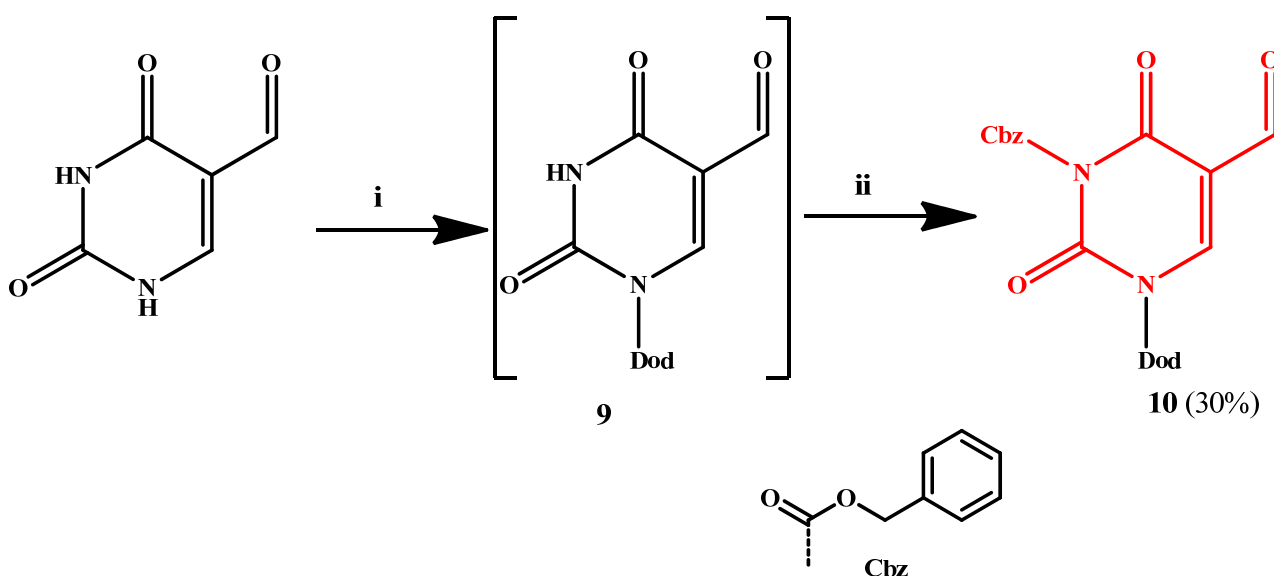
Compound **7** was then used in a model reaction with bromoacetophenone. In particular, since required geometry of the carbon-carbon double bond is *E*, we worked under Schlosser conditions, in which *trans* isomer is favored by heating and by an excess of base (Scheme 5.3).²⁸⁶



Scheme 5.3: Synthesis of (E)-1,3-bis(bis(4-methoxyphenyl)methyl)-5-(3-oxo-3-phenylprop-1-en-1-yl)pyrimidine-2,4(1H,3H)-dione. (i) [BrCH₂COPh, PPh₃, Na₂CO₃, o.n., 100 °C].

J coupling between alkene hydrogens in the product **8** is 15.4 Hz, that is in the range 12 – 18 Hz that is typical for *E* geometry.²⁸⁷

The two different types of uracil to be coupled to the central linker through Wittig reaction were then obtained from a doubly protected 5-formyluracil derivative, using Dod-protection for N(1) and Cbz-protection for N(3); this can be used as a precursor for both units.



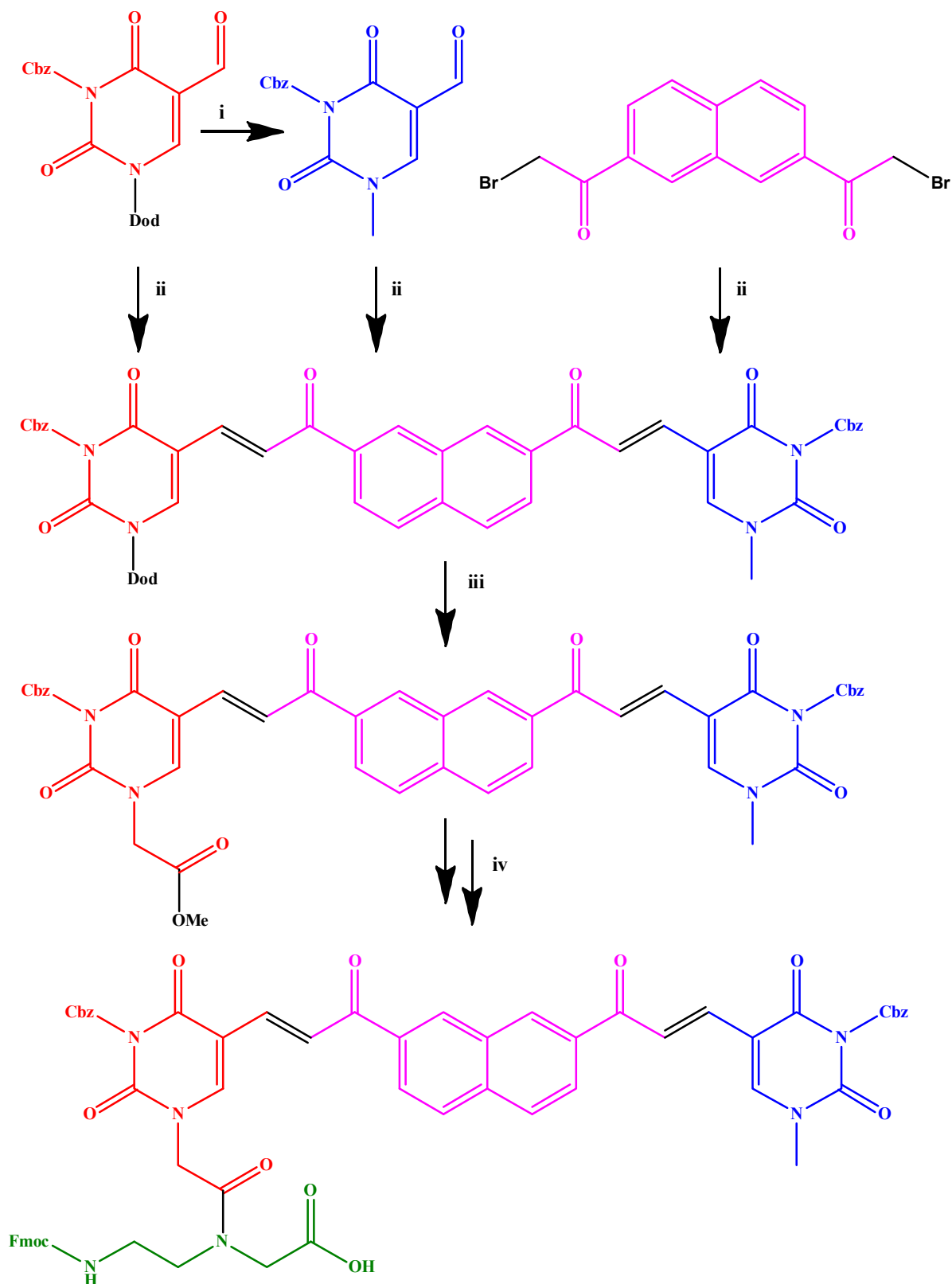
Scheme 5.4: Synthesis of 1-Dod-3-Cbz-5-formyl uracil. (i) [Dod-Cl, K₂CO₃, dry DMF, 1 h, r.t.]; (ii) [Cbz-Cl, Dod-Cl, K₂CO₃, dry DMF].

Compound **6** was protected with Dod chloride to obtain product **9**. Attack in N(1) position was favored since Dod is a bulky group. Using NMR it was possible to confirm the presence of desired product thanks to disappearance of the coupling between hydrogen in N(1) position and that one in C(6). The crude product was then protected with Cbz chloride to obtain final product **10**. Yield was particularly low probably due to oxidation of the aldehyde during purification by flash chromatography. Cbz and Dod are both cleaved in strong acidic media, but Cbz is removed under harsh conditions. Therefore is possible to deprotect N(1) position using a solution of TFA and DCM without cleaving Cbz group and thus introducing alkyl group of interest. In this way it will be possible to initially obtain uracil with methylated N1 position and then, after Wittig reactions, to insert methylenecarboxylic linker for coupling between base and backbone (Scheme 5.5).

5.7. Conclusions and future work

We have applied Molecular Dynamics simulations for a virtual screening of modified dimeric bases analogues. This is a novel approach in which rational design is used together a dynamic testing of system from a molecular point of view. Examples explored showed difficulty in predicting modification effects, since even small modifications led to completely different results. With this novel strategy was possible to find a promising candidate for adenine binding, saving time and money in experimental trials. Certainly also simulations have an intrinsic cost,²⁸⁸ but the price has rapidly decreased during the years due to progressive increased computational power. We believe that this may lead, in future, to a more routinely use of simulations as support to organic synthesis design and understanding.

We have also synthesized building blocks to be used for obtaining final monomer. Synthetic pathway to be carried starting from building blocks produced in the present thesis is reported in the scheme 1.1. Completion of synthesis will be carried in the host laboratory, and represent one of the major goals of European Project ULTRAPLACAD.²⁸⁹



Scheme 5.5: Proposed synthetic pathway to obtain Mod-4 monomer. (i) Deprotection of Dod moiety, with 40% TFA solution in DCM, followed by N(1) alkylation with iodo methane to obtain second uracil; (ii) Wittig reaction between central linker **5** and two different uracil units; (iii) Dod removal in the same conditions of i and alkylation with bromo acetate; (iv) hydrolysis of the linker ester with 1M NaOH, coupling with the backbone (compound **3** of chapter 4) and hydrolysis of the final monomer with Ba(OH)₂.

5.8. Experimental Section

General. Reagents were purchased from Sigma-Aldrich, Fluka, Merck, Carlo Erba, TCI Europe, LINK and were used without further purifications. TLC were run on Supelco 56524-25EA silica gel on aluminium foils with fluorescence indicator (254 nm). Column chromatography was performed as flash chromatography on Merck silica 60 (0.040 – 0.063) under 1.1-1.2 bar of air pressure. NMR spectra were obtained on Bruker Avance 300 MHz or 400 MHz spectrometers. Chemical shifts are presented as δ values and expressed in ppm units. FT-IR spectra were recorded on Thermo Nicolet 5700. ESI-MS spectra were recorded on Acquity Ultra Performance LC. GC-MS spectra were recorded on HP instrument (oven 6890N, mass detector 5973 and column SE-52). Melting points were measured with GALLENKAMP melting point apparatus.

Ab initio and Molecular Dynamics simulations. Calculations were performed on Brutus cluster, the central high-performance cluster of ETH Zurich,²⁹⁰ or on CSCS servers (Dora²⁹¹ and Mönch)²⁹². Ab initio calculations were performed by means of Gaussian G09 D1 using 8 cores. Molecular Dynamics simulation were performed using 32 – 64 cores and were conducted in the conditions reported in section 2.5.

Naphthalene-2,7-diyl bis(trifluoromethanesulfonate) (1). 2,7-dihydroxy naphthalene (500.5 mg, 3.13 mmol) and $(\text{CF}_3\text{SO}_2)_2\text{NPh}$ (2.344 g, 6.56 mmol) were dispersed in DCM (5 ml) and then NEt_3 (913 μl , 6.56 mmol) was slowly added at 0°C. Temperature was raised to room temperature and mixture stirred for 3 h. Reaction mixture was diluted with DCM (35 ml) and then extracted with KHSO_4 . The water phase was extracted with DCM (3 x 40 ml). The organic layers were combined, dried over sodium sulfate and evaporated under vacuum to afford pure product (90 %, 1.19 g).

(1) $^1\text{H-NMR}$ (400 MHz, 25°C, CDCl_3) δ (ppm): 8.03 (d, 2H, $J = 9.2$ Hz, CH naphth.), 7.84 (d, 2H, $J = 2.4$ Hz, CH naphth.), 7.50 (dd, 2H, $J_1 = 9.1$ Hz, $J_2 = 2.4$ Hz, CH naphth.). $^{13}\text{C-NMR}$ (100 MHz, 25°C, CDCl_3) δ (ppm): 148.3, 132.1, 131.0, 130.8, 130.0, 121.2, 119.5. FT-IR ν (cm^{-1}) 1413, 1211, 1187, 1135, 1111. Full characterization is reported in literature.²⁹³

2,7-bis(trimethylsilyl)ethynyl)naphthalene (2). Vacuum-nitrogen cycles were performed on Schlenk tube containing compound **1** (814.6 mg, 1.91 mmol) followed by addition of dry DMF and

solution was stirred at room temperature. Then, in the following order, trimethylsilylacetylene (641.1 μ l, 4.78 mmol), Pd(PPh₃)₂Cl₂ (63.7 mg, 0.10 mmol), CuI (17.3 mg, 0.10 mmol), NEt₃ (1.14 ml, 8.60 mmol) were added to the solution. When palladium was added to solution it became yellow, then with copper it turned orange and finally with triethylamine it went back to the yellow color. Temperature was raised to 40°C and solution stirred overnight. At the end of the reaction, mixture was diluted with AcOEt (200 ml) and washed with saturated KHSO₄ (2 x 200 ml), NaHCO₃ (2 x 200 ml), and brine (200 ml). The organic layer was dried over sodium sulfate and evaporated under vacuum. The crude was purified by flash column chromatography on silica gel with hexane as eluent (R_f = 0.1) to afford pure product (65%, 115 mg).

(2) ¹H-NMR (400 MHz, 25°C, CDCl₃) δ (ppm): 7.93 (s, 2H, CH naphth.), 7.74 (d, 2H, J = 8.4 Hz, CH naphth.) 7.52 (d, 2H, J = 8.6 Hz, CH naphth.), 0.31 (s, 18H, CH₃-Si). ¹³C-NMR (100 MHz, 25°C, CDCl₃) δ (ppm): 132.3, 131.7, 129.5, 127.7, 121.2, 105.0, 95.2, 0.0. MS (EI, 70 eV) m/z: 320.20 (M⁺, 50), 321.20 (18), 306.2 (35), 305.20 (100). Spectra were compared with those reported in literature.²⁹⁴

2,7-diethynylnaphthalene (3). Compound **2** (380 mg, 1.19 mmol) was dissolved in THF (25 ml) followed by addition of 1M solution of NaOH (25 ml). The mixture was vigorously stirred for 2 h and then THF was removed under vacuum. Product was extracted with DCM from water phase to afford, after drying and evaporation, pure product (99 %, 206 mg). TLC (Silica gel, hexane-AcOEt 9:1) R_f = 0.57.

(3) ¹H-NMR (400 MHz, 25°C, CDCl₃) δ (ppm): 7.99 (s, 2H, CH naphth.) 7.79 (d, 2H, J = 8.6 Hz, CH naphth.), 7.56 (dd, 2H, J₁ = 8.5 Hz, J₂ = 1.5 Hz, CH naphth.), 3.20 (s, 2H, CH alkyne). ¹³C-NMR (100 MHz, 25°C, CDCl₃) δ (ppm): 132.6, 132.3, 132.0, 129.6, 127.9, 120.3, 83.6, 78.1. MS (EI, 70 eV) m/z: 176.10 (M⁺,100), 177.10 (17), 150.10 (12). FT-IR ν (cm⁻¹) 2987, 2889, 1687, 1629, 1429. Decomposes above 250 °C.

1,1'-(naphthalene-2,7-diyl)diethanone (4a). Compound **3** (70 mg, 0.397 mmol) and Fe₂(SO₄)₃ x 5H₂O (16.7 mg, 0.03 mmol) were dissolved in glacial acetic acid (3 ml) and stirred at 95°C for 24 h. Reaction was followed by GC-MS. At the end of reaction, the mixture was diluted with AcOEt (150 ml) and washed with saturated NaHCO₃ solution (150 ml) and brine (3 x 100 ml). Organic layer was dried over sodium sulfate and evacuated under vacuum. Crude product was purified by flash column

chromatography on silica gel with hexane-AcOEt 9:1 as eluent ($R_f = 0.24$) to afford pure product (75%, 63 mg).

(4a) $^1\text{H-NMR}$ (400 MHz, 25°C, CDCl_3) δ (ppm): 8.61 (bs, 2H, CH naphth.), 8.17, (dd, 2H, $J_1 = 8.7$, $J_2 = 1.6$ Hz, CH naphth.), 7.96 (d, 2H, $J = 8.7$ Hz), 2.77 (s, 6H, Me). $^{13}\text{C-NMR}$ (100 MHz, 25°C, CDCl_3) δ (ppm): 197.6, 137.7, 135.3, 131.8, 131.5, 128.4, 126.5, 26.7. MS (EI, 70 eV) m/z : 212.17 (M^+ , 50), 197.10 (100), 169.10 (25), 154.10 (35), 126.10 (24). FT-IR ν (cm^{-1}) 2921, 2849, 1685, 1665, 1625, 1460, 1420, 1353, 1284, 1273, 1249, 1225, 1185, 1132. m.p. 108 – 110 °C.

1-(7-ethynynaphthalen-2-yl)ethanone (4b). The procedure was the same as one used to synthesize product **4a**, except that reaction time was 5 h. After column chromatography pure product was obtained (42%, 47 mg). Additionally, product **4a** was also collected as a byproduct (51 %, 62 mg).

(4b) $^1\text{H-NMR}$ (400 MHz, 25°C, CDCl_3) δ (ppm): 8.43 (s, 1H, CH naphth.), 8.16 (s, 1H, CH naphth), 8.07 (dd, 1H, $J_1 = 8.6$ Hz, $J_2 = 1.7$ Hz, CH naphth.), 7.90 (d, 1H, $J = 8.6$ Hz, CH naphth.), 7.85 (d, 1H, $J = 8.5$ Hz, CH naphth.), 7.66 (dd, 1H, $J_1 = 8.5$ Hz, $J_2 = 1.6$ Hz, CH naphth.), 3.22 (s, 1H, CH alkyne), 2.75 (s, 3H, Me). $^{13}\text{C-NMR}$ (100 MHz, 25°C, CDCl_3) δ (ppm): 197.8, 135.2, 135.1, 133.7, 132.1, 131.1, 129.8, 128.4, 127.9, 125.0, 120.6, 83.3, 78.3, 26.7. MS (EI, 70 eV) m/z : 194.10 (M^+ , 64), 179.10 (100), 151.10 (80) 150.10 (36). FT-IR ν (cm^{-1}) 3225, 2922, 2850, 1677, 1623, 1595, 1563, 1456, 1413, 1366, 1351, 1274, 1259, 1224, 1192, 1164. m.p. 121 – 122 °C.

1,1'-(naphthalene-2,7-diyl)bis(2-bromoethanone) (5). Compound **4a** (30 mg, 0.13 mmol) was dissolved in CH_3CN (10 ml) in together with *p*-toluenesulfonic acid (80.5 mg, 0.39 mmol). NBS (50.2 mg, 0.26 mmol) was dissolved in CH_3CN (600 μl) and added to the solution and stirred for 2 h. Mixture was diluted with DCM and washed with brine two times. Crude product was purified by flash column chromatography on silica gel with hexane-DCM 5:95 as eluent to afford pure product (75%, 39.2 mg).

(5). $^1\text{H-NMR}$ (400 MHz, 25°C, CDCl_3) δ (ppm): 8.68 (s, 2H, CH naphth.), 8.21 (dd, 2H, $J_1 = 8.7$ Hz, $J_2 = 1.7$ Hz, CH naphth.), 8.02 (d, 2H, $J = 8.7$ Hz, CH naphth.), 4.60 (s, 4H, CH_2). $^{13}\text{C-NMR}$ (100 MHz, 25°C, CDCl_3) δ (ppm): 190.9, 138.2, 132.4, 132.7, 131.7, 128.9, 127.3, 30.4. FT-IR ν (cm^{-1}) 2991, 2944, 1692,

1670, 1623, 1458, 1418, 1389, 1335, 1279, 1235, 1196, 1169, 1151, 1132, 1100, 1023, 1012. m.p. 155 – 177.

5-formyl uracil (6). 5-hydroxymethyl uracil (500 mg, 3.52 mmol) was dispersed in water (13 ml) and reaction carried at 70°C. Cerium ammonium nitrate was then added (4.8 g, 2.5 eq) and temperature increased to 90°C. Reaction was followed by color change from orange/yellow to transparent and after 1 h reaction was complete. Reaction mixture was cooled and kept in the refrigerator for 4 h to obtain a white precipitate that was filtered and washed with cold water to afford pure product (74%, 363 mg).

(6) ¹H-NMR (300 MHz, 25°C, d₆-DMSO) δ(ppm): 11.89 (bs, 1H, NH uracil), 11.50 (s, 1H, NH uracil), 9.74 (s, 1 H, CHO), 8.13 (d, 1H, J = 6.1 Hz, CH uracil). ¹³C-NMR (75 MHz, 25°C, d₆-DMSO) δ (ppm): 186.9, 162.9, 150.9, 149.8, 110.5. FT-IR ν (cm⁻¹) 3031, 2844, 1762, 1685, 1625, 1598, 1494, 1443, 1417, 1332, 1260, 1227, 1171. m.p. decomposes at 270-280 °C.

1,3-di Dod- 5- formyl uracil (7). Compound **6** (34.3 mg, 0.25 mmol) was dissolved in dry DMF (2 ml) with 4,4'-(chloromethylene)bis(methoxybenzene) (Dod chloride, 160.9 mg, 0.61 mmol) and K₂CO₃ (84.5 mg, 0.61 mmol) and stirred at room temperature for 4 h. Mixture was then diluted with AcOEt (100 ml) and washed with brine (2 x 100 ml). Organic phase was dried over sodium sulfate and evaporated under vacuum. Crude product was purified by column chromatography on silica gel with hexane-AcOEt 8:2 as eluent (R_f = 0.2) to afford pure product (94%, 137.2 mg).

(7) ¹H-NMR (300 Mhz, 25°C, CDCl₃) δ(ppm): 9.99 (s, 1H, CHO), 7.95 (s, 1H, CH uracil), 7.31 (s, 1H, CH Dod), 7.27 (d, 4H, J = 8 Hz, CH aromatic Dod), 7.04 (d, 4H, J = 8.7 Hz, CH aromatic Dod), 6.95 (s, 1 H, CH Dod), 6.92 (d, 4H, J = 8.7 Hz, CH aromatic Dod), 6.84 (d, 4H, J = 8.7 Hz, CH aromatic Dod), 3.84 (s, 6H, OMe), 3.81 (s, 6H, OMe).

(E)-1,3-bis(bis(4-methoxyphenyl)methyl)-5-(3-oxo-3-phenylprop-1-en-1-yl)pyrimidine-2,4(1H,3H)-dione (8). Bromo acetophenone (51 mg, 0.25 mmol) was dissolved in toluene and stirred with PPh₃ (132 mg, 0.50 mmol) for 30 min at 50°C. Aldehyde **7** (99.4 mg, 0.17 mmol) was then added together with Na₂CO₃ (33.4 mg, 1.88 eq), temperature raised to 100°C and reaction carried overnight. Toluene was removed under vacuum and solid dissolved in AcOEt (100 ml) and washed with brine

(2 x 100 ml). Organic layer was dried over sodium sulfate and evaporated under vacuum. Crude product was purified by gradient flash column chromatography on silica gel (hexane, hexane-AcOEt 8:2, 7:3) to afford pure product (95%, 115 mg).

(8) ¹H-NMR (300 MHz, 25°C, d₆-DMSO) δ(ppm): 8.13 (s, 1H, CH uracil), 8.08 (d, 1H, J = 15.6 Hz, CH alkene), 7.91 (d, 2H, J = 7.2 Hz, CH aromatic), 7.64 (t, 1H, J = 7.2 Hz, CH aromatic), 7.55 (t, 2H, J = 7.9 Hz, CH aromatic), 7.50 (d, 1H, J = 15.1 Hz, CH alkene), 7.22 (s, 1H, CH Dod), 7.19 (d, 4H, J = 8.8 Hz, CH aromatic Dod), 7.18 (d, 4H, J = 8.8 Hz, CH aromatic Dod), 6.98 (d, 4H, J = 8.8 Hz, CH aromatic Dod), 6.88 (s, 1H, CH Dod), 6.87 (d, 4H, J = 8.7 Hz, CH aromatic Dod), 3.77 (s, 6H, OMe), 3.74 (s, 6H, OMe). ¹³C-NMR (75 MHz, 25°C, CDCl₃) δ (ppm): 190.8, 159.7, 158.8, 138.1, 137.3, 132.8, 130.4, 129.9, 129.6, 129.2, 128.6, 128.5, 122.4, 114.6, 113.6, 63.3, 55.4, 55.2. FT-IR ν (cm⁻¹) 1663, 1654, 1510, 1442, 1247, 1174. m.p. 99-101 °C.

1-(bis (4-methoxyphenyl) methyl) -2,4- dioxo- 1,2,3,4- tetrahydropyrimidine -5- carbaldehyde (9).

Compound **6** (140 mg, 1.00 mmol) was dissolved in dry DMF (7 ml) and stirred under nitrogen atmosphere, at room temperature, for 1 h, with Dod chloride (289 mg, 1.10 mmol) and potassium carbonate (151.8 mg, 1.10 mmol). At the end of reaction, crude product was diluted with AcOEt (125 ml) and washed with brine (2 x 100 ml). Organic layer was dried over sodium sulfate and evacuated under vacuum. Product was obtained by re-crystallization in hot acetate (30%, 110 mg). TLC (Silica gel, hexane/AcOEt 5:5) R_f = 0.45.

(9) ¹H-NMR (400 MHz, 25°C, d₆-DMSO) δ(ppm): 11.89 (s, 1H, NH uracil), 9.72 (s, 1H, CHO), 7.79 (s, 1H, CH uracil), 7.14 (d, 4H, J = 8.7 Hz, CH aromatic Dod), 6.98 (d, 4H, J = 8.8 Hz, CH aromatic Dod), 6.84 (s, 1H, CH Dod), 3.77 (s, 6H, OMe). ¹³C-NMR (100 MHz, 25°C, d₆-DMSO) δ (ppm): 186.7, 162.2, 159.6, 150.4, 148.0, 130.2, 129.7, 114.8, 110.9, 62.9, 55.7. FT-IR ν (cm⁻¹) 3012, 2840, 1729, 1689, 1608, 1597, 1583, 1514, 1488, 1462, 1442, 1396, 1370, 1324, 1301, 1251, 1180, 1118, 1077, 1031. m.p. 198 – 199 °C.

1-Dod-3-Cbz-5-formyl uracil (10). Compound **6** (560.4 mg, 4.0 mmol) was dissolved in dry DMF (10 ml) and stirred under nitrogen atmosphere, at room temperature, for 1 h, with Dod chloride (1.1561 g, 4.4 mmol) and potassium carbonate (607.2 mg, 4.4 mmol). At the end of the reaction Cbz chloride (1.42 ml, 10.0 mmol) was added and solution stirred overnight. Mixture was diluted with AcOEt (150

ml) and washed with brine (3 x 100 ml). Organic layer was dried over sodium sulfate and evacuated under vacuum. Crude product was purified by gradient flash column chromatography on silica gel (hexane-AcOEt 9:1, 8:2, 7:3, 6:4, 0:1) to afford pure product (30%, 600 mg).

10 $^1\text{H-NMR}$ (400 MHz, 25°C, $\text{d}_6\text{-DMSO}$) δ (ppm): 9.78 (s, 1H, CHO), 7.84 (s, 1H, CH uracil), 7.33 – 7.20 (m, 5H, CH aromatic Cbz), 7.16 (d, 4H, $J = 8.3$ Hz, CH aromatic Dod), 6.98 (d, 4H, $J = 8.3$ Hz, CH aromatic Dod), 6.91 (s, 1H, CH Dod), 5.02 (s, 2H, CH_2 Cbz), 3.77 (s, 6H, OMe). $^{13}\text{C-NMR}$ (100 MHz, 25°C, $\text{d}_6\text{-DMSO}$) δ (ppm): 186.90, 161.19, 159.64, 150.74, 146.61, 136.86, 130.24, 129.56, 128.83, 128.06, 127.78, 114.84, 110.18, 64.29, 55.67, 44.46. FT-IR ν (cm^{-1}) 3005, 2961, 2839, 1718, 1698, 1663, 1602, 1586, 1511, 1496, 1462, 1450, 1443, 1420, 1380, 1353, 1330, 1310, 1300, 1279, 1248, 1229, 1177, 1146, 1105, 1022. m.p. 170 – 172 °C.

6. Modelling and synthesis of hydrolytic unit for reactive PNAs.

6.1. Introduction

As briefly explained in chapter 1, the introduction of hydrolytic units on systems able to discriminate target DNA or RNA sequences would be extremely useful in order to obtain artificial nucleases. First trials in this direction have been done modifying oligonucleotides.²⁹⁵ Catalytic moieties have been inserted both on bases or backbone. An example of modified base is reported in Figure 6.1 A, in which terpyridine unit is bound to uracil through a spacer.²⁹⁶ In Figure 6.1 B a second example in which a lutetium catalytic complex was bound at the 5' backbone end of a 15 mer oligonucleotide.²⁹⁷

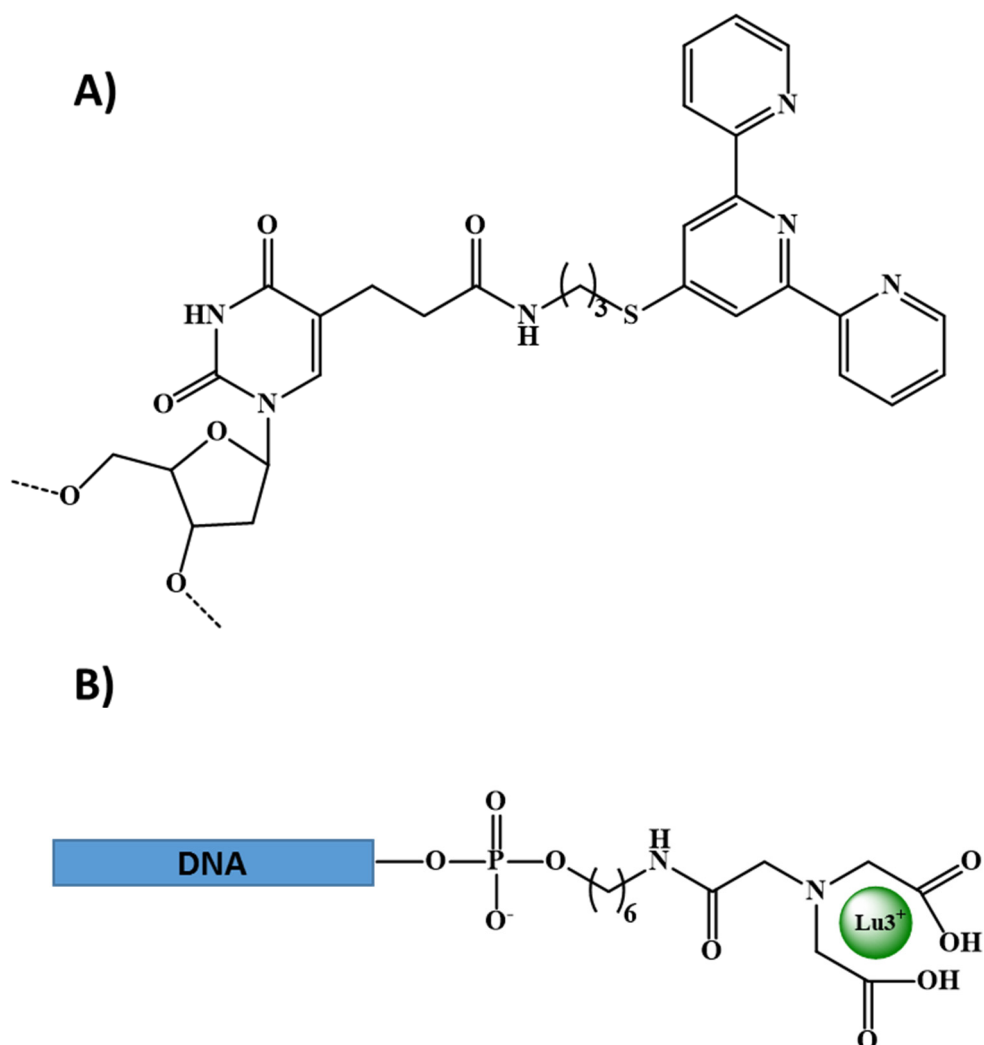


Figure 6.1: Oligonucleotides bearing hydrolytic units. A) example of base modification; B) example of backbone modification.

In a similar manner, PNAs have been also used as artificial nucleases. Stromberg and coworkers reported several examples of PNAs able to cleave RNA (Figure 6.2 A) in presence of a bulge due to

mispairing of several nucleotides on the RNA. PNA bearing phenanthroline derivative complex with Zn^{2+} and Cu^{2+} were able to hydrolyze RNA, preserving binding and selectivity typical of PNA.^{153,152} They have also synthesized a metal free nuclease (Figure 6.2 B) coupling a tris(2-aminobenzimidazole) derivative to PNA.²⁹⁸ Hydrolysis occurred in this case without necessity of a bulge.

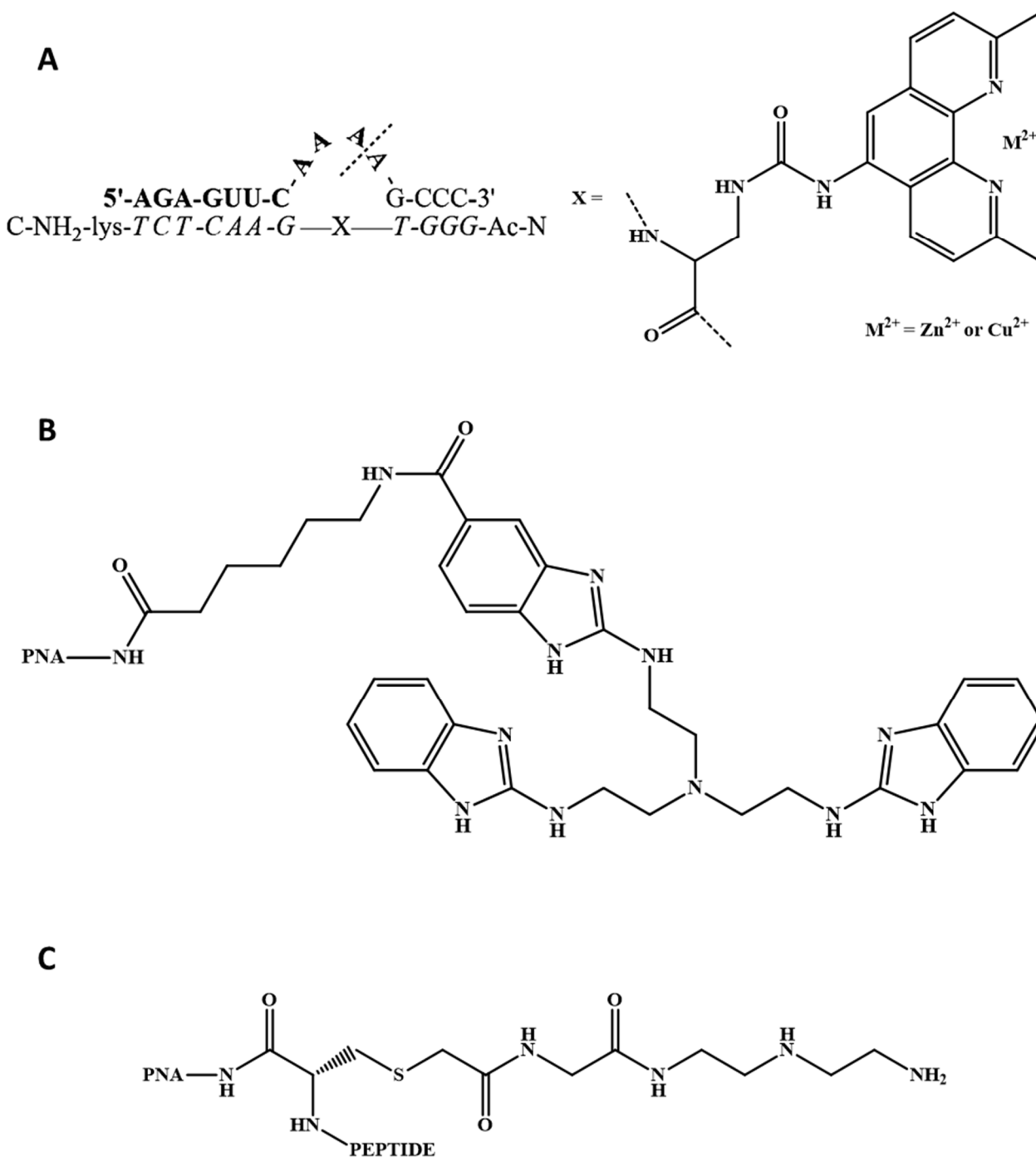


Figure 6.2: **A)** PNA bearing phenanthroline derivative interacting with RNA bulge; **B)** PNA bearing tris(2-aminobenzimidazole) derivative; **C)** example of DETA modified PNA.

Other examples of PNAs bearing hydrolytic moieties have been reported by Boom and coworkers. In particular, they attached diethylenetriamine (DETA) to N terminus of PNA backbone using different spacers (Figure 6.2 C).^{299,300}

One of the most studied and effective catalytic unit used for artificial metallo-nucleases is 1,4,7-triazacyclononane (TACN) which has been used in combination with Cu^{2+} and Zn^{2+} ; binding to this ligand is very strong, thus preventing metal release; the hydrolytic properties of metal-, in particular of zinc-complexes have been well studied and this unit has been coupled to different molecules retaining its activity.^{301,302} Here we report the modelling studies and the synthesis of a modified triazacyclononane (Figure 6.3), with a carboxylic moiety that allows linkage to PNA (1,4,7-triazacyclononane-N-acetate, TACNA).

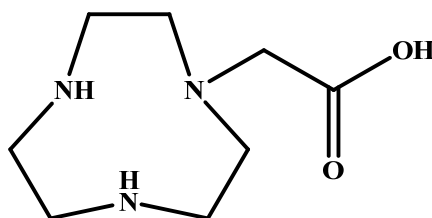


Figure 6.3: Chemical structure of the TACN derivative TACNA.

6.2. *Ab initio* calculation

TACNA is less studied complex compared to TACN,^{303,304} but its hydrolytic properties have been tested with copper³⁰⁵ and manganese.³⁰⁶ We performed an explorative study on this compound using *ab initio* DFT calculation on zinc complex and we compared results with TACN. Initial starting geometries were generated with Avogadro, a freeware software for input preparation.³⁰⁷ In order to mimic binding with a spacer linked to PNA, we have simulated TACNA capped with an *N*-ethylamide moiety. The introduction of this group is also important to remove complex extra stabilization due to carboxylic acid which could bias the results.

Calculations were performed using Gaussian 09 D1 (G09),²⁴⁶ at B3LYP level of theory. Structures were optimized, in vacuum, initially with 3-21G* basis set, then refined with 6-31G and 6-31G*. For the Zn atom the LANL2DZ core potential was used. To obtain more reliable results, the final optimized structures were refined with implicit solvent (Polarizable Continuum Model, PCM)³⁰⁸, implemented in G09, using 6-31G* as basis set. Finally in order to fill vacant coordination position

we inserted an explicit molecule of water. RESP charges (Figure 6.4) were calculated with procedure explained in chapter 2.

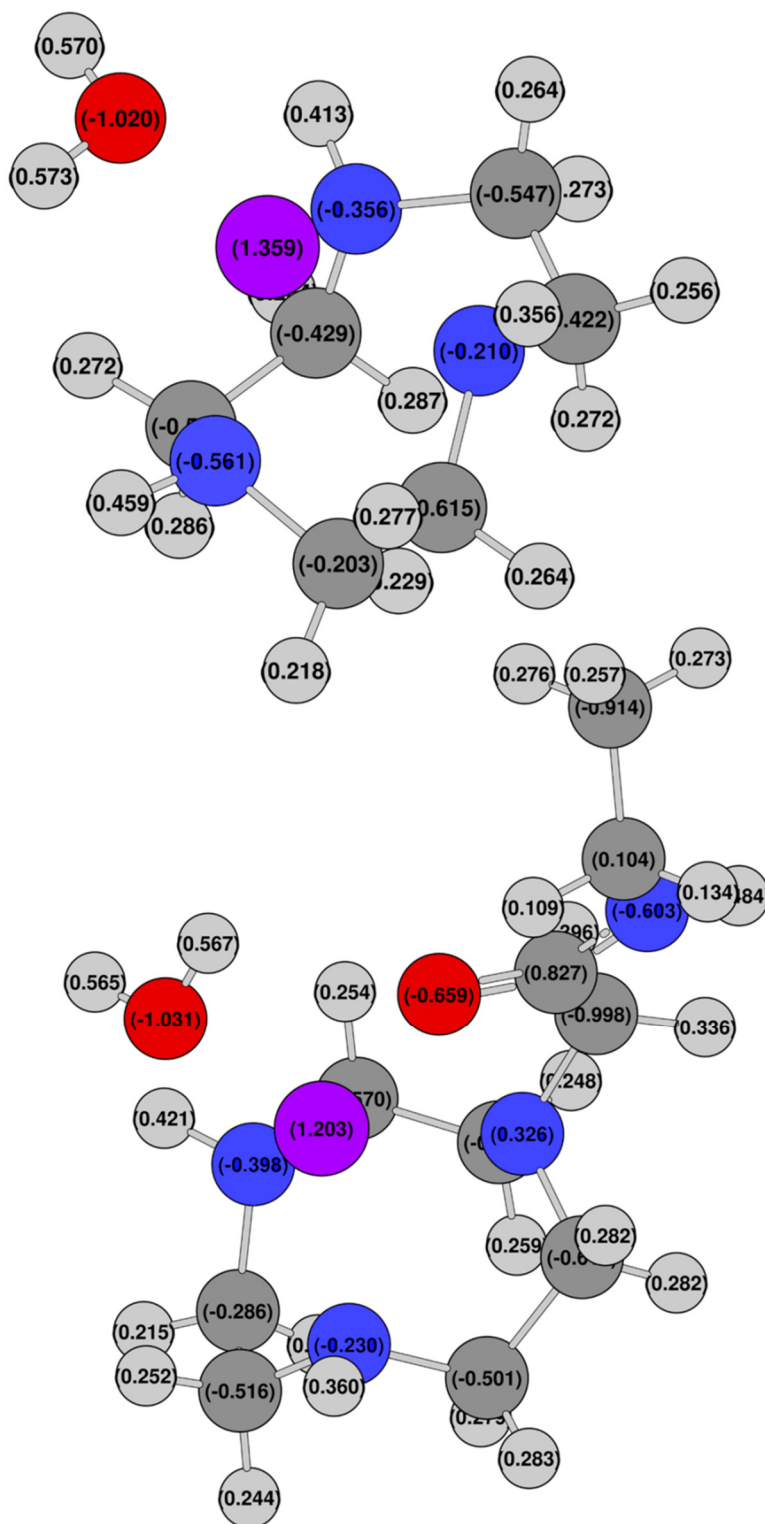
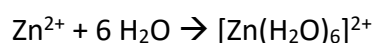


Figure 6.4: RESP charges for TACN (top) and TACNA (bottom) complexes with ZINC (violet).

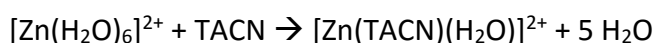
Acetamide group on TACNA affects the charge distribution of the cycle, changing in particular that of bonded nitrogen (from -0.210 to +0.326). This may slightly reduce binding properties compared

to TACN, but the effect is counterbalanced by an additional interaction with carbonyl group. Partial charge on zinc was reduced (from 1.359 to 1.203), but fortunately this poorly affected polarization of coordinated water molecule. We also considered stability of the complexes by evaluation of free energy variation. For Zinc we decided to consider an octahedral complex $[\text{Zn}(\text{H}_2\text{O})_6]^{2+}$, since in water it should be preferred to the tetrahedral one^{309,310}.

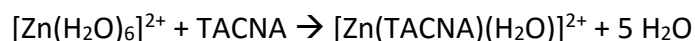
Reactions involved are then:



and consequently



and



From ΔG^0 value it is possible to calculate stability constants β of complexes ($\log \beta_{\text{TACNA}} = 42.03$, $\log \beta_{\text{TACN}} = 46.4$). These values in themselves are not fully meaningful since to have more accurate and realistic results equilibrium between species, second sphere coordination and other factors should be considered. However, assuming that approximations are equal in both cases, their ratio represents relative stability of the complexes:

$$\frac{\log \beta_{\text{TACNA}}}{\log \beta_{\text{TACN}}} = 0.92$$

From these results we deduced that complex with TACNA has a stability comparable to TACN, and considering that experimental $\log \beta_{\text{TACN}}$ is 7.18,³¹¹ we expect a $\log \beta_{\text{TACNA}}$ of 6.61, making it suitable for zinc complexation. Moreover coordinated water molecule exhibited polarization similar to that one found in unmodified cycle, suggesting retention of hydrolytic properties. Optimized structures obtained from this study are reported in Appendix 8.8.

6.3. Synthesis of modified TACNA

In Figure 6.5 it is reported structure of modified TACNA synthesized during PhD activity. Protection of amino groups was necessary in order to avoid side reactions during PNA synthesis.

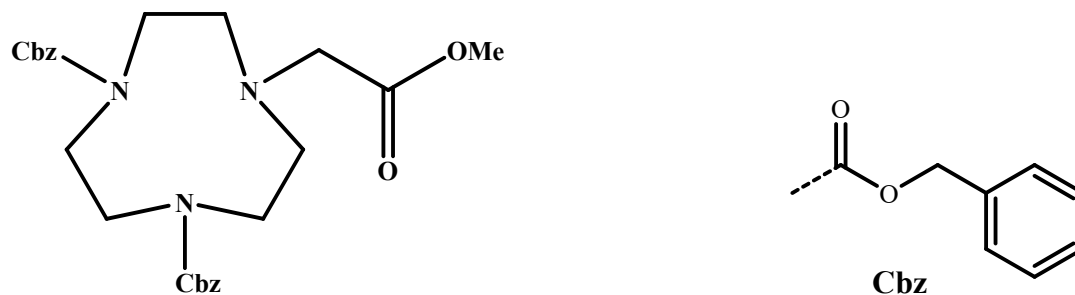
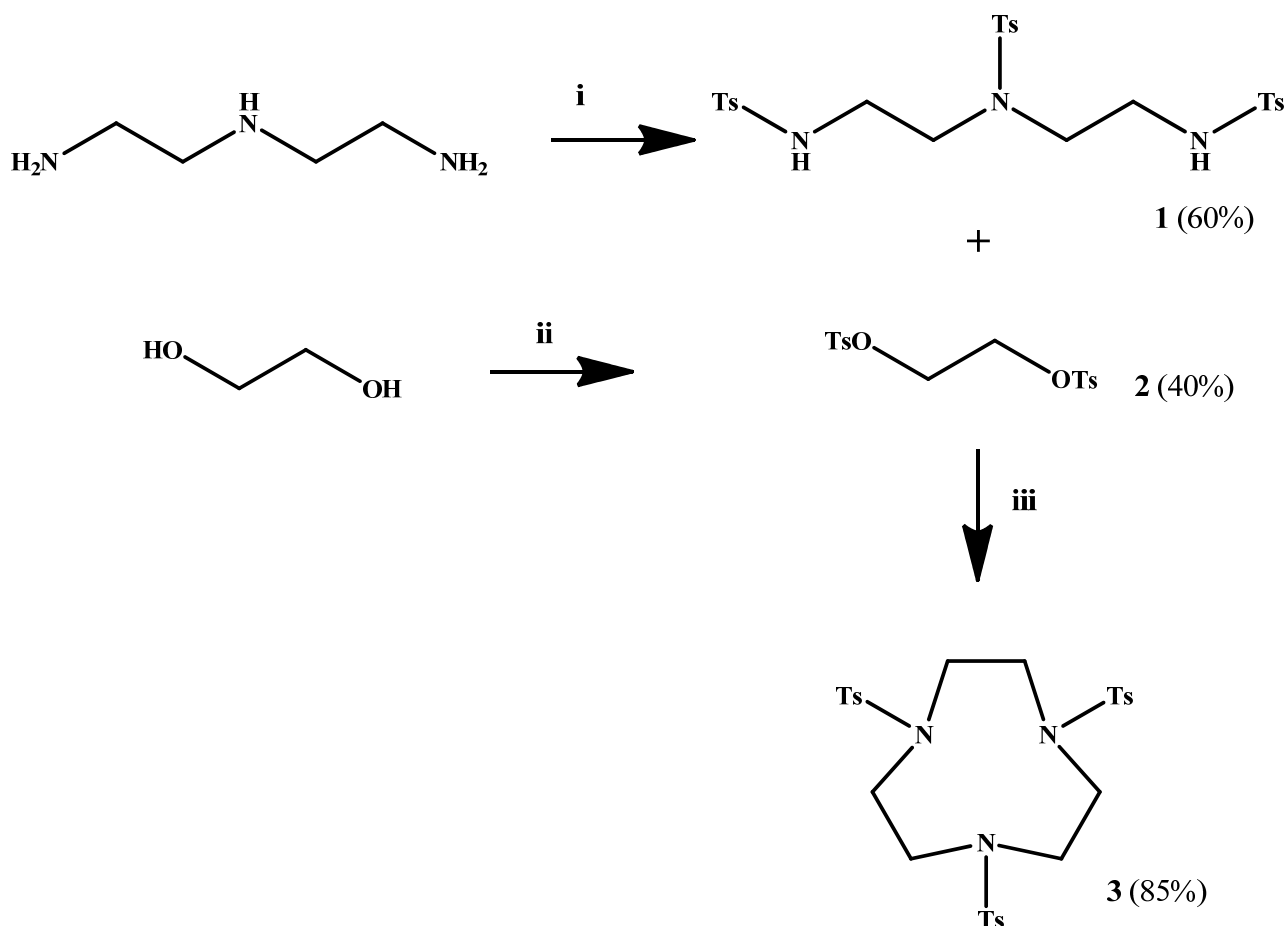


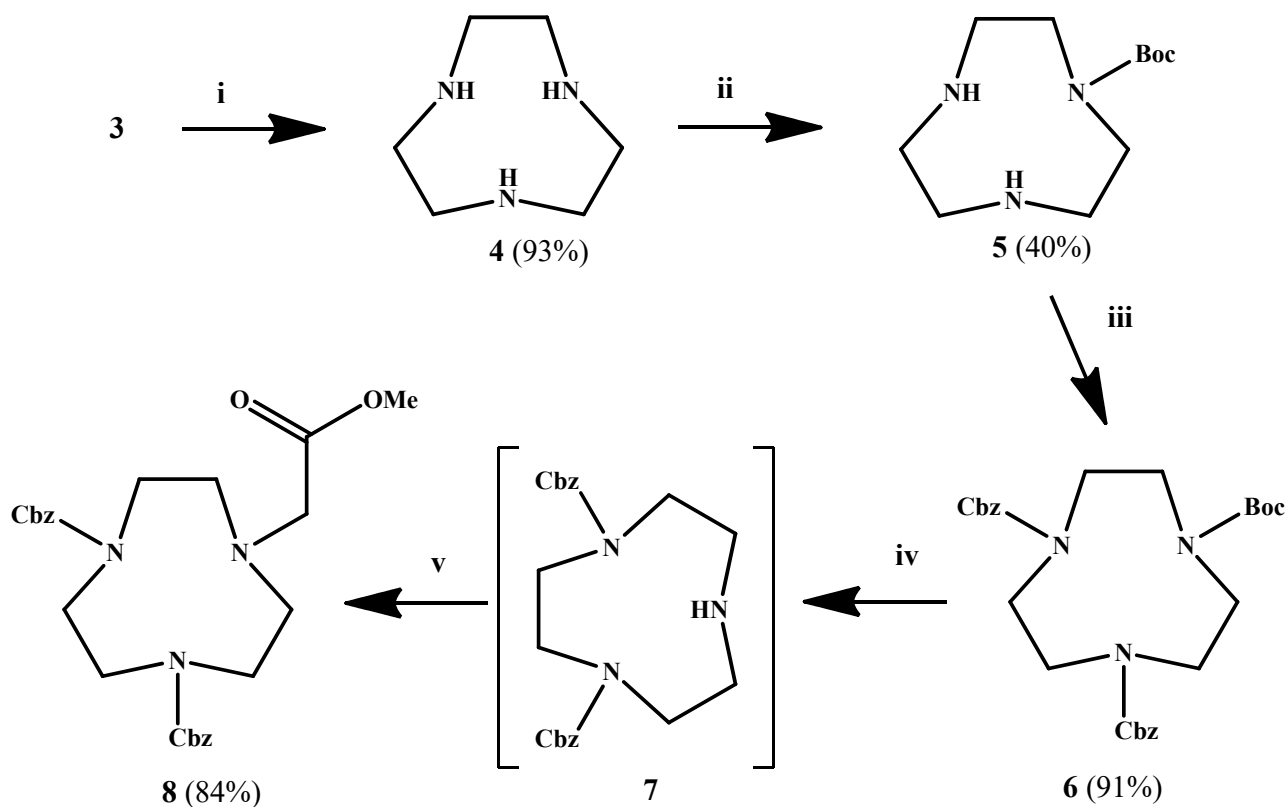
Figure 6.5: Designed hydrolytic unit for coupling with PNA. Original TACNA structure was protected with Cbz in order to avoid side reaction during PNA chain growth.



Scheme 6.1: Synthesis of TACN 3Ts. (i) [K₂CO₃, TsCl, dry DMF/DCM 1:1, o.n., r.t.]; (ii) [(a) TsCl, K₂CO₃, pestle 40 min; (b) NaOH, pestle 10 min]; (iii) [N₂, CsCO₃, dry DMF, o.n. r.t.].

Synthesis of TACN was performed starting from two building blocks: diethylenetriamine tritosilate (DETA 3Ts) and glycol ditosilate.³¹² DETA 3Ts was obtained from DETA protecting amino groups with tosyl chloride and product **2** was obtained from reaction of ethylene glycol with tosyl chloride, with

a solid phase reaction. Low yields obtained in this case were probably due to presence of water in the starting reagent. In order to favor cyclization of compounds **2** and **3** over polymerization, reaction was conducted in diluted solutions. Cesium carbonate was also necessary to generate reactive nucleophile (loose ion pair).³¹²



Scheme 6.2: Synthesis of modified TACNA. (i) [(a) conc. H_2SO_4 , N_2 , 30 min, 160°C ; (b) HCl 37%, EtOH ; (c) $\text{H}_2\text{O}/\text{NaOH}$, continuously extracted with CH_3Cl]; (ii) [Boc_2O , Phosphate buffer ($\text{pH} = 6.8$), o.n., r.t.]; (iii) [Cbz chloride, K_2CO_3 , CH_3CN , o.n., r.t.]; (iv) [40% TFA/DCM , 1h, r.t.]; (v) [$\text{BrCH}_2\text{COOMe}$, K_2CO_3 , CH_3CN , o.n., r.t.].

Compound **3** was deprotected using hot concentrated sulfuric acid and the crude was precipitated as hydrochloric salt. This was then dissolved in 1M NaOH solution and continuously extracted with chloroform. Product **4** was mono protected with $(\text{Boc})_2\text{O}$ in presence of phosphate buffer ($\text{pH} = 6.8$). Without pH control, major products were the macrocycle bearing three Boc -groups and unprotected TACN, while mono- and di-protection occurred in traces; with pH control it was possible to obtain about 40% of **5** with 10% of three protected compound and traces of di protected. About 50% remained unreacted TACN. At the end of the reaction it was possible to extract with chloroform a first fraction containing Boc products and then, rising pH of the solution, to collect unreacted TACN. Product **6** was obtained protecting **5** with Cbz chloride. Treating **6** with a 40% solution of TFA

in DCM allowed selective deprotection of Boc moiety since Cbz protecting groups require harsher acidic conditions. Intermediate **7** was then made react with bromoacetate to obtain product **8**.

6.4. Molecular Dynamics simulation on Model 1 with coupled TACNA

Positioning of hydrolytic moieties on PNA is not trivial, in particular if selective cut is needed. The model developed in the previous chapter could be very useful for properly directing TACNA hydrolytic moiety towards the target phosphate groups. The second uracil of the dimeric base described in Chapter 5 could allow this catalytic unit to be precisely localized in the PNA:DNA or PNA:RNA duplex in an inner base, unlike many derivatives reported in the literature, thanks to the vicinity of the latter to the DNA/RNA strand. In order to better focus on this possibility we have performed a Molecular Dynamics simulation after introduction of the TACNA unit. The PNA:DNA duplex simulated using the Model 1 structure of Chapter 5, which was then modified to bear the hydrolytic unit linked in N1 position, in place of methyl group, through an ethyl amino linker (Figure 6.6).

From the structures obtained, the hydrolytic moiety was found to be in a favorable position and could effectively interact with target phosphate even if, as shown before, Model1 was not so efficient in adenine targeting. From simulation TACNA and Zn^{2+} presented average distances from target phosphate of respectively $3.47 \pm 0.10 \text{ \AA}$ and $1.83 \pm 0.11 \text{ \AA}$.

Interaction with the target was quite stable and durable even if one considers that Molecular Dynamics tend to overestimate coulombic interactions between charged groups.³¹³

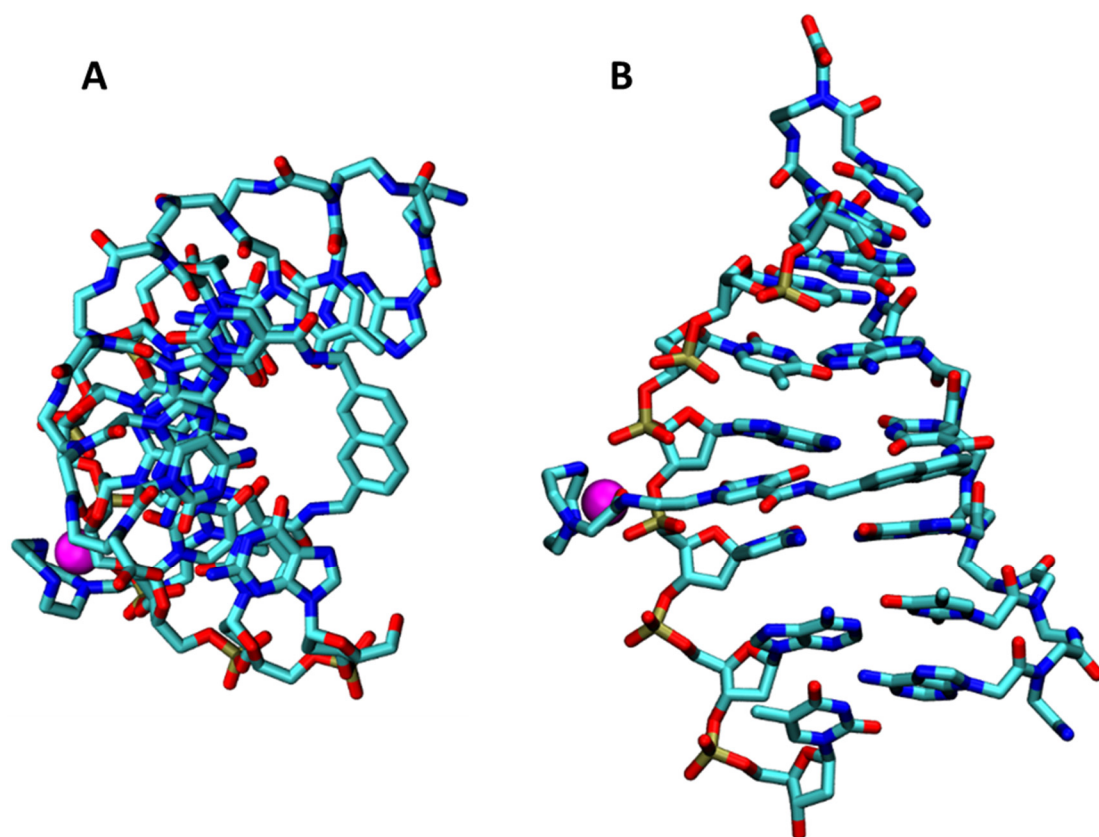
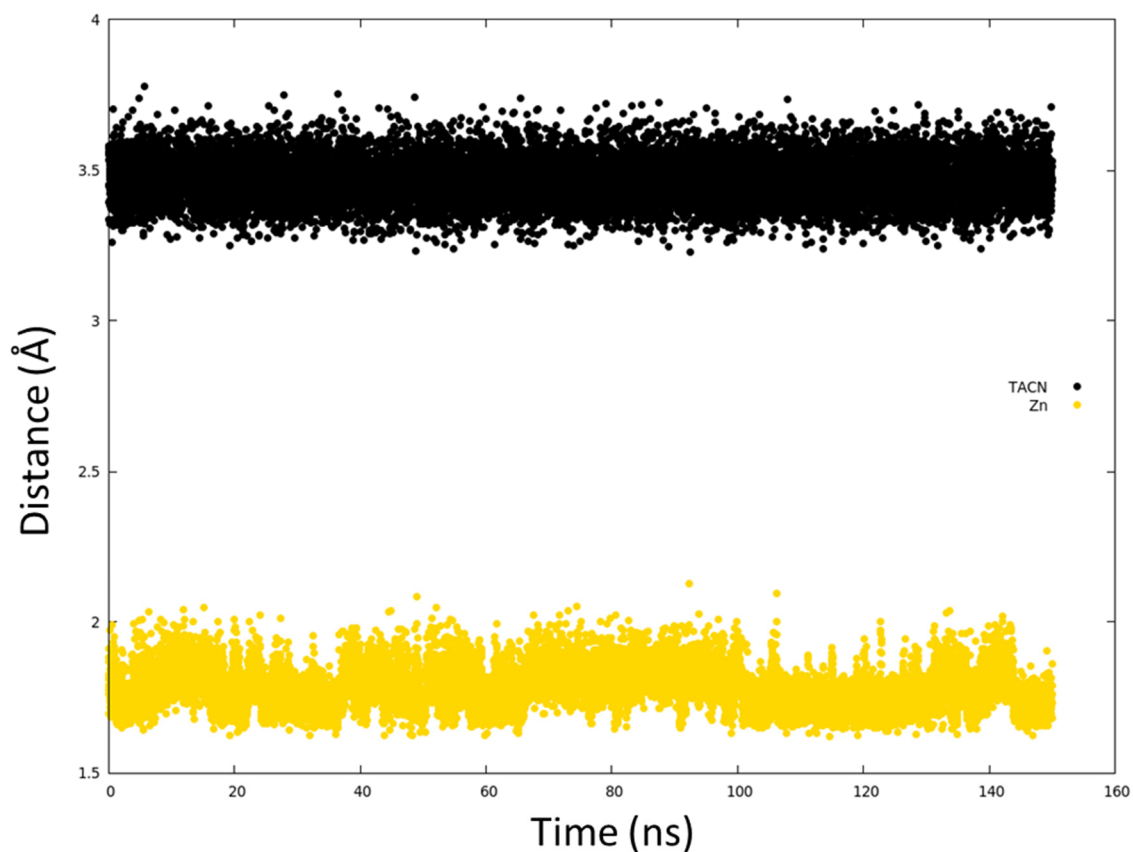


Figure 6.6: Structure of PNA:DNA duplex containing TACNA bonded to Model 1 dimeric base. **A)** top view; **B)** side view. Zn ion is shown in magenta. The system was solvated with TIP3P water in a cubic box of 66 X 63 X 74 Å³ for a total of 30100 atoms.



Graph 6.1: Distance between phosphate and TACN (black) or Zinc ion (gold).

6.5. Conclusions and future works

Ab initio calculations on TACNA complex and MD simulation on a duplex containing dimeric base linked to TACNA suggested a possible combination of these elements to obtain a favorable disposition of the catalytic unit with respect to phosphate groups of complementary DNA. This should be a starting point for the design of new artificial nucleases. The vicinity effect was evaluated using Model1 modified base (chapter 5) already tested in our lab, though this presents a constrained interaction with adenine counterparts; further studies will be performed also on Model4 nucleobase, once the synthesis of the latter is completed.

For the final experiments, product **8**, after ester deprotection, will be coupled to the second uracil base of PNA, in order to test the hydrolytic properties of the final engineered molecule.

The aim of this work is therefore, in perspective, to endow PNA not only with DNA/RNA binding properties, but also with the ability to cleave the target sequence and release it, thus acting as a real artificial nuclease. Application of a similar system can have an impact both in the production of more effective toolkits for molecular biologists, and in the ability to selectively and efficiently deactivate unwanted gene expression, as in the case of genetic diseases or cancer.

6.6. Experimental section

General. Reagents were purchased from Sigma-Aldrich, Fluka, Merck, Carlo Erba, TCI Europe, LINK and used without further purifications. TLC were run on Supelco 56524-25EA silica gel on aluminium foils with fluorescence indicator 254 nm. Column chromatography were performed as flash chromatography on Merck silica 60 (0.040 – 0.063) under 0.1-0.2 bar of air overpressure. NMR spectra were obtained on Bruker Avance 300 MHz or 400 MHz spectrometers. δ values are expressed in ppm. FT-IR were recorded on Thermo Nicolet 5700. ESI-MS spectra were recorded on a Acquity Ultra Performance LC. Melting points were measured with GALLENKAMP melting point apparatus.

Ab initio and Molecular Dynamics simulations. Calculations were performed on Brutus cluster, the central high-performance cluster of ETH Zurich.²⁹⁰ Ab initio calculation were performed by means of Gaussian G09 D1 using 8 cores. Molecular Dynamics simulation were performed using 32 cores and were conducted in the conditions reported in section 2.5.

4-methyl-N,N-bis(2-(4-methylphenylsulfonamido)ethyl)benzenesulfonamide (DETA 3Ts, 1). DETA (2g, 19.385 mmol) was vigorously stirred in dry DMF (10 ml) with K₂CO₃ (9.36 g, 67.846 mmol), at room temperature. Tosyl chloride (12.82 g, 63.971 mmol) was dissolved in DCM (30 ml) and slowly added to the mixture over a period of 2 h. Solution was kept reacting overnight. Solvent was evacuated under vacuum and white solid partitioned between brine (250 ml) and DCM (250 ml). Aqueous phase was extracted several times with DCM. Compound **1** was purified by crystallization in DCM-MeOH 1:3 (60%, 6.5 g). TLC (Silica gel, hexane-AcOEt 5:5, R_f = 0.49).

(1) ¹H-NMR (400 MHz, 25°C, CDCl₃) δ(ppm): 7.78 (d, J = 8.1 Hz, 4H, CH aromatic of external Ts), 7.64 (d, J = 8.1 Hz, 2H, CH aromatic internal Ts), 7.33 (d, J = 8.1 Hz, 4H, CH aromatic of external Ts), 7.31 (d, J = 8.1 Hz, 2H, CH aromatic internal Ts), 5.24 (t, J = 4.7, 2H, NH), 3.25 – 3.11 (m, 8H, CH₂), 2.45 (s, 9H, Me Ts). ¹³C-NMR (100 MHz, 25°C, d₆-DMSO) δ(ppm): 143.9, 143.2, 137.8, 135.8, 130.4, 130.2, 127.3, 127.0, 48.8, 42.0, 21.4. FT-IR ν (cm⁻¹) 3285, 2910, 1598, 1495, 1438, 1423, 1372, 1335, 1322, 1307, 1256, 1154, 1092, 1077. Full characterization is reported in literature.³¹⁴

Ethylene glycol di tosylate (2). Ethylene glycol (1g, 16.111 mmol), K₂CO₃ (13.4 g, 96.666 mmol) and tosyl chloride (9.2 g, 48.333 mmol) were placed in a mortar. Reagents were ground together for about 40 min (when reaction started becoming very exothermic). Excess of TsCl was quenched with NaOH (3.22g, 80.555 mmol). The resulting solid was transferred into a flask, AcOEt was added (150 ml) and kept stirring for 10 min. Organic phase was filtered on sintered glass filter (G4) and concentrated under vacuum. Solid was crystallized from hot methanol to afford pure product (40%, 2.35 g). TLC (Silica gel, hexane-AcOEt 3:7, R_f = 0.62).

(2) ¹H-NMR (300 MHz, 25°C, CDCl₃) δ(ppm): 7.75 (d, 4H, J= 8 Hz, CH aromatic), 7.36 (d, 4H, J= 8 Hz, CH aromatic), 4.20 (s, 4H, CH₂), 2.48 (s, 6H, Me). ¹³C-NMR (100 MHz, 25°C, CDCl₃) δ(ppm): 145.3, 132.3, 130.0, 128.0, 66.7, 21.7. Full characterization is reported in literature.^{315,316}

1,4,7-Triazacyclononane tritosylate (3). Compound **1** (1.3 g, 2.310 mmol) was dissolved in dry DMF (15 ml), under nitrogen. Cs₂CO₃ (1.582 g, 4.851 mmol) was added and mixture kept stirring for 1 h. Compound **2** (865.2 mg, 2.333 mmol) was dissolved in dry DMF (6 ml), slowly added over a period of 2 h and reaction conducted overnight. Crude product was divided into four centrifuge tubes (50

ml). They were filled with water and ice and kept at 4 °C for 4 h. Solid was separated by centrifuge and washed with ethanol and ethyl ether (85%, 1.2 g). TLC (Silica gel, hexane-AcOEt 5:5, R_f = 0.66).

(3) $^1\text{H-NMR}$ (400 MHz, 25°C, CDCl_3) δ (ppm): 7.72 (d, J = 8.2 Hz, 6H, CH aromatic), 7.35 (d, J = 8.2 Hz, 6H, CH aromatic), 3.44 (s, 12H, CH_2), 2.46 (s, 9H, Me). $^{13}\text{C-NMR}$ (100 MHz, 25°C, CDCl_3) δ (ppm): 143.9, 134.6, 129.9, 127.5, 51.9, 21.5. MS (ESI+) m/z : $[\text{M}+\text{Cs}]^+$ calcd. 724.06, found 724.24. FT-IR ν (cm^{-1}) 1336, 1320, 1308, 1161, 1149, 1089. m.p. 218 – 219 °C.

1,4,7-Triazacylononane (4). Compound **3** (3.4 g, 5.74 mmol) was introduced in a tube and stirred with concentrated H_2SO_4 (6 ml) at 160°C for 30 min, under nitrogen. Tube was cooled to room temperature and cold ethanol was slowly added (3 ml), followed by addition of ethyl ether (45 ml). Mixture was kept in freezer for 4 h. Precipitate was separated by centrifugation and washed with diethyl ether. Concentrated HCl was then added (5 ml) and solid sonicated till dissolved. Ethanol and diethyl ether were added again and mixture kept in freezer overnight. White solid was separated by centrifugation and washed with diethyl ether. Solid was dissolved in 1 M NaOH ($\text{pH} > 12$) and then continuously extracted with chloroform for 48 h to afford pure product (93%, 693.7 mg, viscous liquid).

(4) $^1\text{H-NMR}$ (300 MHz, 25°C, CDCl_3) δ (ppm): 2.80 (s, 12H, CH_2), 2.38 (bs, 3H, NH). $^{13}\text{C-NMR}$ (75 MHz, 25°C, CDCl_3) δ (ppm): 47.2. MS (ESI+) m/z : $[\text{M}+\text{H}]^+$ calcd. 130.13, found. 130.23. FT-IR ν (cm^{-1}) 3339, 2919, 1646, 1449, 1402, 1350, 1282, 1149.

tert-butyl 1,4,7-triazonane-1-carboxylate (5). TACN (350 mg, 2.709 mmol) was dissolved in water (150 ml) containing a phosphate buffer ($\text{pH} = 6.8$, 186 mM). Boc anhydride (1.182 g, 5.418 mmol) was dissolved in water/methanol 1:1 mixture (40 ml) and slowly added in the flask with buffered solution of TACN. Mixture was stirred overnight. Water solution was extracted with chloroform (3 x 200 ml) to separate the first fraction containing mainly three Boc protected byproduct. Water solution pH was adjusted to ~ 10 and product continuously extracted with CHCl_3 overnight to collect crude mono Boc product. Concentrated organic layer was purified using gradient flash chromatography (hexane-AcOEt 7:3, 5:5, 4:6 + 1% NH_3 , 2:8 + 1% NH_3 , AcOEt + 1% NH_3) to afford pure product (40%, 250 mg, viscous liquid). TLC (Silica gel, hexane-AcOEt 5:5, R_f = 0.33). Rising pH of water solution above 12 was possible to collect unreacted TACN.

(5) $^1\text{H-NMR}$ (300 MHz, 25°C, CDCl_3) δ (ppm): 5.73 (bs, 2H, NH), 3.75 – 2.65 (m, 12H, CH_2), 1.49 (s, 9H, ^tBu). MS (ESI+) m/z : $[\text{M}+\text{H}]^+$ calcd. 230.19, found 230.05, $[\text{2M}+\text{H}]^+$ calcd. 459.37, found 459.52, $[\text{M}-^t\text{Bu}+\text{H}]^+$ calcd. 174.12, found 173.99. FT-IR ν (cm^{-1}) 3298, 3166, 2975, 2917, 2848, 1659, 1458, 1397, 1364, 1248, 1149.

1,4-dibenzyl 7-tert-butyl 1,4,7-triazonane-1,4,7-tricarboxylate (6). Compound **5** (147.6 mg, 0.643 mmol) was dissolved in CH_3CN (15 ml) in the presence of K_2CO_3 (266.6 mg, 1.929 mmol). Benzyl chloroformate (218 μl , 1.929 mmol) was added and mixture stirred overnight. Solvent was evacuated under vacuum and rinsed with AcOEt (150 ml). Organic phase was washed with brine (3 x 125 ml), dried over Na_2SO_4 and concentrated under vacuum. The crude product was purified by gradient flash chromatography (hexane-AcOEt 75:25, 7:3, 6:4, 5:5) to afford the pure product (91%, 290 mg, viscous liquid). TLC (Silica gel, hexane-AcOEt 7:3, R_f = 0.3, with ninhydrin yielded blue spot).

(6) $^1\text{H-NMR}$ (300 MHz, 25°C, CDCl_3) δ (ppm): 7.40 – 7.29 (m, 10H, CH Cbz), 5.20 – 4.95 (m, 4H, CH_2 Cbz), 3.60 – 3.30 (m, 12H, CH_2), 1.49 – 1.43 (m, 9H, ^tBu). $^{13}\text{C-NMR}$ (100 MHz, 25°C, CDCl_3) δ (ppm): 156.4, 155.7, 136.6, 128.5, 128.1, 128.0, 80.1, 67.3, 50.7, 50.4, 50.1, 50.0, 49.9, 49.7, 49.6, 49.5, 49.4, 49.3, 49.1, 49.0, 48.9, 48.8, 48.7, 28.4. MS (ESI+) m/z : $[\text{M}+\text{H}]^+$ calcd. 498.26, found 498.40, $[\text{M}+\text{Na}]^+$ calcd. 520.24, found 520.44, $[\text{M}+\text{K}]^+$ calcd. 536.22, found 536.38, $[\text{2M}+\text{Na}]^+$ calcd. 1017.49, found 1017.75, $[\text{M}-^t\text{Bu}+\text{H}]^+$ calcd. 442.20, found 442.35, $[\text{M}-\text{Boc}+\text{H}]^+$ calcd. 398.21, found 398.35. FT-IR ν (cm^{-1}) 2973, 2924, 1688, 1497, 1466, 1419, 1408, 1361, 1317, 1231, 1167, 1135, 1090, 1034.

Dibenzyl 7-(2-methoxy-2-oxoethyl)-1,4,7-triazonane-1,4-dicarboxylate, TACNA (8). Compound **6** (200 mg, 0.402 mmol) was dissolved in 40% TFA solution in DCM (5 ml) and stirred for 1 h. Solvent was removed under vacuum and crude product dissolved in CH_3CN (5 ml). K_2CO_3 (500 mg, 3.618 mmol) was added together with 2-bromoacetate (92 μl , 0.965 mmol) and reaction conducted overnight. At the end of reaction, suspension was filtered on G5 sintered-glass filter to remove KBr and solvent. The crude product was dissolved in water and continuously extracted with chloroform overnight. The extract was purified by preparative HPLC (Phenomenex Jupiter RP C18, 5 μM , 300 \AA , 250 X 15 mm), under isocratic conditions (water- CH_3CN 50:50) affording to pure **8** (84%, 158.8 mg).

(8) $^1\text{H-NMR}$ (400 MHz, 25°C, CDCl_3) $\delta(\text{ppm})$: 7.45 – 7.30 (m, 10H, CH aromatic), 5.2 (M), 5.19 (m), 5.15 (m) (s, 4H, $\underline{\text{CH}_2}$ Cbz), 4.12 (M), 4.03 (m) (s, 2H, $\underline{\text{CH}_2}\text{COOMe}$), 3.79 (s, 3H, OMe), 3.76 – 3.17 (m, 12H, CH cycle). $^{13}\text{C-NMR}$ (100 MHz, 25°C, CDCl_3) $\delta(\text{ppm})$: 166.9, 156.3, 155.8, 136.0, 128.7, 128.5, 68.1, 55.2, 53.2, 52.8, 49.2, 48.5, 46.6, 46.3, 45.4. MS (ESI+) m/z: $[\text{M}+\text{H}]^+ = 471.06$, $[2\text{M}+\text{H}]^+ = 939.67$. FT-IR ν (cm^{-1}) 2953, 1752, 1696, 1674, 1499, 1466, 1423, 1413, 1372, 1298, 1237, 1174, 1155, 1129, 1089. m.p. 152 – 153 °C.

7. References

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8. Appendix

8.1. Standard PNA Force Field

Triplos1: APN

@<TRIPOS>MOLECULE

APN

33 34 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	3.621054	-1.450373	-0.068677	N	1	APN	-0.4030	****
2	H1'1	4.246165	-1.271793	0.682602	H	1	APN	0.2842	****
3	C2'	3.457784	-0.420083	-1.077294	CT	1	APN	-0.1241	****
4	H2'1	3.628216	-0.847867	-2.056986	H1	1	APN	0.1147	****
5	H2'2	4.227242	0.322257	-0.905941	H1	1	APN	0.1147	****
6	C3'	2.080030	0.240647	-1.080497	CT	1	APN	-0.0522	****
7	H3'1	1.323813	-0.470938	-1.363061	H1	1	APN	0.1033	****
8	H3'2	2.071373	1.016457	-1.836440	H1	1	APN	0.1033	****
9	N4'	1.741210	0.867720	0.196197	N	1	APN	-0.0941	****
10	C5'	2.298716	2.197163	0.430342	CT	1	APN	-0.1112	****
11	H5'1	2.391804	2.356515	1.493133	H1	1	APN	0.0678	****
12	H5'2	3.275899	2.250678	-0.024827	H1	1	APN	0.0678	****
13	C'	1.449604	3.289214	-0.217087	C	1	APN	0.6209	****
14	O1'	1.767998	3.770682	-1.273205	O	1	APN	-0.5807	****
15	C7'	0.857680	0.402551	1.099946	C	1	APN	0.4391	****
16	O7'	0.505502	1.058337	2.054036	O	1	APN	-0.5502	****
17	C8'	0.288243	-1.011489	0.977878	CT	1	APN	-0.1214	****
18	H8'1	0.638353	-1.548180	1.850536	H1	1	APN	0.1124	****
19	H8'2	0.593037	-1.557702	0.104279	H1	1	APN	0.1124	****
20	N9	-1.153273	-0.974097	1.005139	N*	1	APN	-0.0578	****
21	C8	-1.981619	-0.898844	2.098267	CK	1	APN	0.1298	****
22	H8	-1.577259	-0.897330	3.089174	H5	1	APN	0.1677	****
23	N7	-3.225847	-0.831369	1.805401	NB	1	APN	-0.5582	****
24	C6	-4.278736	-0.782229	-0.521089	CA	1	APN	0.6597	****
25	N6	-5.576223	-0.734082	-0.171904	N2	1	APN	-0.8883	****
26	H61	-5.821383	-0.567634	0.776793	H	1	APN	0.4048	****
27	H62	-6.246165	-0.541126	-0.880495	H	1	APN	0.4048	****
28	C5	-3.247873	-0.843054	0.423001	CB	1	APN	0.0300	****
29	C4	-1.971958	-0.920512	-0.084086	CB	1	APN	0.4343	****
30	N3	-1.628350	-0.929692	-1.366627	NC	1	APN	-0.7127	****
31	C2	-2.681040	-0.862225	-2.150429	CQ	1	APN	0.5660	****
32	H2	-2.485791	-0.862420	-3.207408	H5	1	APN	0.0560	****
33	N1	-3.961295	-0.789586	-1.810556	NC	1	APN	-0.7398	****

@<TRIPOS>BOND

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6	6	7	1
7	6	8	1
8	6	9	1
9	9	10	1

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10      9      15 1
11     10     11 1
12     10     12 1
13     10     13 1
14     13     14 1
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17     17     18 1
18     17     19 1
19     17     20 1
20     20     21 1
21     20     29 1
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23     21     23 1
24     23     28 1
25     24     25 1
26     24     28 1
27     24     33 1
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33     31     32 1
34     31     33 1

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@<TRIPOS>SUBSTRUCTURE

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1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

C' 1

@<TRIPOS>RESIDUECONNECT

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Triplos2: NAP

@<TRIPOS>MOLECULE

NAP

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

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3	H1'2	-0.372155	0.470271	0.814530	H	1	NAP	0.2531	****
4	H1'3	-0.372159	0.470265	-0.814535	H	1	NAP	0.2531	****
5	C2'	1.451021	0.000000	0.000000	CT	1	NAP	0.0723	****
6	H2'1	1.809090	0.728963	-0.715711	HP	1	NAP	0.0819	****
7	H2'2	1.772427	-0.976835	-0.339099	HP	1	NAP	0.0819	****
8	C3'	2.077421	0.318830	1.356743	CT	1	NAP	-0.0606	****
9	H3'1	1.853667	1.331448	1.644142	H1	1	NAP	0.0894	****
10	H3'2	3.154718	0.255838	1.262557	H1	1	NAP	0.0894	****
11	N4'	1.673355	-0.612895	2.408686	N	1	NAP	-0.0615	****
12	C5'	2.391832	-1.883556	2.456313	CT	1	NAP	-0.1251	****
13	H5'1	1.755750	-2.629775	2.905913	H1	1	NAP	0.0674	****
14	H5'2	2.636268	-2.183962	1.448873	H1	1	NAP	0.0674	****
15	C'	3.712813	-1.758991	3.212768	C	1	NAP	0.6140	****
16	O1'	4.752967	-1.641945	2.618671	O	1	NAP	-0.5754	****
17	C7'	0.814276	-0.353206	3.412835	C	1	NAP	0.4500	****

18 O7'	0.656345	-1.115185	4.339559	O	1 NAP	-0.5598	****
19 C8'	-0.040831	0.914581	3.399618	CT	1 NAP	-0.1055	****
20 H8'1	-1.067893	0.578481	3.333829	H1	1 NAP	0.1129	****
21 H8'2	0.144283	1.587359	2.582595	H1	1 NAP	0.1129	****
22 N9	0.128970	1.645092	4.631539	N*	1 NAP	-0.0808	****
23 C8	-0.484234	1.426978	5.841149	CK	1 NAP	0.1431	****
24 H8	-1.217445	0.654840	5.948895	H5	1 NAP	0.1651	****
25 N7	-0.092748	2.212538	6.772847	NB	1 NAP	-0.5626	****
26 C6	1.677778	4.047283	6.622865	CA	1 NAP	0.6559	****
27 N6	1.615237	4.513686	7.882340	N2	1 NAP	-0.8852	****
28 H61	1.101561	4.007469	8.566237	H	1 NAP	0.4041	****
29 H62	2.320174	5.146225	8.184189	H	1 NAP	0.4041	****
30 C5	0.862351	3.006437	6.165260	CB	1 NAP	0.0362	****
31 C4	1.016266	2.658603	4.843739	CB	1 NAP	0.4357	****
32 N3	1.862590	3.199779	3.975449	NC	1 NAP	-0.7175	****
33 C2	2.573771	4.158240	4.525565	CQ	1 NAP	0.5728	****
34 H2	3.286378	4.646507	3.885945	H5	1 NAP	0.0549	****
35 N1	2.533154	4.605154	5.773949	NC	1 NAP	-0.7416	****

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35	33	34	1
36	33	35	1

@<TRIPOS>SUBSTRUCTURE

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@<TRIPOS>HEADTAIL

0 0

C' 1

@<TRIPOS>RESIDUECONNECT

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Triplos3: CAP

@<TRIPOS>MOLECULE

CAP

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	C'	0.000000	0.000000	0.000000	C	1	CAP	0.7881	****
2	O1'	-0.511689	-1.124427	0.000000	O2	1	CAP	-0.8206	****
3	OXT	-0.540795	1.108412	0.000000	O2	1	CAP	-0.8206	****
4	N1'	4.658960	2.043595	1.148355	N	1	CAP	-0.4302	****
5	H1'1	5.197266	1.232456	0.949976	H	1	CAP	0.2931	****
6	C2'	3.403976	1.870867	1.855917	CT	1	CAP	-0.0573	****
7	H2'1	3.389308	2.519417	2.722526	H1	1	CAP	0.0960	****
8	H2'2	3.373611	0.848591	2.211622	H1	1	CAP	0.0960	****
9	C3'	2.164235	2.172976	1.015351	CT	1	CAP	-0.0479	****
10	H3'1	2.132830	3.216818	0.755607	H1	1	CAP	0.0959	****
11	H3'2	1.284270	1.978060	1.616247	H1	1	CAP	0.0959	****
12	N4'	2.068693	1.343761	-0.185166	N	1	CAP	-0.0212	****
13	C5'	1.527328	0.000000	0.000000	CT	1	CAP	-0.4798	****
14	H5'1	1.915655	-0.644423	-0.772983	H1	1	CAP	0.1621	****
15	H5'2	1.839381	-0.371939	0.963957	H1	1	CAP	0.1621	****
16	C7'	2.293194	1.748333	-1.449955	C	1	CAP	0.4669	****
17	O7'	2.032945	1.052610	-2.405262	O	1	CAP	-0.5706	****
18	C8'	2.935922	3.107040	-1.731879	CT	1	CAP	-0.1309	****
19	H8'1	3.884219	2.892995	-2.208391	H1	1	CAP	0.1143	****
20	H8'2	3.125602	3.715935	-0.867188	H1	1	CAP	0.1143	****
21	N9	2.119337	3.866285	-2.646673	N*	1	CAP	-0.0583	****
22	C8	2.068388	3.776811	-4.016398	CK	1	CAP	0.1330	****
23	H8	2.712150	3.103014	-4.542682	H5	1	CAP	0.1697	****
24	N7	1.204275	4.550897	-4.557254	NB	1	CAP	-0.5627	****
25	C6	-0.402401	6.153321	-3.384917	CA	1	CAP	0.6591	****
26	N6	-1.010141	6.679483	-4.462658	N2	1	CAP	-0.8878	****
27	H61	-0.863299	6.268980	-5.355831	H	1	CAP	0.4046	****
28	H62	-1.820927	7.236423	-4.319130	H	1	CAP	0.4046	****
29	C5	0.614390	5.197601	-3.486992	CB	1	CAP	0.0379	****
30	C4	1.164160	4.775405	-2.299305	CB	1	CAP	0.4242	****
31	N3	0.818087	5.172773	-1.080538	NC	1	CAP	-0.7142	****
32	C2	-0.147642	6.063306	-1.117589	CQ	1	CAP	0.5704	****
33	H2	-0.487004	6.434296	-0.167575	H5	1	CAP	0.0554	****
34	N1	-0.767261	6.567034	-2.176915	NC	1	CAP	-0.7415	****

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0 **** ****

@<TRIPOS>HEADTAIL

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Trip4: MAPN

@<TRIPOS>MOLECULE

MAPN

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

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2	HH31	2.948500	-4.543905	0.795444	HC	1	ACE	0.0639	****
3	HH32	3.800485	-3.308336	1.734877	HC	1	ACE	0.0639	****
4	HH33	4.620396	-4.154872	0.429549	HC	1	ACE	0.0639	****
5	C	3.208694	-2.725793	-0.275020	C	1	ACE	0.6513	****
6	O	2.527394	-3.029687	-1.215905	O	1	ACE	-0.5590	****
7	N1'	3.621054	-1.450373	-0.068677	N	2	APN	-0.5256	****
8	H1'1	4.246165	-1.271793	0.682602	H	2	APN	0.3154	****
9	C2'	3.457784	-0.420082	-1.077294	CT	2	APN	-0.1667	****
10	H2'1	3.628216	-0.847867	-2.056986	H1	2	APN	0.1407	****
11	H2'2	4.227242	0.322257	-0.905941	H1	2	APN	0.1407	****
12	C3'	2.080030	0.240647	-1.080497	CT	2	APN	-0.0601	****
13	H3'1	1.323813	-0.470938	-1.363061	H1	2	APN	0.0992	****
14	H3'2	2.071373	1.016457	-1.836440	H1	2	APN	0.0992	****
15	N4'	1.741210	0.867720	0.196197	N	2	APN	-0.0789	****
16	C5'	2.298716	2.197163	0.430342	CT	2	APN	-0.0497	****
17	H5'1	2.391804	2.356515	1.493133	H1	2	APN	0.0561	****
18	H5'2	3.275899	2.250678	-0.024827	H1	2	APN	0.0561	****
19	C'	1.449604	3.289214	-0.217087	C	2	APN	0.5520	****
20	O1'	1.767998	3.770682	-1.273205	O	2	APN	-0.5667	****
21	C7'	0.857680	0.402551	1.099946	C	2	APN	0.4519	****
22	O7'	0.505502	1.058337	2.054036	O	2	APN	-0.5582	****
23	C8'	0.288243	-1.011489	0.977878	CT	2	APN	-0.1384	****

24	H8'1	0.638353	-1.548180	1.850536	H1	2	APN	0.0996	****
25	H8'2	0.593037	-1.557702	0.104279	H1	2	APN	0.0996	****
26	N9	-1.153273	-0.974097	1.005139	N*	2	APN	-0.0306	****
27	C8	-1.981619	-0.898844	2.098267	CK	2	APN	0.1329	****
28	H8	-1.577259	-0.897330	3.089174	H5	2	APN	0.1676	****
29	N7	-3.225847	-0.831369	1.805401	NB	2	APN	-0.5631	****
30	C6	-4.278736	-0.782229	-0.521089	CA	2	APN	0.6557	****
31	N6	-5.576223	-0.734082	-0.171904	N2	2	APN	-0.8908	****
32	H61	-5.821383	-0.567634	0.776793	H	2	APN	0.4050	****
33	H62	-6.246165	-0.541126	-0.880495	H	2	APN	0.4050	****
34	C5	-3.247873	-0.843054	0.423001	CB	2	APN	0.0465	****
35	C4	-1.971958	-0.920512	-0.084086	CB	2	APN	0.3838	****
36	N3	-1.628350	-0.929692	-1.366627	NC	2	APN	-0.6473	****
37	C2	-2.681040	-0.862225	-2.150429	CQ	2	APN	0.5169	****
38	H2	-2.485791	-0.862420	-3.207408	H5	2	APN	0.0639	****
39	N1	-3.961295	-0.789586	-1.810556	NC	2	APN	-0.7226	****
40	N	0.351653	3.646064	0.468753	N	3	NME	-0.4149	****
41	H	0.105581	3.102507	1.266835	H	3	NME	0.3019	****
42	CH3	-0.608954	4.579103	-0.078842	CT	3	NME	-0.1405	****
43	HH31	-0.104131	5.471506	-0.420612	H1	3	NME	0.0930	****
44	HH32	-1.311496	4.847295	0.698620	H1	3	NME	0.0930	****
45	HH33	-1.153819	4.153269	-0.914556	H1	3	NME	0.0930	****

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Trip5: CPN

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SMALL
USER_CHARGES
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  1 N1'      3.312760  -1.543352   0.212583  N    1 CPN   -0.4030  ****
  2 H1'1     3.702917  -1.507731   1.125329  H    1 CPN    0.2842  ****
  3 C2'      3.409872  -0.355316  -0.616244  CT   1 CPN   -0.1241  ****
  4 H2'1     3.831200  -0.625077  -1.576213  H1   1 CPN    0.1147  ****
  5 H2'2     4.106279   0.317173  -0.130994  H1   1 CPN    0.1147  ****
  6 C3'      2.078572   0.351137  -0.861740  CT   1 CPN   -0.0522  ****
  7 H3'1     1.419465  -0.270223  -1.440286  H1   1 CPN    0.1033  ****
  8 H3'2     2.265962   1.235931  -1.457963  H1   1 CPN    0.1033  ****
  9 N4'      1.418014   0.778575   0.370475  N    1 CPN   -0.0941  ****
 10 C5'      1.868150   2.048826   0.930762  CT   1 CPN   -0.1112  ****
 11 H5'1     1.604383   2.082404   1.976237  H1   1 CPN    0.0678  ****
 12 H5'2     2.941525   2.112645   0.832466  H1   1 CPN    0.0678  ****
 13 C'       1.286287   3.248636   0.184378  C    1 CPN    0.6209  ****
 14 O1'      1.962137   3.876561  -0.588907  O    1 CPN   -0.5807  ****
 15 C7'      0.289974   0.243420   0.877641  C    1 CPN    0.4391  ****
 16 O7'     -0.352871   0.790958   1.744232  O    1 CPN   -0.5502  ****
 17 C8'     -0.168725  -1.146666   0.438193  CT   1 CPN   -0.1769  ****
 18 H8'1     0.067701  -1.813354   1.258285  H1   1 CPN    0.1116  ****
 19 H8'2     0.309278  -1.520521  -0.446642  H1   1 CPN    0.1116  ****
 20 N1      -1.603343  -1.156389   0.242311  N*   1 CPN   -0.0586  ****
 21 C2      -2.082244  -0.489847  -0.899052  C    1 CPN    0.8504  ****
 22 O2      -1.272044   0.028044  -1.621377  O    1 CPN   -0.6164  ****
 23 N3      -3.421586  -0.470877  -1.105266  NC   1 CPN   -0.8062  ****
 24 C4      -4.227530  -0.981286  -0.225184  CA   1 CPN    0.8936  ****
 25 N4      -5.545324  -0.953373  -0.495430  N2   1 CPN   -0.9941  ****
 26 H41     -6.217094  -1.159663   0.205670  H    1 CPN    0.4267  ****
 27 H42     -5.840068  -0.434706  -1.291886  H    1 CPN    0.4267  ****
 28 C5      -3.781194  -1.582483   1.002879  CM   1 CPN   -0.5360  ****
 29 H5      -4.464124  -1.972947   1.730361  HA   1 CPN    0.2051  ****
 30 C6      -2.453831  -1.628734   1.178645  CM   1 CPN   -0.0567  ****
 31 H6      -2.006105  -2.050482   2.057232  H4   1 CPN    0.2189  ****

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@<TRIPOS>SUBSTRUCTURE

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C' 1

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Tripos6: NCP

@<TRIPOS>MOLECULE

NCP

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	0.000000	0.000000	0.000000	N3	1	NCP	0.0049	****
2	H1'1	-0.372154	-0.940539	0.000008	H	1	NCP	0.2531	****
3	H1'2	-0.372155	0.470271	0.814530	H	1	NCP	0.2531	****
4	H1'3	-0.372159	0.470265	-0.814535	H	1	NCP	0.2531	****
5	C2'	1.451831	0.000000	0.000000	CT	1	NCP	0.0723	****
6	H2'1	1.807299	0.707039	-0.738626	HP	1	NCP	0.0819	****
7	H2'2	1.771691	-0.986601	-0.311450	HP	1	NCP	0.0819	****
8	C3'	2.081023	0.358531	1.344350	CT	1	NCP	-0.0606	****
9	H3'1	1.858758	1.376326	1.608771	H1	1	NCP	0.0894	****
10	H3'2	3.157960	0.296916	1.245012	H1	1	NCP	0.0894	****

11	N4'	1.683160	-0.542622	2.424638	N	1	NCP	-0.0615	****
12	C5'	2.432859	-1.790720	2.526052	CT	1	NCP	-0.1251	****
13	H5'1	1.845848	-2.506708	3.079674	H1	1	NCP	0.0674	****
14	H5'2	2.612995	-2.171254	1.531762	H1	1	NCP	0.0674	****
15	C'	3.801844	-1.591844	3.175304	C	1	NCP	0.6140	****
16	O1'	4.802339	-1.580938	2.506033	O	1	NCP	-0.5754	****
17	C7'	0.880256	-0.217213	3.456886	C	1	NCP	0.4500	****
18	O7'	0.790585	-0.904948	4.448353	O	1	NCP	-0.5598	****
19	C8'	-0.037062	1.001789	3.365122	CT	1	NCP	-0.1631	****
20	H8'1	-1.034977	0.614161	3.201317	H1	1	NCP	0.1106	****
21	H8'2	0.194123	1.681153	2.567497	H1	1	NCP	0.1106	****
22	N1	-0.029154	1.723758	4.620227	N*	1	NCP	-0.0668	****
23	C2	1.135831	2.454055	4.913163	C	1	NCP	0.8503	****
24	O2	2.026180	2.428175	4.105080	O	1	NCP	-0.6188	****
25	N3	1.179491	3.131882	6.085923	NC	1	NCP	-0.8037	****
26	C4	0.205489	3.031106	6.937845	CA	1	NCP	0.8897	****
27	N4	0.294463	3.745115	8.074801	N2	1	NCP	-0.9926	****
28	H41	-0.319525	3.575671	8.836168	H	1	NCP	0.4265	****
29	H42	1.153857	4.212176	8.257903	H	1	NCP	0.4265	****
30	C5	-0.957698	2.214143	6.717936	CM	1	NCP	-0.5319	****
31	H5	-1.738204	2.121610	7.446177	HA	1	NCP	0.2039	****
32	C6	-1.007101	1.578182	5.539786	CM	1	NCP	-0.0541	****
33	H6	-1.823841	0.938782	5.267118	H4	1	NCP	0.2174	****

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0 **** ****

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C' 1

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Tripos7:CCP

@<TRIPOS>MOLECULE

CCP

32 32 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	C'	0.000000	0.000000	0.000000	C	1	CCP	0.7881	****
2	O1'	-0.511689	-1.124427	0.000000	O2	1	CCP	-0.8206	****
3	OXT	-0.540795	1.108412	0.000000	O2	1	CCP	-0.8206	****
4	N1'	4.547802	2.222175	1.204150	N	1	CCP	-0.4302	****
5	H1'1	5.114203	1.413832	1.092941	H	1	CCP	0.2931	****
6	C2'	3.247174	2.061524	1.828949	CT	1	CCP	-0.0573	****
7	H2'1	3.150528	2.775088	2.637228	H1	1	CCP	0.0960	****
8	H2'2	3.221350	1.069960	2.263488	H1	1	CCP	0.0960	****
9	C3'	2.065683	2.252761	0.880682	CT	1	CCP	-0.0479	****
10	H3'1	2.019998	3.267760	0.530088	H1	1	CCP	0.0959	****
11	H3'2	1.151130	2.079500	1.434785	H1	1	CCP	0.0959	****
12	N4'	2.080412	1.327413	-0.251091	N	1	CCP	-0.0212	****
13	C5'	1.528135	0.000000	0.000000	CT	1	CCP	-0.4798	****
14	H5'1	1.911977	-0.683800	-0.740811	H1	1	CCP	0.1621	****
15	H5'2	1.838723	-0.327232	0.980981	H1	1	CCP	0.1621	****
16	C7'	2.318781	1.666236	-1.533457	C	1	CCP	0.4669	****
17	O7'	2.067377	0.924611	-2.455853	O	1	CCP	-0.5706	****
18	C8'	3.020906	2.982673	-1.864926	CT	1	CCP	-0.1739	****
19	H8'1	4.034933	2.720339	-2.140338	H1	1	CCP	0.1088	****
20	H8'2	3.064266	3.686705	-1.056442	H1	1	CCP	0.1088	****
21	N1	2.386611	3.609236	-3.005839	N*	1	CCP	-0.0612	****
22	C2	1.123455	4.183709	-2.780559	C	1	CCP	0.8437	****
23	O2	0.672528	4.116382	-1.667670	O	1	CCP	-0.6180	****
24	N3	0.497863	4.768347	-3.831061	NC	1	CCP	-0.8009	****
25	C4	1.021589	4.716284	-5.017487	CA	1	CCP	0.8885	****
26	N4	0.365905	5.334296	-6.016754	N2	1	CCP	-0.9912	****
27	H41	0.614522	5.180692	-6.965420	H	1	CCP	0.4259	****
28	H42	-0.542564	5.689348	-5.819643	H	1	CCP	0.4259	****
29	C5	2.263387	4.050556	-5.306514	CM	1	CCP	-0.5351	****
30	H5	2.665244	3.993955	-6.298176	HA	1	CCP	0.2044	****
31	C6	2.890965	3.508437	-4.254289	CM	1	CCP	-0.0519	****
32	H6	3.821705	2.984115	-4.349449	H4	1	CCP	0.2183	****

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8	6	9	1
9	9	10	1
10	9	11	1
11	9	12	1
12	12	13	1
13	12	16	1
14	13	14	1

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15 13 15 1
16 16 17 1
17 16 18 1
18 18 19 1
19 18 20 1
20 18 21 1
21 21 22 1
22 21 31 1
23 22 23 1
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25 24 25 1
26 25 26 1
27 25 29 1
28 26 27 1
29 26 28 1
30 29 30 1
31 29 31 1
32 31 32 1

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@<TRIPOS>SUBSTRUCTURE

1 CCP

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

0 0

@<TRIPOS>RESIDUECONNECT

1 N1' 0 0 0 0 0

Triplos8: MCPN

@<TRIPOS>MOLECULE

MCPN

43 43 3 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	CH3	3.144181	-3.930966	0.636165	CT	1	ACE	-0.1642	****
2	HH31	2.416220	-4.688501	0.382760	HC	1	ACE	0.0520	****
3	HH32	3.042308	-3.659752	1.681072	HC	1	ACE	0.0520	****
4	HH33	4.134273	-4.350512	0.488026	HC	1	ACE	0.0520	****
5	C	2.969595	-2.750664	-0.296010	C	1	ACE	0.6303	****
6	O	2.562419	-2.880862	-1.418429	O	1	ACE	-0.5637	****
7	N1'	3.312760	-1.543352	0.212583	N	2	CPN	-0.4588	****
8	H1'1	3.702917	-1.507731	1.125329	H	2	CPN	0.2940	****
9	C2'	3.409872	-0.355316	-0.616244	CT	2	CPN	-0.2064	****
10	H2'1	3.831200	-0.625077	-1.576213	H1	2	CPN	0.1369	****
11	H2'2	4.106279	0.317173	-0.130994	H1	2	CPN	0.1369	****
12	C3'	2.078572	0.351137	-0.861740	CT	2	CPN	-0.0645	****
13	H3'1	1.419465	-0.270223	-1.440286	H1	2	CPN	0.1270	****
14	H3'2	2.265962	1.235931	-1.457963	H1	2	CPN	0.1270	****
15	N4'	1.418014	0.778575	0.370475	N	2	CPN	-0.0696	****
16	C5'	1.868150	2.048826	0.930762	CT	2	CPN	-0.1121	****
17	H5'1	1.604383	2.082404	1.976237	H1	2	CPN	0.0694	****
18	H5'2	2.941525	2.112645	0.832466	H1	2	CPN	0.0694	****
19	C'	1.286287	3.248636	0.184378	C	2	CPN	0.5820	****
20	O1'	1.962137	3.876561	-0.588907	O	2	CPN	-0.5721	****
21	C7'	0.289974	0.243420	0.877641	C	2	CPN	0.4343	****
22	O7'	-0.352871	0.790958	1.744232	O	2	CPN	-0.5623	****
23	C8'	-0.168725	-1.146666	0.438193	CT	2	CPN	-0.0643	****
24	H8'1	0.067701	-1.813354	1.258285	H1	2	CPN	0.0679	****

25	H8'2	0.309278	-1.520521	-0.446642	H1	2	CPN	0.0679	****
26	N1	-1.603343	-1.156389	0.242311	N*	2	CPN	-0.0882	****
27	C2	-2.082244	-0.489847	-0.899052	C	2	CPN	0.8972	****
28	O2	-1.272044	0.028044	-1.621377	O	2	CPN	-0.6563	****
29	N3	-3.421586	-0.470877	-1.105266	NC	2	CPN	-0.7991	****
30	C4	-4.227530	-0.981286	-0.225184	CA	2	CPN	0.8624	****
31	N4	-5.545324	-0.953373	-0.495430	N2	2	CPN	-0.9823	****
32	H41	-6.217094	-1.159663	0.205670	H	2	CPN	0.4253	****
33	H42	-5.840068	-0.434706	-1.291886	H	2	CPN	0.4253	****
34	C5	-3.781194	-1.582483	1.002879	CM	2	CPN	-0.5186	****
35	H5	-4.464124	-1.972947	1.730361	HA	2	CPN	0.2026	****
36	C6	-2.453831	-1.628734	1.178645	CM	2	CPN	-0.0638	****
37	H6	-2.006105	-2.050482	2.057232	H4	2	CPN	0.2201	****
38	N	0.001889	3.528780	0.459686	N	3	NME	-0.4595	****
39	H	-0.505206	2.868136	1.006995	H	3	NME	0.3202	****
40	CH3	-0.729953	4.522394	-0.295657	CT	3	NME	-0.0875	****
41	HH31	-0.147181	5.429231	-0.367286	H1	3	NME	0.0804	****
42	HH32	-1.655365	4.738129	0.221813	H1	3	NME	0.0804	****
43	HH33	-0.959308	4.177959	-1.298297	H1	3	NME	0.0804	****

@<TRIPOS>BOND

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@<TRIPPOS>SUBSTRUCTURE
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    2 CPN          7 ****          0 ****  ****
    3 NME          38 ****         0 ****  ****
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0 0
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2 N1' C' 0 0 0 0
3 N 0 0 0 0 0

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Trip9: GPN

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@<TRIPPOS>MOLECULE
GPN
    34      35      1      0      1
SMALL
USER_CHARGES
@<TRIPPOS>ATOM
  1 N1'      4.122369   -0.011061   -0.354039 N      1 GPN      -0.4030 ****
  2 H1'1     4.685538    0.387828    0.360264 H      1 GPN       0.2842 ****
  3 C2'      3.375242    0.883187   -1.218440 CT     1 GPN      -0.1241 ****
  4 H2'1     3.602902    0.654488   -2.252239 H1     1 GPN       0.1147 ****
  5 H2'2     3.728294    1.888000   -1.020195 H1     1 GPN       0.1147 ****
  6 C3'      1.861091    0.807184   -1.034211 CT     1 GPN      -0.0522 ****
  7 H3'1     1.508956   -0.169250   -1.308675 H1     1 GPN       0.1033 ****
  8 H3'2     1.385586    1.503556   -1.712045 H1     1 GPN       0.1033 ****
  9 N4'      1.442012    1.143452    0.322917 N      1 GPN      -0.0941 ****
 10 C5'      1.274774    2.560901    0.611615 CT     1 GPN      -0.1112 ****
 11 H5'1     1.507660    2.748575    1.647866 H1     1 GPN       0.0678 ****
 12 H5'2     1.958337    3.123362   -0.007239 H1     1 GPN       0.0678 ****
 13 C'       -0.136885    3.034278    0.262312 C      1 GPN       0.6209 ****
 14 O1'      -0.436875    3.291728   -0.881342 O      1 GPN      -0.5807 ****
 15 C7'      0.908973    0.291353    1.227567 C      1 GPN       0.4391 ****
 16 O7'      0.434614    0.681319    2.265912 O      1 GPN      -0.5502 ****
 17 C8'      0.930894   -1.218898    0.973741 CT     1 GPN      -0.2359 ****
 18 H8'1     1.388371   -1.656028    1.850521 H1     1 GPN       0.1418 ****
 19 H8'2     1.499243   -1.518921    0.110264 H1     1 GPN       0.1418 ****
 20 N9       -0.419969   -1.732213    0.885575 N*     1 GPN       0.0268 ****
 21 C8       -0.938170   -2.847500    1.505896 CK     1 GPN       0.0862 ****
 22 H8       -0.350748   -3.413771    2.199667 H5     1 GPN       0.1623 ****
 23 N7       -2.143275   -3.103753    1.168112 NB     1 GPN      -0.5377 ****
 24 C6       -3.680934   -1.852372   -0.458440 C      1 GPN       0.4075 ****
 25 O6       -4.720758   -2.438010   -0.491249 O      1 GPN      -0.5295 ****
 26 C5       -2.470564   -2.111875    0.269409 CB     1 GPN       0.2319 ****
 27 C4       -1.413271   -1.264990    0.091765 CB     1 GPN       0.0882 ****
 28 N3       -1.328232   -0.155400   -0.676135 NC     1 GPN      -0.3651 ****
 29 C2       -2.418087    0.101743   -1.321498 CA     1 GPN       0.4620 ****
 30 N2       -2.476606    1.182902   -2.135232 N2     1 GPN      -0.8775 ****
 31 H21      -1.767274    1.870625   -1.969915 H      1 GPN       0.4042 ****
 32 H22      -3.371292    1.551625   -2.367077 H      1 GPN       0.4042 ****
 33 N1       -3.526182   -0.682293   -1.245267 NA     1 GPN      -0.3251 ****
 34 H1       -4.323926   -0.465858   -1.804032 H      1 GPN       0.3136 ****
@<TRIPPOS>BOND
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  2      1      3 1

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33     30    31 1
34     30    32 1
35     33    34 1

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@<TRIPOS>SUBSTRUCTURE

1 GPN

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

C' 1

@<TRIPOS>RESIDUECONNECT

1 N1' C' 0 0 0 0

Triplos10: NGP

@<TRIPOS>MOLECULE

NGP

36 37 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	0.000000	0.000000	0.000000	N3	1	NGP	0.0049	****
2	H1'1	-0.372154	-0.940539	0.000008	H	1	NGP	0.2531	****
3	H1'2	-0.372155	0.470271	0.814530	H	1	NGP	0.2531	****
4	H1'3	-0.372159	0.470265	-0.814535	H	1	NGP	0.2531	****
5	C2'	1.450885	0.000000	0.000000	CT	1	NGP	0.0723	****
6	H2'1	1.808606	0.667951	-0.773789	HP	1	NGP	0.0819	****
7	H2'2	1.770286	-1.002133	-0.259438	HP	1	NGP	0.0819	****
8	C3'	2.073987	0.428275	1.326911	CT	1	NGP	-0.0606	****
9	H3'1	1.817015	1.449196	1.537716	H1	1	NGP	0.0894	****

10	H3'2	3.151890	0.397925	1.239142	H1	1	NGP	0.0894	****
11	N4'	1.688505	-0.439558	2.435410	N	1	NGP	-0.0615	****
12	C5'	2.476264	-1.652240	2.606625	CT	1	NGP	-0.1251	****
13	H5'1	1.854640	-2.431969	3.017574	H1	1	NGP	0.0674	****
14	H5'2	2.839635	-1.968519	1.639914	H1	1	NGP	0.0674	****
15	C'	3.703062	-1.396381	3.483199	C	1	NGP	0.6140	****
16	O1'	4.697580	-0.885744	3.020161	O	1	NGP	-0.5754	****
17	C7'	0.898835	-0.098127	3.478694	C	1	NGP	0.4500	****
18	O7'	0.764838	-0.818272	4.437175	O	1	NGP	-0.5598	****
19	C8'	0.107932	1.213202	3.453096	CT	1	NGP	-0.2061	****
20	H8'1	-0.919431	0.938764	3.648851	H1	1	NGP	0.1347	****
21	H8'2	0.144783	1.739285	2.514744	H1	1	NGP	0.1347	****
22	N9	0.539700	2.080286	4.529125	N*	1	NGP	0.0149	****
23	C8	-0.250429	2.776796	5.416686	CK	1	NGP	0.0908	****
24	H8	-1.315270	2.662171	5.412165	H5	1	NGP	0.1618	****
25	N7	0.413437	3.532922	6.203830	NB	1	NGP	-0.5404	****
26	C6	2.945591	3.910922	6.349885	C	1	NGP	0.4042	****
27	O6	3.139635	4.708326	7.216936	O	1	NGP	-0.5290	****
28	C5	1.728738	3.345237	5.838641	CB	1	NGP	0.2410	****
29	C4	1.812100	2.452105	4.808104	CB	1	NGP	0.0795	****
30	N3	2.909697	1.973985	4.179794	NC	1	NGP	-0.3622	****
31	C2	4.013894	2.459569	4.643861	CA	1	NGP	0.4564	****
32	N2	5.195199	2.072913	4.105801	N2	1	NGP	-0.8711	****
33	H21	5.155316	1.226248	3.571936	H	1	NGP	0.4024	****
34	H22	6.021302	2.187023	4.648668	H	1	NGP	0.4024	****
35	N1	4.055848	3.374183	5.648907	NA	1	NGP	-0.3229	****
36	H1	4.932940	3.754317	5.934680	H	1	NGP	0.3134	****

@<TRIPOS>BOND

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34	31	35	1

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35      32      33 1
36      32      34 1
37      35      36 1
@<TRIPOS>SUBSTRUCTURE
1 NGP          1 ****          0 ****  ****
@<TRIPOS>HEADTAIL
0 0
C' 1
@<TRIPOS>RESIDUECONNECT
1 0 C' 0 0 0 0

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Triplos11: CGP

```

@<TRIPOS>MOLECULE
CGP
35      36      1      0      1
SMALL
USER_CHARGES
@<TRIPOS>ATOM
1 C'      0.000000      0.000000      0.000000 C      1 CGP      0.7881 ****
2 O1'     -0.511689     -1.124427      0.000000 O2     1 CGP     -0.8206 ****
3 OXT     -0.540795      1.108412      0.000000 O2     1 CGP     -0.8206 ****
4 N1'      4.733360      2.037702      1.112789 N      1 CGP     -0.4302 ****
5 H1'1     5.292873      1.307794      0.737716 H      1 CGP      0.2931 ****
6 C2'      3.569496      1.674956      1.899493 CT     1 CGP     -0.0573 ****
7 H2'1     3.614306      2.178052      2.857491 H1     1 CGP      0.0960 ****
8 H2'2     3.629641      0.609550      2.086275 H1     1 CGP      0.0960 ****
9 C3'      2.237460      2.023037      1.238516 CT     1 CGP     -0.0479 ****
10 H3'1    2.151970      3.087675      1.129016 H1     1 CGP      0.0959 ****
11 H3'2    1.428178      1.718262      1.888649 H1     1 CGP      0.0959 ****
12 N4'      2.056513      1.356583     -0.047407 N      1 CGP     -0.0212 ****
13 C5'      1.529340      0.000000      0.000000 CT     1 CGP     -0.4798 ****
14 H5'1     1.922895     -0.569746     -0.826908 H1     1 CGP      0.1621 ****
15 H5'2     1.844858     -0.462054      0.923881 H1     1 CGP      0.1621 ****
16 C7'      2.034865      1.954718     -1.259991 C      1 CGP      0.4669 ****
17 O7'      1.713460      1.357833     -2.257827 O      1 CGP     -0.5706 ****
18 C8'      2.464593      3.417266     -1.408498 CT     1 CGP     -0.2258 ****
19 H8'1     3.222431      3.422366     -2.179709 H1     1 CGP      0.1344 ****
20 H8'2     2.884856      3.852372     -0.518167 H1     1 CGP      0.1344 ****
21 N9       1.356427      4.219949     -1.881533 N*     1 CGP      0.0345 ****
22 C8       1.364998      5.150565     -2.896952 CK     1 CGP      0.0831 ****
23 H8       2.240954      5.305776     -3.493320 H5     1 CGP      0.1624 ****
24 N7       0.254792      5.768411     -3.030183 NB     1 CGP     -0.5375 ****
25 C6      -1.923394      5.510352     -1.701674 C      1 CGP      0.4092 ****
26 O6      -2.709425      6.290962     -2.146741 O      1 CGP     -0.5305 ****
27 C5      -0.559594      5.233458     -2.055910 CB     1 CGP      0.2325 ****
28 C4       0.113631      4.279918     -1.345862 CB     1 CGP      0.0799 ****
29 N3      -0.326715      3.500988     -0.332227 NC     1 CGP     -0.3583 ****
30 C2      -1.559702      3.725474     -0.016398 CA     1 CGP      0.4443 ****
31 N2      -2.134228      3.020094      0.987022 N2     1 CGP     -0.8613 ****
32 H21     -1.654590      2.176015      1.233849 H      1 CGP      0.3992 ****
33 H22     -3.127153      2.960676      1.014221 H      1 CGP      0.3992 ****
34 N1      -2.322437      4.666699     -0.633431 NA     1 CGP     -0.3215 ****
35 H1      -3.253411      4.833994     -0.316008 H      1 CGP      0.3139 ****
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3      1     13 1

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35    31    33 1
36    34    35 1

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@<TRIPOS>SUBSTRUCTURE

1 CGP

1 ****

0 **** ****

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Triplos12: MGPN

@<TRIPOS>MOLECULE

MGPN

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

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2	HH31	4.956259	-3.121745	0.274719	HC	1	ACE	0.0719	****
3	HH32	5.319264	-1.679633	1.235446	HC	1	ACE	0.0719	****
4	HH33	6.233092	-2.026775	-0.226492	HC	1	ACE	0.0719	****
5	C	4.263906	-1.328219	-0.632833	C	1	ACE	0.6551	****
6	O	3.650754	-1.861271	-1.518940	O	1	ACE	-0.5668	****
7	N1'	4.122369	-0.011061	-0.354039	N	2	GPN	-0.5393	****
8	H1'1	4.685538	0.387828	0.360264	H	2	GPN	0.3199	****
9	C2'	3.375242	0.883187	-1.218440	CT	2	GPN	-0.1343	****

10	H2'1	3.602902	0.654488	-2.252239	H1	2	GPN	0.1313	****
11	H2'2	3.728294	1.888000	-1.020195	H1	2	GPN	0.1313	****
12	C3'	1.861091	0.807184	-1.034211	CT	2	GPN	-0.0753	****
13	H3'1	1.508956	-0.169250	-1.308675	H1	2	GPN	0.0928	****
14	H3'2	1.385586	1.503556	-1.712045	H1	2	GPN	0.0928	****
15	N4'	1.442012	1.143452	0.322917	N	2	GPN	-0.0660	****
16	C5'	1.274774	2.560901	0.611615	CT	2	GPN	-0.0463	****
17	H5'1	1.507660	2.748575	1.647866	H1	2	GPN	0.0638	****
18	H5'2	1.958337	3.123362	-0.007239	H1	2	GPN	0.0638	****
19	C'	-0.136885	3.034278	0.262312	C	2	GPN	0.5106	****
20	O1'	-0.436875	3.291728	-0.881342	O	2	GPN	-0.5496	****
21	C7'	0.908973	0.291353	1.227567	C	2	GPN	0.3502	****
22	O7'	0.434614	0.681319	2.265912	O	2	GPN	-0.5188	****
23	C8'	0.930894	-1.218898	0.973741	CT	2	GPN	-0.1247	****
24	H8'1	1.388371	-1.656028	1.850521	H1	2	GPN	0.1220	****
25	H8'2	1.499243	-1.518921	0.110264	H1	2	GPN	0.1220	****
26	N9	-0.419969	-1.732213	0.885575	N*	2	GPN	-0.0216	****
27	C8	-0.938170	-2.847500	1.505896	CK	2	GPN	0.0950	****
28	H8	-0.350748	-3.413771	2.199667	H5	2	GPN	0.1612	****
29	N7	-2.143275	-3.103753	1.168112	NB	2	GPN	-0.5400	****
30	C6	-3.680934	-1.852372	-0.458440	C	2	GPN	0.3790	****
31	O6	-4.720758	-2.438010	-0.491249	O	2	GPN	-0.5262	****
32	C5	-2.470564	-2.111875	0.269409	CB	2	GPN	0.2702	****
33	C4	-1.413271	-1.264990	0.091765	CB	2	GPN	0.0213	****
34	N3	-1.328232	-0.155400	-0.676135	NC	2	GPN	-0.2338	****
35	C2	-2.418087	0.101743	-1.321498	CA	2	GPN	0.3562	****
36	N2	-2.476606	1.182902	-2.135232	N2	2	GPN	-0.8521	****
37	H21	-1.767274	1.870625	-1.969915	H	2	GPN	0.4017	****
38	H22	-3.371292	1.551625	-2.367077	H	2	GPN	0.4017	****
39	N1	-3.526182	-0.682293	-1.245267	NA	2	GPN	-0.2798	****
40	H1	-4.323926	-0.465858	-1.804032	H	2	GPN	0.3090	****
41	N	-0.974453	3.159616	1.301021	N	3	NME	-0.4710	****
42	H	-0.720015	2.669624	2.131110	H	3	NME	0.3430	****
43	CH3	-2.367489	3.503665	1.118477	CT	3	NME	-0.0758	****
44	HH31	-2.447842	4.310199	0.405014	H1	3	NME	0.0796	****
45	HH32	-2.772615	3.831079	2.066429	H1	3	NME	0.0796	****
46	HH33	-2.951120	2.661155	0.762137	H1	3	NME	0.0796	****

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Triplos13: IPN

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IPN

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SMALL

USER_CHARGES

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1 N1' 3.559657 -1.542274 -0.052240 N 1 IPN -0.4030 ****
2 H1'1 4.203196 -1.390317 0.689405 H 1 IPN 0.2842 ****
3 C2' 3.435242 -0.512090 -1.066643 CT 1 IPN -0.1241 ****
4 H2'1 3.600955 -0.948354 -2.043642 H1 1 IPN 0.1147 ****
5 H2'2 4.224092 0.208605 -0.891637 H1 1 IPN 0.1147 ****
6 C3' 2.077227 0.187561 -1.082969 CT 1 IPN -0.0522 ****
7 H3'1 1.302946 -0.511790 -1.346386 H1 1 IPN 0.1033 ****
8 H3'2 2.088007 0.945093 -1.857007 H1 1 IPN 0.1033 ****
9 N4' 1.758896 0.854591 0.178000 N 1 IPN -0.0941 ****
10 C5' 2.344620 2.178591 0.373362 CT 1 IPN -0.1112 ****
11 H5'1 2.460129 2.359702 1.430258 H1 1 IPN 0.0678 ****
12 H5'2 3.314042 2.204119 -0.100560 H1 1 IPN 0.0678 ****
13 C' 1.499276 3.265679 -0.286995 C 1 IPN 0.6209 ****
14 O1' 1.779240 3.680739 -1.381926 O 1 IPN -0.5807 ****
15 C7' 0.874076 0.433347 1.102883 C 1 IPN 0.4391 ****

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16	O7'	0.553995	1.118469	2.046300	O	1	IPN	-0.5502	****
17	C8'	0.262332	-0.965693	1.015962	CT	1	IPN	-0.0806	****
18	H8'1	0.605179	-1.494691	1.895814	H1	1	IPN	0.0966	****
19	H8'2	0.541523	-1.538143	0.151025	H1	1	IPN	0.0966	****
20	N9	-1.179020	-0.886381	1.060590	N*	1	IPN	-0.0288	****
21	C8	-1.985563	-0.743800	2.160091	CK	1	IPN	0.0720	****
22	H8	-1.569220	-0.695999	3.144244	H5	1	IPN	0.1798	****
23	N7	-3.232071	-0.672045	1.871360	NB	1	IPN	-0.5189	****
24	C6	-4.397681	-0.731078	-0.404605	C	1	IPN	0.5198	****
25	O6	-5.568101	-0.634105	-0.193600	O	1	IPN	-0.5429	****
26	C5	-3.276262	-0.755586	0.501537	CB	1	IPN	0.1278	****
27	C4	-2.015744	-0.881579	-0.008061	CB	1	IPN	0.3258	****
28	N3	-1.636599	-0.981220	-1.311904	NC	1	IPN	-0.5586	****
29	C2	-2.632626	-0.957938	-2.110315	CQ	1	IPN	0.2453	****
30	H2	-2.463275	-1.030037	-3.168844	H5	1	IPN	0.1476	****
31	N1	-3.926211	-0.845529	-1.730193	NA	1	IPN	-0.4020	****
32	H1	-4.639593	-0.837005	-2.428008	H	1	IPN	0.3202	****

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Tripos14: NIP

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SMALL

USER_CHARGES

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2	H1'1	-0.372154	-0.940539	0.000008	H	1	NIP	0.2531	****
3	H1'2	-0.372155	0.470271	0.814530	H	1	NIP	0.2531	****
4	H1'3	-0.372159	0.470265	-0.814535	H	1	NIP	0.2531	****
5	C2'	1.451128	0.000000	0.000000	CT	1	NIP	0.0723	****
6	H2'1	1.810174	0.717285	-0.727257	HP	1	NIP	0.0819	****
7	H2'2	1.772793	-0.982310	-0.322352	HP	1	NIP	0.0819	****
8	C3'	2.075668	0.341031	1.351899	CT	1	NIP	-0.0606	****
9	H3'1	1.829710	1.351129	1.629745	H1	1	NIP	0.0894	****
10	H3'2	3.153618	0.299269	1.255016	H1	1	NIP	0.0894	****
11	N4'	1.695025	-0.590007	2.412363	N	1	NIP	-0.0615	****
12	C5'	2.448174	-1.840266	2.474299	CT	1	NIP	-0.1251	****
13	H5'1	1.828027	-2.602908	2.918110	H1	1	NIP	0.0674	****
14	H5'2	2.714474	-2.136679	1.471176	H1	1	NIP	0.0674	****
15	C'	3.754015	-1.667339	3.247160	C	1	NIP	0.6140	****
16	O1'	4.790076	-1.471621	2.665955	O	1	NIP	-0.5754	****
17	C7'	0.825303	-0.348270	3.412817	C	1	NIP	0.4500	****
18	O7'	0.679636	-1.113192	4.337779	O	1	NIP	-0.5598	****
19	C8'	-0.054693	0.902491	3.395346	CT	1	NIP	-0.0662	****
20	H8'1	-1.074689	0.546787	3.327489	H1	1	NIP	0.0979	****
21	H8'2	0.119606	1.577580	2.578255	H1	1	NIP	0.0979	****
22	N9	0.093992	1.637487	4.629627	N*	1	NIP	-0.0507	****
23	C8	-0.504236	1.392065	5.838607	CK	1	NIP	0.0847	****
24	H8	-1.193964	0.582940	5.955980	H5	1	NIP	0.1775	****
25	N7	-0.144589	2.207433	6.759544	NB	1	NIP	-0.5241	****
26	C6	1.504439	4.160721	6.672818	C	1	NIP	0.5174	****
27	O6	1.526128	4.640554	7.765108	O	1	NIP	-0.5423	****
28	C5	0.757460	3.043819	6.149533	CB	1	NIP	0.1370	****
29	C4	0.916174	2.696999	4.838428	CB	1	NIP	0.3238	****
30	N3	1.724375	3.269196	3.904037	NC	1	NIP	-0.5633	****
31	C2	2.384424	4.256723	4.372222	CQ	1	NIP	0.2549	****
32	H2	3.058679	4.793525	3.730672	H5	1	NIP	0.1456	****
33	N1	2.309411	4.702297	5.647497	NA	1	NIP	-0.4068	****
34	H1	2.864429	5.484515	5.923219	H	1	NIP	0.3212	****

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Trip15: CIP

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CIP

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SMALL

USER_CHARGES

@<TRIPPOS>ATOM

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2	O1'	-0.511689	-1.124427	0.000000	O2	1	CIP	-0.8206	****
3	OXT	-0.540795	1.108412	0.000000	O2	1	CIP	-0.8206	****
4	N1'	4.664268	2.092576	1.132439	N	1	CIP	-0.4302	****
5	H1'1	5.232991	1.304479	0.925731	H	1	CIP	0.2931	****
6	C2'	3.423499	1.866503	1.850182	CT	1	CIP	-0.0573	****
7	H2'1	3.403315	2.487593	2.736835	H1	1	CIP	0.0960	****
8	H2'2	3.422817	0.833296	2.173904	H1	1	CIP	0.0960	****
9	C3'	2.166747	2.163102	1.033736	CT	1	CIP	-0.0479	****
10	H3'1	2.122073	3.210049	0.788997	H1	1	CIP	0.0959	****
11	H3'2	1.298816	1.949844	1.645554	H1	1	CIP	0.0959	****
12	N4'	2.060980	1.348561	-0.175248	N	1	CIP	-0.0212	****
13	C5'	1.527232	0.000000	0.000000	CT	1	CIP	-0.4798	****
14	H5'1	1.919242	-0.637197	-0.776840	H1	1	CIP	0.1621	****
15	H5'2	1.840731	-0.376559	0.961746	H1	1	CIP	0.1621	****
16	C7'	2.270971	1.765056	-1.439446	C	1	CIP	0.4669	****
17	O7'	2.014053	1.072097	-2.396358	O	1	CIP	-0.5706	****
18	C8'	2.890620	3.135760	-1.715681	CT	1	CIP	-0.0900	****
19	H8'1	3.837370	2.940752	-2.202670	H1	1	CIP	0.0987	****
20	H8'2	3.078639	3.740180	-0.847875	H1	1	CIP	0.0987	****
21	N9	2.055654	3.890301	-2.620820	N*	1	CIP	-0.0296	****
22	C8	1.983143	3.792577	-3.986447	CK	1	CIP	0.0759	****
23	H8	2.605105	3.106501	-4.521840	H5	1	CIP	0.1819	****
24	N7	1.117264	4.580279	-4.508057	NB	1	CIP	-0.5250	****
25	C6	-0.469997	6.247354	-3.392885	C	1	CIP	0.5174	****

26	O6	-1.095631	6.763843	-4.267890	O	1	CIP	-0.5427	****
27	C5	0.559973	5.239285	-3.440122	CB	1	CIP	0.1399	****
28	C4	1.127025	4.816678	-2.272079	CB	1	CIP	0.3135	****
29	N3	0.844046	5.215824	-1.001531	NC	1	CIP	-0.5606	****
30	C2	-0.069064	6.107422	-0.964717	CQ	1	CIP	0.2509	****
31	H2	-0.381701	6.504926	-0.016766	H5	1	CIP	0.1466	****
32	N1	-0.700734	6.612927	-2.049128	NA	1	CIP	-0.4040	****
33	H1	-1.403395	7.310109	-1.922096	H	1	CIP	0.3205	****

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MIPN

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SMALL

USER_CHARGES

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2	HH31	2.760822	-4.601552	0.826535	HC	1	ACE	0.0719	****
3	HH32	3.677193	-3.403266	1.753367	HC	1	ACE	0.0719	****
4	HH33	4.444325	-4.288559	0.441684	HC	1	ACE	0.0719	****
5	C	3.091040	-2.799280	-0.251333	C	1	ACE	0.6711	****
6	O	2.387473	-3.074039	-1.185429	O	1	ACE	-0.5676	****
7	N1'	3.559657	-1.542274	-0.052240	N	2	IPN	-0.5493	****
8	H1'1	4.203196	-1.390317	0.689405	H	2	IPN	0.3236	****
9	C2'	3.435242	-0.512090	-1.066643	CT	2	IPN	-0.1739	****
10	H2'1	3.600955	-0.948354	-2.043642	H1	2	IPN	0.1477	****
11	H2'2	4.224092	0.208605	-0.891637	H1	2	IPN	0.1477	****
12	C3'	2.077227	0.187561	-1.082969	CT	2	IPN	-0.0463	****
13	H3'1	1.302946	-0.511790	-1.346386	H1	2	IPN	0.0834	****
14	H3'2	2.088007	0.945093	-1.857007	H1	2	IPN	0.0834	****
15	N4'	1.758896	0.854591	0.178000	N	2	IPN	-0.0811	****
16	C5'	2.344620	2.178591	0.373362	CT	2	IPN	-0.0374	****
17	H5'1	2.460129	2.359702	1.430258	H1	2	IPN	0.0523	****
18	H5'2	3.314042	2.204119	-0.100560	H1	2	IPN	0.0523	****
19	C'	1.499276	3.265679	-0.286995	C	2	IPN	0.5520	****
20	O1'	1.779240	3.680739	-1.381926	O	2	IPN	-0.5680	****
21	C7'	0.874076	0.433347	1.102883	C	2	IPN	0.4496	****
22	O7'	0.553995	1.118469	2.046300	O	2	IPN	-0.5590	****
23	C8'	0.262332	-0.965693	1.015962	CT	2	IPN	-0.0753	****
24	H8'1	0.605179	-1.494691	1.895814	H1	2	IPN	0.0810	****
25	H8'2	0.541523	-1.538143	0.151025	H1	2	IPN	0.0810	****
26	N9	-1.179020	-0.886381	1.060590	N*	2	IPN	-0.0092	****
27	C8	-1.985563	-0.743800	2.160091	CK	2	IPN	0.0814	****
28	H8	-1.569220	-0.695999	3.144244	H5	2	IPN	0.1778	****
29	N7	-3.232071	-0.672045	1.871360	NB	2	IPN	-0.5319	****
30	C6	-4.397681	-0.731078	-0.404605	C	2	IPN	0.4676	****
31	O6	-5.568101	-0.634105	-0.193600	O	2	IPN	-0.5361	****
32	C5	-3.276262	-0.755586	0.501537	CB	2	IPN	0.1935	****
33	C4	-2.015744	-0.881579	-0.008061	CB	2	IPN	0.2153	****
34	N3	-1.636599	-0.981220	-1.311904	NC	2	IPN	-0.4249	****
35	C2	-2.632626	-0.957938	-2.110315	CQ	2	IPN	0.1272	****
36	H2	-2.463275	-1.030037	-3.168844	H5	2	IPN	0.1677	****
37	N1	-3.926211	-0.845529	-1.730193	NA	2	IPN	-0.3258	****
38	H1	-4.639593	-0.837005	-2.428008	H	2	IPN	0.3083	****
39	N	0.448069	3.692648	0.430458	N	3	NME	-0.4086	****
40	H	0.218097	3.191295	1.260374	H	3	NME	0.3051	****
41	CH3	-0.506205	4.633386	-0.115671	CT	3	NME	-0.1585	****
42	HH31	0.011491	5.490432	-0.522330	H1	3	NME	0.0978	****
43	HH32	-1.161562	4.961288	0.679602	H1	3	NME	0.0978	****
44	HH33	-1.104509	4.189635	-0.904257	H1	3	NME	0.0978	****

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Trip17: TPN

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TPN

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SMALL

USER_CHARGES

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1 N1' -3.164169 -2.076590 -0.366700 N 1 TPN -0.4030 ****
2 H1'1 -3.506187 -2.157192 -1.296066 H 1 TPN 0.2842 ****
3 C2' -3.574308 -0.918849 0.407734 CT 1 TPN -0.1241 ****
4 H2'1 -4.043432 -1.248639 1.326301 H1 1 TPN 0.1147 ****
5 H2'2 -4.323694 -0.395224 -0.172374 H1 1 TPN 0.1147 ****
6 C3' -2.434601 0.029400 0.773185 CT 1 TPN -0.0522 ****
7 H3'1 -1.736380 -0.458108 1.429867 H1 1 TPN 0.1033 ****
8 H3'2 -2.847904 0.863153 1.327406 H1 1 TPN 0.1033 ****
9 N4' -1.739615 0.576891 -0.391242 N 1 TPN -0.0941 ****
10 C5' -2.365484 1.731600 -1.029253 CT 1 TPN -0.1112 ****

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11	H5'1	-2.030029	1.789884	-2.052832	H1	1	TPN	0.0678	****
12	H5'2	-3.436772	1.599072	-1.013366	H1	1	TPN	0.0678	****
13	C'	-2.071035	3.030494	-0.280901	C	1	TPN	0.6209	****
14	O1'	-2.885875	3.506896	0.465867	O	1	TPN	-0.5807	****
15	C7'	-0.493775	0.252814	-0.788018	C	1	TPN	0.4391	****
16	O7'	0.116709	0.906616	-1.602495	O	1	TPN	-0.5502	****
17	C8'	0.167399	-1.027414	-0.277164	CT	1	TPN	-0.0664	****
18	H8'1	0.112897	-1.739168	-1.090636	H1	1	TPN	0.0917	****
19	H8'2	-0.302938	-1.469186	0.579881	H1	1	TPN	0.0917	****
20	N1	1.565822	-0.792717	0.023138	N*	1	TPN	-0.0415	****
21	C2	1.842742	-0.187207	1.221262	C	1	TPN	0.6366	****
22	O2	0.985969	0.112374	2.004874	O	1	TPN	-0.5738	****
23	N3	3.169727	0.044652	1.449507	NA	1	TPN	-0.5240	****
24	H3	3.389568	0.465526	2.327773	H	1	TPN	0.3452	****
25	C4	4.231392	-0.185367	0.586981	C	1	TPN	0.6300	****
26	O4	5.349034	0.081073	0.916386	O	1	TPN	-0.5603	****
27	C5	3.845209	-0.751534	-0.709285	CM	1	TPN	0.0069	****
28	C6	2.555494	-1.002612	-0.917275	CM	1	TPN	-0.2988	****
29	H6	2.203698	-1.400670	-1.848115	H4	1	TPN	0.2494	****
30	C7	4.928309	-0.997851	-1.721763	CT	1	TPN	-0.2318	****
31	H71	5.445785	-0.076709	-1.964474	HC	1	TPN	0.0816	****
32	H72	5.668103	-1.689340	-1.334509	HC	1	TPN	0.0816	****
33	H73	4.515420	-1.410559	-2.635231	HC	1	TPN	0.0816	****

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Tripo18: NTP

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NTP

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SMALL

USER_CHARGES

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2	H1'1	-0.372154	-0.940539	0.000008	H	1	NTP	0.2531	****
3	H1'2	-0.372155	0.470271	0.814530	H	1	NTP	0.2531	****
4	H1'3	-0.372159	0.470265	-0.814535	H	1	NTP	0.2531	****
5	C2'	1.452008	0.000000	0.000000	CT	1	NTP	0.0723	****
6	H2'1	1.811485	0.709863	-0.734490	HP	1	NTP	0.0819	****
7	H2'2	1.771785	-0.985591	-0.314071	HP	1	NTP	0.0819	****
8	C3'	2.081072	0.353118	1.345826	CT	1	NTP	-0.0606	****
9	H3'1	1.845385	1.366929	1.616100	H1	1	NTP	0.0894	****
10	H3'2	3.158194	0.304327	1.243103	H1	1	NTP	0.0894	****
11	N4'	1.700248	-0.561133	2.421819	N	1	NTP	-0.0615	****
12	C5'	2.457440	-1.806866	2.505076	CT	1	NTP	-0.1251	****
13	H5'1	1.863229	-2.543139	3.023203	H1	1	NTP	0.0674	****
14	H5'2	2.662844	-2.156139	1.504430	H1	1	NTP	0.0674	****
15	C'	3.809063	-1.612312	3.190000	C	1	NTP	0.6140	****
16	O1'	4.817371	-1.511348	2.540609	O	1	NTP	-0.5754	****
17	C7'	0.878322	-0.269011	3.448307	C	1	NTP	0.4500	****
18	O7'	0.792780	-0.971906	4.429195	O	1	NTP	-0.5598	****
19	C8'	-0.056744	0.937589	3.365388	CT	1	NTP	-0.0520	****
20	H8'1	-1.042726	0.540421	3.161924	H1	1	NTP	0.0911	****
21	H8'2	0.180976	1.642556	2.592461	H1	1	NTP	0.0911	****
22	N1	-0.104447	1.632386	4.636542	N*	1	NTP	-0.0517	****
23	C2	0.939154	2.476729	4.913690	C	1	NTP	0.6341	****
24	O2	1.837971	2.664708	4.142556	O	1	NTP	-0.5750	****
25	N3	0.870934	3.085554	6.134928	NA	1	NTP	-0.5177	****
26	H3	1.612843	3.720536	6.342740	H	1	NTP	0.3442	****
27	C4	-0.072382	2.883646	7.131613	C	1	NTP	0.6257	****
28	O4	0.000058	3.479989	8.164933	O	1	NTP	-0.5597	****
29	C5	-1.106094	1.899713	6.795010	CM	1	NTP	0.0099	****
30	C6	-1.052923	1.330010	5.594062	CM	1	NTP	-0.2934	****
31	H6	-1.767408	0.590947	5.290795	H4	1	NTP	0.2468	****
32	C7	-2.148437	1.589409	7.832373	CT	1	NTP	-0.2418	****
33	H71	-1.689595	1.210718	8.738601	HC	1	NTP	0.0843	****
34	H72	-2.702209	2.481776	8.101512	HC	1	NTP	0.0843	****
35	H73	-2.848081	0.846914	7.465728	HC	1	NTP	0.0843	****

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@<TRIPOS>SUBSTRUCTURE

1 NTP

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C' 1

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Triplos19: CTP

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CTP

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SMALL

USER_CHARGES

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2	O1'	-0.511689	-1.124427	0.000000	O2	1	CTP	-0.8206	****
3	OXT	-0.540795	1.108412	0.000000	O2	1	CTP	-0.8206	****
4	N1'	4.594918	2.187026	1.178241	N	1	CTP	-0.4302	****
5	H1'1	5.184625	1.392533	1.087575	H	1	CTP	0.2931	****
6	C2'	3.310261	2.005883	1.830291	CT	1	CTP	-0.0573	****
7	H2'1	3.231111	2.690954	2.665166	H1	1	CTP	0.0960	****
8	H2'2	3.293666	0.999863	2.230210	H1	1	CTP	0.0960	****
9	C3'	2.105344	2.229808	0.919414	CT	1	CTP	-0.0479	****
10	H3'1	2.063583	3.254873	0.597069	H1	1	CTP	0.0959	****
11	H3'2	1.204634	2.050699	1.493665	H1	1	CTP	0.0959	****
12	N4'	2.076301	1.332626	-0.235083	N	1	CTP	-0.0212	****
13	C5'	1.527696	0.000000	0.000000	CT	1	CTP	-0.4798	****
14	H5'1	1.914893	-0.674301	-0.747689	H1	1	CTP	0.1621	****
15	H5'2	1.839074	-0.336457	0.977402	H1	1	CTP	0.1621	****
16	C7'	2.306081	1.688413	-1.513829	C	1	CTP	0.4669	****
17	O7'	2.031510	0.967337	-2.445592	O	1	CTP	-0.5706	****
18	C8'	3.016889	3.004414	-1.830052	CT	1	CTP	-0.0673	****

19	H8'1	4.031033	2.740822	-2.100850	H1	1	CTP	0.0902	****
20	H8'2	3.063321	3.701184	-1.015644	H1	1	CTP	0.0902	****
21	N1	2.400705	3.654210	-2.969760	N*	1	CTP	-0.0463	****
22	C2	1.245600	4.351399	-2.727920	C	1	CTP	0.6285	****
23	O2	0.772164	4.453010	-1.630929	O	1	CTP	-0.5743	****
24	N3	0.680895	4.920280	-3.834368	NA	1	CTP	-0.5149	****
25	H3	-0.149541	5.449972	-3.671221	H	1	CTP	0.3440	****
26	C4	1.094352	4.802409	-5.153138	C	1	CTP	0.6249	****
27	O4	0.491041	5.348029	-6.028886	O	1	CTP	-0.5597	****
28	C5	2.285142	3.967683	-5.340862	CM	1	CTP	0.0064	****
29	C6	2.849082	3.438753	-4.258333	CM	1	CTP	-0.2916	****
30	H6	3.711306	2.805822	-4.326463	H4	1	CTP	0.2495	****
31	C7	2.781780	3.753244	-6.743094	CT	1	CTP	-0.2329	****
32	H71	2.017752	3.288368	-7.355748	HC	1	CTP	0.0818	****
33	H72	3.037418	4.697613	-7.210250	HC	1	CTP	0.0818	****
34	H73	3.659725	3.117307	-6.748038	HC	1	CTP	0.0818	****

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@<TRIPOS>SUBSTRUCTURE

1 CTP

1 ****

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N1' 1

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Triplos20: MTPN

@<TRIPOS>MOLECULE

MTPN

45 45 3 0 1

SMALL

USER_CHARGES

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2	HH31	-1.683830	-5.002251	-0.315556	HC	1	ACE	0.0513	****
3	HH32	-2.368080	-4.148382	-1.708026	HC	1	ACE	0.0513	****
4	HH33	-3.418621	-4.990733	-0.577015	HC	1	ACE	0.0513	****
5	C	-2.637354	-3.178981	0.220977	C	1	ACE	0.6257	****
6	O	-2.298076	-3.186372	1.373090	O	1	ACE	-0.5596	****
7	N1'	-3.164169	-2.076590	-0.366700	N	2	TPN	-0.4761	****
8	H1'1	-3.506187	-2.157192	-1.296066	H	2	TPN	0.3011	****
9	C2'	-3.574308	-0.918849	0.407734	CT	2	TPN	-0.1798	****
10	H2'1	-4.043432	-1.248639	1.326301	H1	2	TPN	0.1351	****
11	H2'2	-4.323694	-0.395224	-0.172374	H1	2	TPN	0.1351	****
12	C3'	-2.434601	0.029400	0.773185	CT	2	TPN	-0.0676	****
13	H3'1	-1.736380	-0.458108	1.429867	H1	2	TPN	0.1221	****
14	H3'2	-2.847904	0.863153	1.327406	H1	2	TPN	0.1221	****
15	N4'	-1.739615	0.576891	-0.391242	N	2	TPN	-0.1000	****
16	C5'	-2.365484	1.731600	-1.029253	CT	2	TPN	-0.0992	****
17	H5'1	-2.030029	1.789884	-2.052832	H1	2	TPN	0.0703	****
18	H5'2	-3.436772	1.599072	-1.013366	H1	2	TPN	0.0703	****
19	C'	-2.071035	3.030494	-0.280901	C	2	TPN	0.5707	****
20	O1'	-2.885875	3.506896	0.465867	O	2	TPN	-0.5674	****
21	C7'	-0.493775	0.252814	-0.788018	C	2	TPN	0.4407	****
22	O7'	0.116709	0.906616	-1.602495	O	2	TPN	-0.5491	****
23	C8'	0.167399	-1.027414	-0.277164	CT	2	TPN	-0.0520	****
24	H8'1	0.112897	-1.739168	-1.090636	H1	2	TPN	0.0755	****
25	H8'2	-0.302938	-1.469186	0.579881	H1	2	TPN	0.0755	****
26	N1	1.565822	-0.792717	0.023138	N*	2	TPN	-0.0412	****
27	C2	1.842742	-0.187207	1.221262	C	2	TPN	0.6620	****
28	O2	0.985969	0.112374	2.004874	O	2	TPN	-0.5893	****
29	N3	3.169727	0.044652	1.449507	NA	2	TPN	-0.5381	****
30	H3	3.389568	0.465526	2.327773	H	2	TPN	0.3504	****
31	C4	4.231392	-0.185367	0.586981	C	2	TPN	0.6296	****
32	O4	5.349034	0.081073	0.916386	O	2	TPN	-0.5598	****
33	C5	3.845209	-0.751534	-0.709285	CM	2	TPN	0.0092	****
34	C6	2.555494	-1.002612	-0.917275	CM	2	TPN	-0.3059	****
35	H6	2.203698	-1.400670	-1.848115	H4	2	TPN	0.2503	****
36	C7	4.928309	-0.997851	-1.721763	CT	2	TPN	-0.2157	****
37	H71	5.445785	-0.076709	-1.964474	HC	2	TPN	0.0772	****
38	H72	5.668103	-1.689340	-1.334509	HC	2	TPN	0.0772	****
39	H73	4.515420	-1.410559	-2.635231	HC	2	TPN	0.0772	****
40	N	-0.865254	3.571266	-0.519214	N	3	NME	-0.4370	****
41	H	-0.213735	3.033768	-1.047775	H	3	NME	0.3127	****
42	CH3	-0.399200	4.726533	0.216784	CT	3	NME	-0.1122	****
43	HH31	-1.150884	5.502623	0.198654	H1	3	NME	0.0868	****
44	HH32	0.500950	5.096914	-0.255148	H1	3	NME	0.0868	****
45	HH33	-0.181298	4.484551	1.251482	H1	3	NME	0.0868	****

@<TRIPOS>BOND

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7	7	8	1

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     20     19     20 1
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@<TRIPOS>SUBSTRUCTURE

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      2 TPN         7 *****
      3 NME        40 *****

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@<TRIPOS>HEADTAIL

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@<TRIPOS>RESIDUECONNECT

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Tripos21: UPN

@<TRIPOS>MOLECULE

UPN

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	-3.254846	-1.627229	-0.278295	N	1	UPN	-0.4030	****
2	H1'1	-3.684063	-1.618138	-1.174397	H	1	UPN	0.2842	****
3	C2'	-3.417411	-0.453557	0.561161	CT	1	UPN	-0.1241	****
4	H2'1	-3.861967	-0.744434	1.504820	H1	1	UPN	0.1147	****
5	H2'2	-4.116231	0.203849	0.059476	H1	1	UPN	0.1147	****
6	C3'	-2.118053	0.290355	0.861095	CT	1	UPN	-0.0522	****
7	H3'1	-1.458527	-0.326589	1.444893	H1	1	UPN	0.1033	****
8	H3'2	-2.351584	1.154613	1.470650	H1	1	UPN	0.1033	****
9	N4'	-1.434427	0.771128	-0.339044	N	1	UPN	-0.0941	****
10	C5'	-1.916897	2.033395	-0.892647	CT	1	UPN	-0.1112	****
11	H5'1	-1.656731	2.078589	-1.938533	H1	1	UPN	0.0678	****
12	H5'2	-2.991117	2.068858	-0.791737	H1	1	UPN	0.0678	****
13	C'	-1.362836	3.239471	-0.136210	C	1	UPN	0.6209	****
14	O1'	-2.029484	3.803357	0.692087	O	1	UPN	-0.5807	****
15	C7'	-0.290973	0.272717	-0.845814	C	1	UPN	0.4391	****
16	O7'	0.350918	0.853260	-1.691252	O	1	UPN	-0.5502	****
17	C8'	0.198827	-1.112754	-0.424350	CT	1	UPN	-0.0516	****
18	H8'1	-0.027035	-1.775518	-1.249653	H1	1	UPN	0.0866	****
19	H8'2	-0.265877	-1.511529	0.456298	H1	1	UPN	0.0866	****
20	N1	1.636338	-1.105349	-0.233737	N*	1	UPN	-0.0156	****
21	C2	2.097211	-0.613701	0.965992	C	1	UPN	0.5893	****
22	O2	1.354926	-0.236002	1.827302	O	1	UPN	-0.5655	****
23	N3	3.456597	-0.592628	1.095482	NA	1	UPN	-0.4340	****
24	H3	3.799540	-0.257334	1.971222	H	1	UPN	0.3197	****
25	C4	4.409555	-0.925403	0.138007	C	1	UPN	0.6418	****
26	O4	5.574047	-0.843589	0.387444	O	1	UPN	-0.5643	****
27	C5	3.828236	-1.349290	-1.130494	CM	1	UPN	-0.3150	****
28	H5	4.487720	-1.598074	-1.935910	HA	1	UPN	0.1763	****
29	C6	2.505560	-1.401481	-1.258114	CM	1	UPN	-0.1842	****
30	H6	2.035852	-1.690560	-2.176543	H4	1	UPN	0.2296	****

@<TRIPOS>BOND

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@<TRIPOS>SUBSTRUCTURE

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@<TRIPOS>HEADTAIL
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C' 1
@<TRIPOS>RESIDUECONNECT
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Tripes22: NUP

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NUP
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SMALL
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USER_CHARGES
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1	N1'	0.000000	0.000000	0.000000	N3	1	NUP	0.0049	****
2	H1'1	-0.372154	-0.940539	0.000008	H	1	NUP	0.2531	****
3	H1'2	-0.372155	0.470271	0.814530	H	1	NUP	0.2531	****
4	H1'3	-0.372159	0.470265	-0.814535	H	1	NUP	0.2531	****
5	C2'	1.452109	0.000000	0.000000	CT	1	NUP	0.0723	****
6	H2'1	1.812299	0.711263	-0.732871	HP	1	NUP	0.0819	****
7	H2'2	1.771672	-0.985033	-0.315927	HP	1	NUP	0.0819	****
8	C3'	2.081304	0.350353	1.346500	CT	1	NUP	-0.0606	****
9	H3'1	1.846313	1.363945	1.618205	H1	1	NUP	0.0894	****
10	H3'2	3.158368	0.300601	1.244060	H1	1	NUP	0.0894	****
11	N4'	1.699564	-0.565259	2.421094	N	1	NUP	-0.0615	****
12	C5'	2.453774	-1.813144	2.501840	CT	1	NUP	-0.1251	****
13	H5'1	1.856556	-2.549965	3.015681	H1	1	NUP	0.0674	****
14	H5'2	2.661033	-2.159104	1.500469	H1	1	NUP	0.0674	****
15	C'	3.803854	-1.622760	3.190926	C	1	NUP	0.6140	****
16	O1'	4.813082	-1.515316	2.544106	O	1	NUP	-0.5754	****
17	C7'	0.875750	-0.275429	3.445811	C	1	NUP	0.4500	****
18	O7'	0.784373	-0.981482	4.424055	O	1	NUP	-0.5598	****
19	C8'	-0.055249	0.934575	3.367104	CT	1	NUP	-0.0400	****
20	H8'1	-1.042748	0.541471	3.162874	H1	1	NUP	0.0869	****
21	H8'2	0.183561	1.643560	2.598454	H1	1	NUP	0.0869	****
22	N1	-0.100003	1.622919	4.642646	N*	1	NUP	-0.0241	****
23	C2	0.939335	2.481954	4.917068	C	1	NUP	0.5863	****
24	O2	1.825630	2.680119	4.135588	O	1	NUP	-0.5667	****
25	N3	0.879040	3.086369	6.140256	NA	1	NUP	-0.4267	****
26	H3	1.617910	3.727759	6.339359	H	1	NUP	0.3185	****
27	C4	-0.050122	2.870611	7.153040	C	1	NUP	0.6353	****
28	O4	0.029837	3.463892	8.185793	O	1	NUP	-0.5631	****
29	C5	-1.060964	1.878469	6.805825	CM	1	NUP	-0.3089	****
30	H5	-1.801481	1.627518	7.536691	HA	1	NUP	0.1749	****
31	C6	-1.028849	1.305058	5.606387	CM	1	NUP	-0.1830	****
32	H6	-1.740853	0.559276	5.315589	H4	1	NUP	0.2282	****

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@<TRIPOS>SUBSTRUCTURE

1 NUP

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

0 0

C' 1

@<TRIPOS>RESIDUECONNECT

1 0 C' 0 0 0 0

Tripos23: CUP

@<TRIPOS>MOLECULE

CUP

31 31 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	C'	0.000000	0.000000	0.000000	C	1	CUP	0.7881	****
2	O1'	-0.511689	-1.124427	0.000000	O2	1	CUP	-0.8206	****
3	OXT	-0.540795	1.108412	0.000000	O2	1	CUP	-0.8206	****
4	N1'	4.598729	2.180074	1.176275	N	1	CUP	-0.4302	****
5	H1'1	5.190931	1.386592	1.092655	H	1	CUP	0.2931	****
6	C2'	3.315434	1.999899	1.831492	CT	1	CUP	-0.0573	****
7	H2'1	3.239051	2.683291	2.668076	H1	1	CUP	0.0960	****
8	H2'2	3.298284	0.993071	2.229263	H1	1	CUP	0.0960	****
9	C3'	2.108361	2.227674	0.924414	CT	1	CUP	-0.0479	****
10	H3'1	2.067159	3.253702	0.605067	H1	1	CUP	0.0959	****
11	H3'2	1.208917	2.047858	1.500351	H1	1	CUP	0.0959	****
12	N4'	2.075114	1.333719	-0.232548	N	1	CUP	-0.0212	****
13	C5'	1.527678	0.000000	0.000000	CT	1	CUP	-0.4798	****
14	H5'1	1.915516	-0.672557	-0.748902	H1	1	CUP	0.1621	****
15	H5'2	1.839314	-0.337875	0.976787	H1	1	CUP	0.1621	****
16	C7'	2.304821	1.691703	-1.509967	C	1	CUP	0.4669	****
17	O7'	2.032312	0.972538	-2.443969	O	1	CUP	-0.5706	****
18	C8'	3.012295	3.009724	-1.825188	CT	1	CUP	-0.0535	****
19	H8'1	4.026105	2.748689	-2.099861	H1	1	CUP	0.0856	****
20	H8'2	3.059604	3.707024	-1.011525	H1	1	CUP	0.0856	****

21	N1	2.390707	3.658532	-2.963390	N*	1	CUP	-0.0186	****
22	C2	1.241358	4.373070	-2.714590	C	1	CUP	0.5820	****
23	O2	0.785902	4.483801	-1.611958	O	1	CUP	-0.5661	****
24	N3	0.667579	4.942185	-3.815511	NA	1	CUP	-0.4276	****
25	H3	-0.155136	5.480476	-3.641340	H	1	CUP	0.3191	****
26	C4	1.058777	4.812173	-5.144303	C	1	CUP	0.6385	****
27	O4	0.448336	5.358730	-6.012399	O	1	CUP	-0.5640	****
28	C5	2.232367	3.964335	-5.319130	CM	1	CUP	-0.3144	****
29	H5	2.588400	3.784728	-6.312352	HA	1	CUP	0.1756	****
30	C6	2.816473	3.428732	-4.251220	CM	1	CUP	-0.1798	****
31	H6	3.669815	2.786407	-4.334471	H4	1	CUP	0.2297	****

@<TRIPOS>BOND

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@<TRIPOS>SUBSTRUCTURE

1 CUP

1 ****

0 **** *

@<TRIPOS>HEADTAIL

N1' 1

0 0

@<TRIPOS>RESIDUECONNECT

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Tripas24: MUPN

@<TRIPOS>MOLECULE

MUPN

42 42 3 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	CH3	-3.003190	-4.010912	-0.692202	CT	1	ACE	-0.1596	****
2	HH31	-2.257828	-4.748586	-0.431447	HC	1	ACE	0.0528	****
3	HH32	-2.903897	-3.742706	-1.738084	HC	1	ACE	0.0528	****
4	HH33	-3.983526	-4.453537	-0.545924	HC	1	ACE	0.0528	****
5	C	-2.861669	-2.820591	0.231677	C	1	ACE	0.6302	****
6	O	-2.433479	-2.925728	1.348865	O	1	ACE	-0.5606	****
7	N1'	-3.254846	-1.627229	-0.278295	N	2	UPN	-0.4780	****
8	H1'1	-3.684063	-1.618138	-1.174397	H	2	UPN	0.3011	****
9	C2'	-3.417411	-0.453557	0.561161	CT	2	UPN	-0.1800	****
10	H2'1	-3.861967	-0.744434	1.504820	H1	2	UPN	0.1370	****
11	H2'2	-4.116231	0.203849	0.059476	H1	2	UPN	0.1370	****
12	C3'	-2.118053	0.290355	0.861095	CT	2	UPN	-0.0797	****
13	H3'1	-1.458527	-0.326589	1.444893	H1	2	UPN	0.1269	****
14	H3'2	-2.351584	1.154613	1.470650	H1	2	UPN	0.1269	****
15	N4'	-1.434427	0.771128	-0.339044	N	2	UPN	-0.1075	****
16	C5'	-1.916897	2.033395	-0.892647	CT	2	UPN	-0.0994	****
17	H5'1	-1.656731	2.078589	-1.938533	H1	2	UPN	0.0708	****
18	H5'2	-2.991117	2.068858	-0.791737	H1	2	UPN	0.0708	****
19	C'	-1.362836	3.239471	-0.136210	C	2	UPN	0.5717	****
20	O1'	-2.029484	3.803357	0.692087	O	2	UPN	-0.5673	****
21	C7'	-0.290973	0.272717	-0.845814	C	2	UPN	0.4544	****
22	O7'	0.350918	0.853260	-1.691252	O	2	UPN	-0.5498	****
23	C8'	0.198827	-1.112754	-0.424350	CT	2	UPN	-0.0603	****
24	H8'1	-0.027035	-1.775518	-1.249653	H1	2	UPN	0.0739	****
25	H8'2	-0.265877	-1.511529	0.456298	H1	2	UPN	0.0739	****
26	N1	1.636338	-1.105349	-0.233737	N*	2	UPN	-0.0052	****
27	C2	2.097211	-0.613701	0.965992	C	2	UPN	0.6187	****
28	O2	1.354926	-0.236002	1.827302	O	2	UPN	-0.5825	****
29	N3	3.456597	-0.592628	1.095482	NA	2	UPN	-0.4525	****
30	H3	3.799540	-0.257334	1.971222	H	2	UPN	0.3261	****
31	C4	4.409555	-0.925403	0.138007	C	2	UPN	0.6427	****
32	O4	5.574047	-0.843589	0.387444	O	2	UPN	-0.5638	****
33	C5	3.828236	-1.349290	-1.130494	CM	2	UPN	-0.3153	****
34	H5	4.487720	-1.598074	-1.935910	HA	2	UPN	0.1770	****
35	C6	2.505560	-1.401481	-1.258114	CM	2	UPN	-0.1892	****
36	H6	2.035852	-1.690560	-2.176543	H4	2	UPN	0.2305	****
37	N	-0.110798	3.598190	-0.462810	N	3	NME	-0.4360	****
38	H	0.403391	2.990697	-1.062166	H	3	NME	0.3119	****
39	CH3	0.587826	4.638500	0.260751	CT	3	NME	-0.1202	****
40	HH31	-0.031804	5.521352	0.326512	H1	3	NME	0.0890	****
41	HH32	1.494443	4.883562	-0.275972	H1	3	NME	0.0890	****
42	HH33	0.846659	4.325829	1.266638	H1	3	NME	0.0890	****

@<TRIPOS>BOND

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@<TRIPOS>SUBSTRUCTURE

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1 ACE 1 **** 0 **** ****
2 UPN 7 **** 0 **** ****
3 NME 37 **** 0 **** ****

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@<TRIPOS>HEADTAIL

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0 0

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@<TRIPOS>RESIDUECONNECT

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frcmod.pna

PNA frcmod file: frcmod.pna
MASS

BOND

CQ-NA 502.0 1.324 by analogy to CQ-NC, parm99.dat

ANGL

C -CT-N* 50.00 110.13 FF cst by analogy to CT-CT-N*; Eq.val. from
QM geo.opt.
CQ-NA-C 70.0 118.60 by analogy to CA-NC-CQ, parm99.dat
H5-CQ-NA 50.0 115.45 by analogy to H5-CQ-NC, parm99.dat
NC-CQ-NA 70.0 129.10 by analogy to NC-CQ-NC, parm99.dat
CQ-NA-H 50.0 118.00 by analogy to CA-NA-H, parm99.dat

DIHEDRAL

X -CQ-NA-X 4 6.00 180.0 2.0 by analogy to X -CA-NA-X,
parm99.dat

NONB

8.2. Gamma modified PNA

GAP

@<TRIPOS>MOLECULE

GAP

37 38 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	2.658286	-1.774021	0.309197	N	1	GAP	-0.3871	0.0000	****
2	H1'1	2.517951	-2.759747	0.268759	H	1	GAP	0.2784	0.0000	****
3	C2'	2.512277	-1.069824	-0.955646	CT	1	GAP	0.0528	0.0000	****
4	H2'1	3.410703	-0.490010	-1.113776	H1	1	GAP	0.1363	0.0000	****
5	C3'	1.288324	-0.144835	-0.973096	CT	1	GAP	-0.1234	0.0000	****
6	H3'1	0.383739	-0.730991	-0.927254	H1	1	GAP	0.0809	0.0000	****
7	H3'2	1.264585	0.390541	-1.914799	H1	1	GAP	0.0809	0.0000	****
8	N4'	1.282584	0.854551	0.086253	N	1	GAP	-0.1034	0.0000	****
9	C5'	2.106475	2.040867	-0.129854	CT	1	GAP	-0.1424	0.0000	****
10	H5'1	2.401438	2.436687	0.827763	H1	1	GAP	0.0802	0.0000	****
11	H5'2	2.992408	1.751034	-0.670394	H1	1	GAP	0.0802	0.0000	****
12	C'	1.360971	3.072713	-0.969332	C	1	GAP	0.6199	0.0000	****
13	O1'	1.478995	3.099056	-2.168198	O	1	GAP	-0.5806	0.0000	****
14	C7'	0.496122	0.835159	1.175840	C	1	GAP	0.5312	0.0000	****
15	O7'	0.374438	1.789833	1.910183	O	1	GAP	-0.5649	0.0000	****
16	C8'	-0.284550	-0.426263	1.541418	CT	1	GAP	-0.0325	0.0000	****
17	H8'1	-0.142787	-0.557902	2.605388	H1	1	GAP	0.0716	0.0000	****
18	H8'2	0.046395	-1.316442	1.039739	H1	1	GAP	0.0716	0.0000	****
19	C9'	2.387422	-2.077500	-2.092261	CT	1	GAP	0.1284	0.0000	****
20	H9'1	2.350846	-1.540092	-3.035476	H1	1	GAP	0.0497	0.0000	****
21	H9'2	1.458038	-2.637070	-1.992424	H1	1	GAP	0.0497	0.0000	****
22	O10'	3.491037	-2.940017	-2.041878	OH	1	GAP	-0.6690	0.0000	****
23	H10'	3.469878	-3.530498	-2.781888	HO	1	GAP	0.4382	0.0000	****
24	N9	-1.696155	-0.258620	1.262813	N*	1	GAP	-0.1390	0.0000	****
25	C8	-2.584459	0.581549	1.893238	CK	1	GAP	0.1714	0.0000	****
26	H8	-2.252210	1.236539	2.671358	H5	1	GAP	0.1526	0.0000	****
27	N7	-3.783280	0.494507	1.451181	NB	1	GAP	-0.5554	0.0000	****
28	C6	-4.658517	-1.015694	-0.409182	CA	1	GAP	0.6677	0.0000	****
29	N6	-5.953672	-0.665697	-0.378225	N2	1	GAP	-0.8735	0.0000	****
30	H61	-6.241331	0.111927	0.169797	H	1	GAP	0.4008	0.0000	****
31	H62	-6.551887	-0.995486	-1.100244	H	1	GAP	0.4008	0.0000	****
32	C5	-3.709216	-0.461543	0.457170	CB	1	GAP	0.0129	0.0000	****
33	C4	-2.424041	-0.934376	0.328921	CB	1	GAP	0.4418	0.0000	****
34	N3	-2.004789	-1.856143	-0.534156	NC	1	GAP	-0.6782	0.0000	****
35	C2	-2.986448	-2.293220	-1.291757	CQ	1	GAP	0.5001	0.0000	****
36	H2	-2.725981	-3.040636	-2.019527	H5	1	GAP	0.0648	0.0000	****
37	N1	-4.262161	-1.935453	-1.282934	NC	1	GAP	-0.7139	0.0000	****

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8	5	8	1
9	8	9	1
10	8	14	1

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11      9      10 1
12      9      11 1
13      9      12 1
14     12      13 1
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18     16      18 1
19     16      24 1
20     19      20 1
21     19      21 1
22     19      22 1
23     22      23 1
24     24      25 1
25     24      33 1
26     25      26 1
27     25      27 1
28     27      32 1
29     28      29 1
30     28      32 1
31     28      37 1
32     29      30 1
33     29      31 1
34     32      33 1
35     33      34 1
36     34      35 1
37     35      36 1
38     35      37 1

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@<TRIPOS>SUBSTRUCTURE

1 GAP

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

C' 1

@<TRIPOS>RESIDUECONNECT

1 N1' C' 0 0 0 0

GNA

@<TRIPOS>MOLECULE

GNA

39 40 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	0.000000	0.000000	-0.000000	N3	1	GNA	-0.0679	0.0000	****
2	H1'1	-0.372149	0.940540	-0.000000	H	1	GNA	0.2683	0.0000	****
3	H1'2	-0.372151	-0.470283	-0.814524	H	1	GNA	0.2683	0.0000	****
4	H1'3	-0.372144	-0.470264	0.814532	H	1	GNA	0.2683	0.0000	****
5	C2'	1.507376	-0.000000	0.000000	CT	1	GNA	0.3376	0.0000	****
6	H2'1	1.835302	0.530722	-0.882699	HP	1	GNA	0.0685	0.0000	****
7	C3'	2.093047	-1.418083	-0.000000	CT	1	GNA	-0.0833	0.0000	****
8	H3'1	1.860282	-1.912064	0.930461	H1	1	GNA	0.0545	0.0000	****
9	H3'2	3.173169	-1.353284	-0.055978	H1	1	GNA	0.0545	0.0000	****
10	N4'	1.656411	-2.242214	-1.118563	N	1	GNA	-0.0870	0.0000	****
11	C5'	2.335753	-2.027012	-2.393328	CT	1	GNA	-0.1820	0.0000	****
12	H5'1	1.665262	-2.295744	-3.192661	H1	1	GNA	0.0902	0.0000	****
13	H5'2	2.576470	-0.980542	-2.482883	H1	1	GNA	0.0902	0.0000	****
14	C'	3.639722	-2.815302	-2.452241	C	1	GNA	0.6203	0.0000	****
15	O1'	4.682808	-2.314574	-2.115868	O	1	GNA	-0.5801	0.0000	****

16	C7'	0.778763	-3.257425	-1.046602	C	1	GNA	0.5146	0.0000	****
17	O7'	0.614652	-4.043909	-1.952130	O	1	GNA	-0.5620	0.0000	****
18	C8'	-0.071201	-3.454963	0.207519	CT	1	GNA	-0.0347	0.0000	****
19	H8'1	-1.074053	-3.647828	-0.148221	H1	1	GNA	0.0860	0.0000	****
20	H8'2	-0.099131	-2.606384	0.865363	H1	1	GNA	0.0860	0.0000	****
21	C9'	2.020273	0.747366	1.225265	CT	1	GNA	0.0891	0.0000	****
22	H9'1	3.103980	0.801223	1.175501	H1	1	GNA	0.0461	0.0000	****
23	H9'2	1.755925	0.202261	2.130705	H1	1	GNA	0.0461	0.0000	****
24	O10'	1.448284	2.026892	1.234387	OH	1	GNA	-0.6783	0.0000	****
25	H10'	1.807919	2.533213	1.949252	HO	1	GNA	0.4459	0.0000	****
26	N9	0.393781	-4.594065	0.972105	N*	1	GNA	-0.1541	0.0000	****
27	C8	0.341518	-5.922617	0.619089	CK	1	GNA	0.1833	0.0000	****
28	H8	-0.051243	-6.215725	-0.332068	H5	1	GNA	0.1524	0.0000	****
29	N7	0.803974	-6.714993	1.512642	NB	1	GNA	-0.5620	0.0000	****
30	C6	1.778116	-6.105908	3.790308	CA	1	GNA	0.6730	0.0000	****
31	N6	2.050565	-7.339154	4.243912	N2	1	GNA	-0.8765	0.0000	****
32	H61	1.979390	-8.117315	3.629767	H	1	GNA	0.4021	0.0000	****
33	H62	2.578638	-7.431150	5.080936	H	1	GNA	0.4021	0.0000	****
34	C5	1.197929	-5.876982	2.537392	CB	1	GNA	0.0116	0.0000	****
35	C4	0.951607	-4.562632	2.215673	CB	1	GNA	0.4629	0.0000	****
36	N3	1.213693	-3.501901	2.974942	NC	1	GNA	-0.7199	0.0000	****
37	C2	1.759251	-3.848084	4.120007	CQ	1	GNA	0.5327	0.0000	****
38	H2	2.004031	-3.043609	4.790101	H5	1	GNA	0.0597	0.0000	****
39	N1	2.052751	-5.061197	4.564679	NC	1	GNA	-0.7265	0.0000	****

@<TRIPOS>BOND

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37	35	36	1

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38    36    37 1
39    37    38 1
40    37    39 1
@<TRIPOS>SUBSTRUCTURE
1 GNA          1 ****          0 ****    ****
@<TRIPOS>HEADTAIL
C' 1
0 0
@<TRIPOS>RESIDUECONNECT
1 C' 0 0 0 0 0

```

GCA

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@<TRIPOS>MOLECULE
GCA
38    39    1    0    1
SMALL
USER_CHARGES
@<TRIPOS>ATOM
1 N1'  -3.092953  1.875939  -1.401830  N    1  GCA  -0.3796  0.0000  ****
2 H1'1  -3.669103  2.591833  -1.787227  H    1  GCA   0.2738  0.0000  ****
3 C2'  -1.848722  1.624843  -2.113083  CT   1  GCA   0.0724  0.0000  ****
4 H2'1  -1.808558  0.568982  -2.340904  H1   1  GCA   0.1339  0.0000  ****
5 C3'  -0.614803  2.025996  -1.294248  CT   1  GCA  -0.1382  0.0000  ****
6 H3'1  -0.594430  3.096322  -1.160303  H1   1  GCA   0.0801  0.0000  ****
7 H3'2   0.277515  1.765196  -1.850798  H1   1  GCA   0.0801  0.0000  ****
8 N4'  -0.518931  1.365121   0.000000  N    1  GCA  -0.0476  0.0000  ****
9 C5'   0.000000  -0.000000  0.000000  CT   1  GCA  -0.5030  0.0000  ****
10 H5'1  -0.403557  -0.523804  0.850575  H1   1  GCA   0.1684  0.0000  ****
11 H5'2  -0.331675  -0.494469  -0.898085  H1   1  GCA   0.1684  0.0000  ****
12 C7'  -0.747400  1.942265  1.191975  C    1  GCA   0.5677  0.0000  ****
13 O7'  -0.446174  1.416833  2.240142  O    1  GCA  -0.5857  0.0000  ****
14 C8'  -1.420571  3.311601  1.269709  CT   1  GCA  -0.0382  0.0000  ****
15 H8'1  -2.164701  3.226002  2.049710  H1   1  GCA   0.0709  0.0000  ****
16 H8'2  -1.908549  3.620531  0.364135  H1   1  GCA   0.0709  0.0000  ****
17 C9'  -1.843819  2.397817  -3.426620  CT   1  GCA   0.1197  0.0000  ****
18 H9'1  -0.943017  2.145984  -3.978831  H1   1  GCA   0.0523  0.0000  ****
19 H9'2  -1.823057  3.468702  -3.227584  H1   1  GCA   0.0523  0.0000  ****
20 O10'  -2.994762  2.051758  -4.147740  OH   1  GCA  -0.6738  0.0000  ****
21 H10'  -2.976589  2.467427  -4.998398  HO   1  GCA   0.4408  0.0000  ****
22 N9   -0.463618  4.337528  1.630357  N*   1  GCA  -0.1392  0.0000  ****
23 C8    0.192134  4.492598  2.829668  CK   1  GCA   0.1702  0.0000  ****
24 H8    0.044541  3.789953  3.623002  H5   1  GCA   0.1557  0.0000  ****
25 N7    0.962701  5.514703  2.870779  NB   1  GCA  -0.5571  0.0000  ****
26 C6    1.392859  7.224339  1.026923  CA   1  GCA   0.6665  0.0000  ****
27 N6    2.245853  8.031004  1.676982  N2   1  GCA  -0.8738  0.0000  ****
28 H61   2.610992  7.754402  2.558991  H    1  GCA   0.4010  0.0000  ****
29 H62   2.712648  8.741783  1.162367  H    1  GCA   0.4010  0.0000  ****
30 C5    0.826780  6.091802  1.623359  CB   1  GCA   0.0181  0.0000  ****
31 C4   -0.050894  5.373124  0.845616  CB   1  GCA   0.4347  0.0000  ****
32 N3   -0.404461  5.650092  -0.406824  NC   1  GCA  -0.6715  0.0000  ****
33 C2    0.196789  6.731005  -0.852668  CQ   1  GCA   0.4972  0.0000  ****
34 H2   -0.035659  7.019752  -1.861983  H5   1  GCA   0.0651  0.0000  ****
35 N1    1.057721  7.518697  -0.225089  NC   1  GCA  -0.7129  0.0000  ****
36 C'    1.524863  0.000000  -0.000000  C    1  GCA   0.8129  0.0000  ****
37 O1'   2.036542  -1.124430  -0.000000  O2   1  GCA  -0.8266  0.0000  ****
38 OXT   2.065633  1.108429  -0.000000  O2   1  GCA  -0.8266  0.0000  ****
@<TRIPOS>BOND

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12     9     11  1
13     9     36  1
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39    36     38  1

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@<TRIPOS>SUBSTRUCTURE

1 GCA

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

0 0

@<TRIPOS>RESIDUECONNECT

1 N1' 0 0 0 0 0

MAP

@<TRIPOS>MOLECULE

MAP

49 50 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

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1 C1      3.888841 -2.360831  2.319522 CT  1  MAP -0.1796  0.0000 ****
2 H1      3.018350 -2.933619  2.619418 HC  1  MAP  0.0586  0.0000 ****
3 H3      4.634525 -3.052384  1.938797 HC  1  MAP  0.0586  0.0000 ****

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4	H4	4.300237	-1.845023	3.174709	HC	1	MAP	0.0586	0.0000	****
5	C	3.558559	-1.348791	1.244087	C	1	MAP	0.5788	0.0000	****
6	O	4.051954	-0.256753	1.220008	O	1	MAP	-0.5764	0.0000	****
7	N1'	2.658286	-1.774021	0.309197	N	1	MAP	-0.3396	0.0000	****
8	H1'1	2.517951	-2.759747	0.268759	H	1	MAP	0.2738	0.0000	****
9	C2'	2.512277	-1.069824	-0.955646	CT	1	MAP	0.0565	0.0000	****
10	H2'1	3.410703	-0.490010	-1.113776	H1	1	MAP	0.1317	0.0000	****
11	C3'	1.288324	-0.144835	-0.973096	CT	1	MAP	-0.1419	0.0000	****
12	H3'1	0.383739	-0.730991	-0.927254	H1	1	MAP	0.0511	0.0000	****
13	H3'2	1.264585	0.390541	-1.914799	H1	1	MAP	0.0511	0.0000	****
14	N4'	1.282584	0.854551	0.086253	N	1	MAP	0.0039	0.0000	****
15	C5'	2.106475	2.040867	-0.129854	CT	1	MAP	0.0403	0.0000	****
16	H5'1	2.401438	2.436687	0.827763	H1	1	MAP	0.0436	0.0000	****
17	H5'2	2.992408	1.751034	-0.670394	H1	1	MAP	0.0436	0.0000	****
18	C'	1.360971	3.072713	-0.969332	C	1	MAP	0.5508	0.0000	****
19	O1'	1.478995	3.099056	-2.168198	O	1	MAP	-0.5659	0.0000	****
20	C7'	0.496122	0.835159	1.175840	C	1	MAP	0.4353	0.0000	****
21	O7'	0.374438	1.789833	1.910183	O	1	MAP	-0.5726	0.0000	****
22	C8'	-0.284550	-0.426263	1.541418	CT	1	MAP	-0.0170	0.0000	****
23	H8'1	-0.142787	-0.557902	2.605388	H1	1	MAP	0.0335	0.0000	****
24	H8'2	0.046395	-1.316442	1.039739	H1	1	MAP	0.0335	0.0000	****
25	C9'	2.387422	-2.077500	-2.092261	CT	1	MAP	0.1172	0.0000	****
26	H9'1	2.350846	-1.540092	-3.035476	H1	1	MAP	0.0482	0.0000	****
27	H9'2	1.458038	-2.637070	-1.992424	H1	1	MAP	0.0482	0.0000	****
28	O10'	3.491037	-2.940017	-2.041878	OH	1	MAP	-0.6940	0.0000	****
29	H10'	3.469878	-3.530498	-2.781888	HO	1	MAP	0.4523	0.0000	****
30	N9	-1.696155	-0.258620	1.262813	N*	1	MAP	-0.0523	0.0000	****
31	C8	-2.584459	0.581549	1.893238	CK	1	MAP	0.1404	0.0000	****
32	H8	-2.252210	1.236539	2.671358	H5	1	MAP	0.1674	0.0000	****
33	N7	-3.783280	0.494507	1.451181	NB	1	MAP	-0.5545	0.0000	****
34	C6	-4.658517	-1.015694	-0.409182	CA	1	MAP	0.6713	0.0000	****
35	N6	-5.953672	-0.665697	-0.378225	N2	1	MAP	-0.8862	0.0000	****
36	H61	-6.241331	0.111927	0.169797	H	1	MAP	0.4040	0.0000	****
37	H62	-6.551887	-0.995486	-1.100244	H	1	MAP	0.4040	0.0000	****
38	C5	-3.709216	-0.461543	0.457170	CB	1	MAP	0.0243	0.0000	****
39	C4	-2.424041	-0.934376	0.328921	CB	1	MAP	0.3730	0.0000	****
40	N3	-2.004789	-1.856143	-0.534156	NC	1	MAP	-0.5797	0.0000	****
41	C2	-2.986448	-2.293220	-1.291757	CQ	1	MAP	0.4352	0.0000	****
42	H2	-2.725981	-3.040636	-2.019527	H5	1	MAP	0.0749	0.0000	****
43	N1	-4.262161	-1.935453	-1.282934	NC	1	MAP	-0.6954	0.0000	****
44	N	0.570964	3.913920	-0.279606	N	1	MAP	-0.4442	0.0000	****
45	H	0.403803	3.700114	0.679105	H	1	MAP	0.3013	0.0000	****
46	CH3	-0.293905	4.857551	-0.952089	CT	1	MAP	-0.1674	0.0000	****
47	HH31	-1.128685	4.366843	-1.442432	H1	1	MAP	0.1005	0.0000	****
48	HH32	-0.678207	5.557668	-0.222257	H1	1	MAP	0.1005	0.0000	****
49	HH33	0.267223	5.400512	-1.698807	H1	1	MAP	0.1005	0.0000	****

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14	11	14 1
15	14	15 1

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@<TRIPOS>SUBSTRUCTURE

1 MAP

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

0 0

0 0

@<TRIPOS>RESIDUECONNECT

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GTP

@<TRIPOS>MOLECULE

GTP

37 37 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	-3.098596	-1.629931	-0.637122	N	1	GTP	-0.3871	0.0000	****
2	H1'1	-3.847978	-1.763996	-1.277424	H	1	GTP	0.2784	0.0000	****
3	C2'	-3.349478	-0.638782	0.397211	CT	1	GTP	0.0528	0.0000	****
4	H2'1	-4.055355	0.056271	-0.037132	H1	1	GTP	0.1363	0.0000	****
5	C3'	-2.104015	0.144775	0.842890	CT	1	GTP	-0.1234	0.0000	****
6	H3'1	-1.537891	-0.463569	1.523138	H1	1	GTP	0.0809	0.0000	****
7	H3'2	-2.454305	1.016378	1.386112	H1	1	GTP	0.0809	0.0000	****

8	N4'	-1.239246	0.566306	-0.248807	N	1	GTP	-0.1034	0.0000	****
9	C5'	-1.823333	1.401828	-1.291874	CT	1	GTP	-0.1424	0.0000	****
10	H5'1	-1.199862	1.338948	-2.168107	H1	1	GTP	0.0802	0.0000	****
11	H5'2	-2.806879	1.036260	-1.535584	H1	1	GTP	0.0802	0.0000	****
12	C'	-1.973998	2.841207	-0.811414	C	1	GTP	0.6199	0.0000	****
13	O1'	-3.011624	3.225255	-0.334363	O	1	GTP	-0.5806	0.0000	****
14	C7'	0.113600	0.464041	-0.230592	C	1	GTP	0.5312	0.0000	****
15	O7'	0.800996	0.961161	-1.086483	O	1	GTP	-0.5649	0.0000	****
16	C8'	0.766505	-0.312212	0.914680	CT	1	GTP	-0.0062	0.0000	****
17	H8'1	0.372818	-1.315880	0.955700	H1	1	GTP	0.0699	0.0000	****
18	H8'2	0.557977	0.180680	1.851259	H1	1	GTP	0.0699	0.0000	****
19	C9'	-4.041595	-1.251223	1.610554	CT	1	GTP	0.1284	0.0000	****
20	H9'1	-4.210855	-0.477759	2.356256	H1	1	GTP	0.0497	0.0000	****
21	H9'2	-3.411778	-2.018791	2.048887	H1	1	GTP	0.0497	0.0000	****
22	O10'	-5.257488	-1.789157	1.163294	OH	1	GTP	-0.6690	0.0000	****
23	H10'	-5.716378	-2.193252	1.886156	HO	1	GTP	0.4382	0.0000	****
24	N1	2.199095	-0.369228	0.746125	N*	1	GTP	-0.0856	0.0000	****
25	C2	2.931817	0.717666	1.150562	C	1	GTP	0.5546	0.0000	****
26	O2	2.457802	1.640760	1.749217	O	1	GTP	-0.5620	0.0000	****
27	N3	4.262929	0.649281	0.840538	NA	1	GTP	-0.4513	0.0000	****
28	H3	4.813971	1.424838	1.142231	H	1	GTP	0.3319	0.0000	****
29	C4	4.918363	-0.329958	0.112682	C	1	GTP	0.5990	0.0000	****
30	O4	6.092091	-0.245355	-0.101713	O	1	GTP	-0.5614	0.0000	****
31	C5	4.055887	-1.425692	-0.339984	CM	1	GTP	0.0097	0.0000	****
32	C7	4.680287	-2.529774	-1.145733	CT	1	GTP	-0.2784	0.0000	****
33	H73	5.127731	-2.138991	-2.052623	HC	1	GTP	0.0921	0.0000	****
34	H72	5.468181	-3.019677	-0.584610	HC	1	GTP	0.0921	0.0000	****
35	H71	3.939973	-3.272938	-1.419789	HC	1	GTP	0.0921	0.0000	****
36	C6	2.767463	-1.371598	-0.014734	CM	1	GTP	-0.2376	0.0000	****
37	H6	2.077851	-2.136183	-0.317510	H4	1	GTP	0.2348	0.0000	****

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30	29	30	1
31	29	31	1

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@<TRIPOS>SUBSTRUCTURE

1 GTP

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

C' 1

@<TRIPOS>RESIDUECONNECT

1 N1' C' 0 0 0 0

GNT

@<TRIPOS>MOLECULE

GNT

39 39 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	0.000000	0.000000	-0.000000	N3	1	GNT	-0.0679	0.0000	****
2	H1'1	-0.372149	0.940540	-0.000000	H	1	GNT	0.2683	0.0000	****
3	H1'2	-0.372151	-0.470283	-0.814524	H	1	GNT	0.2683	0.0000	****
4	H1'3	-0.372144	-0.470264	0.814532	H	1	GNT	0.2683	0.0000	****
5	C2'	1.507376	0.000000	-0.000000	CT	1	GNT	0.3376	0.0000	****
6	H2'1	1.793921	0.507528	-0.911221	HP	1	GNT	0.0685	0.0000	****
7	C3'	2.143489	-1.399689	0.000000	CT	1	GNT	-0.0833	0.0000	****
8	H3'1	2.115032	-1.792411	0.999137	H1	1	GNT	0.0545	0.0000	****
9	H3'2	3.184252	-1.275903	-0.281027	H1	1	GNT	0.0545	0.0000	****
10	N4'	1.505180	-2.347630	-0.900712	N	1	GNT	-0.0870	0.0000	****
11	C5'	1.433524	-1.996072	-2.314401	CT	1	GNT	-0.1820	0.0000	****
12	H5'1	0.659949	-2.588207	-2.774198	H1	1	GNT	0.0902	0.0000	****
13	H5'2	1.180729	-0.953538	-2.412455	H1	1	GNT	0.0902	0.0000	****
14	C'	2.782154	-2.207860	-2.993884	C	1	GNT	0.6203	0.0000	****
15	O1'	3.562153	-1.296973	-3.110606	O	1	GNT	-0.5801	0.0000	****
16	C7'	1.215070	-3.631808	-0.572529	C	1	GNT	0.5146	0.0000	****
17	O7'	0.826576	-4.425786	-1.391590	O	1	GNT	-0.5620	0.0000	****
18	C8'	1.387935	-4.064328	0.884710	CT	1	GNT	0.0068	0.0000	****
19	H8'1	0.801018	-3.431954	1.532518	H1	1	GNT	0.0696	0.0000	****
20	H8'2	2.425903	-3.981197	1.166344	H1	1	GNT	0.0696	0.0000	****
21	C9'	2.072312	0.829105	1.148800	CT	1	GNT	0.0891	0.0000	****
22	H9'1	3.158967	0.803132	1.110399	H1	1	GNT	0.0461	0.0000	****
23	H9'2	1.752307	0.413373	2.098922	H1	1	GNT	0.0461	0.0000	****
24	O10'	1.597366	2.138730	0.984115	OH	1	GNT	-0.6783	0.0000	****
25	H10'	1.915230	2.687563	1.687072	HO	1	GNT	0.4459	0.0000	****
26	N1	0.982076	-5.437928	1.064923	N*	1	GNT	-0.0851	0.0000	****
27	C2	1.884033	-6.417234	0.734433	C	1	GNT	0.5580	0.0000	****
28	O2	3.020652	-6.186268	0.434550	O	1	GNT	-0.5585	0.0000	****
29	N3	1.387319	-7.690419	0.804634	NA	1	GNT	-0.4595	0.0000	****
30	H3	2.035369	-8.416452	0.583134	H	1	GNT	0.3331	0.0000	****
31	C4	0.089255	-8.083617	1.085166	C	1	GNT	0.6027	0.0000	****
32	O4	-0.208038	-9.242222	1.088865	O	1	GNT	-0.5614	0.0000	****
33	C5	-0.830644	-6.976704	1.364329	CM	1	GNT	0.0100	0.0000	****
34	C7	-2.263834	-7.309006	1.670345	CT	1	GNT	-0.2726	0.0000	****
35	H73	-2.719675	-7.844312	0.845103	HC	1	GNT	0.0912	0.0000	****
36	H72	-2.334551	-7.946642	2.544451	HC	1	GNT	0.0912	0.0000	****

37	H71	-2.837502	-6.407686	1.854711	HC	1	GNT	0.0912	0.0000	****
38	C6	-0.340205	-5.741202	1.322393	CM	1	GNT	-0.2382	0.0000	****
39	H6	-0.957644	-4.883753	1.510486	H4	1	GNT	0.2297	0.0000	****

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12	10	16	1
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@<TRIPOS>SUBSTRUCTURE

1 GNT

1 ****

0 **** *

@<TRIPOS>HEADTAIL

C' 1

0 0

@<TRIPOS>RESIDUECONNECT

1 C' 0 0 0 0 0

GCT

@<TRIPOS>MOLECULE

GCT

38 38 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	-2.529414	0.851945	-2.030472	N	1	GCT	-0.3796	0.0000	****
2	H1'1	-2.783661	0.026734	-2.524353	H	1	GCT	0.2738	0.0000	****
3	C2'	-1.243179	1.416752	-2.406977	CT	1	GCT	0.0724	0.0000	****
4	H2'1	-0.654219	0.579836	-2.757283	H1	1	GCT	0.1339	0.0000	****
5	C3'	-0.486204	2.098481	-1.255454	CT	1	GCT	-0.1382	0.0000	****
6	H3'1	-0.902032	3.076247	-1.099325	H1	1	GCT	0.0801	0.0000	****
7	H3'2	0.542276	2.220401	-1.579260	H1	1	GCT	0.0801	0.0000	****
8	N4'	-0.517724	1.363527	0.000000	N	1	GCT	-0.0476	0.0000	****
9	C5'	0.000000	0.000000	0.000000	CT	1	GCT	-0.5030	0.0000	****
10	H5'1	-0.397032	-0.515445	0.858566	H1	1	GCT	0.1684	0.0000	****
11	H5'2	-0.324673	-0.507018	-0.893262	H1	1	GCT	0.1684	0.0000	****
12	C7'	-0.742178	1.934413	1.210244	C	1	GCT	0.5677	0.0000	****
13	O7'	-0.610527	1.319502	2.238224	O	1	GCT	-0.5857	0.0000	****
14	C8'	-1.178554	3.400173	1.250564	CT	1	GCT	-0.0109	0.0000	****
15	H8'1	-2.074105	3.537887	0.664862	H1	1	GCT	0.0693	0.0000	****
16	H8'2	-0.397613	4.021800	0.841518	H1	1	GCT	0.0693	0.0000	****
17	C9'	-1.370592	2.375357	-3.586448	CT	1	GCT	0.1197	0.0000	****
18	H9'1	-0.388837	2.772112	-3.834861	H1	1	GCT	0.0523	0.0000	****
19	H9'2	-2.019227	3.204519	-3.322387	H1	1	GCT	0.0523	0.0000	****
20	O10'	-1.899140	1.641307	-4.658651	OH	1	GCT	-0.6738	0.0000	****
21	H10'	-2.007475	2.204190	-5.412196	HO	1	GCT	0.4408	0.0000	****
22	N1	-1.427023	3.825499	2.607522	N*	1	GCT	-0.0880	0.0000	****
23	C2	-0.346060	4.193181	3.367863	C	1	GCT	0.5577	0.0000	****
24	O2	0.760713	4.302682	2.922616	O	1	GCT	-0.5631	0.0000	****
25	N3	-0.639807	4.456591	4.678198	NA	1	GCT	-0.4533	0.0000	****
26	H3	0.132861	4.741569	5.242048	H	1	GCT	0.3320	0.0000	****
27	C4	-1.858207	4.317986	5.322066	C	1	GCT	0.5997	0.0000	****
28	O4	-1.961867	4.565562	6.487708	O	1	GCT	-0.5615	0.0000	****
29	C5	-2.949890	3.859175	4.457702	CM	1	GCT	0.0077	0.0000	****
30	C7	-4.307608	3.671292	5.073663	CT	1	GCT	-0.2831	0.0000	****
31	H73	-4.268690	2.944529	5.877199	HC	1	GCT	0.0935	0.0000	****
32	H72	-4.671083	4.600230	5.498634	HC	1	GCT	0.0935	0.0000	****
33	H71	-5.022291	3.328544	4.333922	HC	1	GCT	0.0935	0.0000	****
34	C6	-2.669053	3.629553	3.178189	CM	1	GCT	-0.2275	0.0000	****
35	H6	-3.418014	3.286665	2.490210	H4	1	GCT	0.2296	0.0000	****
36	C'	1.524911	-0.000000	0.000000	C	1	GCT	0.8129	0.0000	****
37	O1'	2.036591	-1.124430	-0.000000	O2	1	GCT	-0.8266	0.0000	****
38	OXT	2.065682	1.108429	0.000000	O2	1	GCT	-0.8266	0.0000	****

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12	9	11	1
13	9	36	1

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@<TRIPOS>SUBSTRUCTURE

1 GCT

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

0 0

@<TRIPOS>RESIDUECONNECT

1 N1' 0 0 0 0 0

MTP

@<TRIPOS>MOLECULE

MTP

49 49 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	C1	-2.294517	-3.685951	-1.627163	CT	1	MTP	-0.2117	0.0000	****
2	H1	-3.037204	-4.420831	-1.331685	HC	1	MTP	0.0663	0.0000	****
3	H2	-2.588060	-3.270281	-2.583803	HC	1	MTP	0.0663	0.0000	****
4	H4	-1.340176	-4.183090	-1.722361	HC	1	MTP	0.0663	0.0000	****
5	C	-2.194082	-2.628541	-0.549692	C	1	MTP	0.6963	0.0000	****
6	O	-1.363114	-2.692738	0.322836	O	1	MTP	-0.5971	0.0000	****
7	N1'	-3.098596	-1.629931	-0.637122	N	1	MTP	-0.5775	0.0000	****
8	H1'1	-3.847978	-1.763996	-1.277425	H	1	MTP	0.3483	0.0000	****
9	C2'	-3.349478	-0.638782	0.397210	CT	1	MTP	0.0634	0.0000	****
10	H2'1	-4.055355	0.056272	-0.037133	H1	1	MTP	0.1441	0.0000	****
11	C3'	-2.104015	0.144775	0.842889	CT	1	MTP	-0.1693	0.0000	****
12	H3'1	-1.537891	-0.463569	1.523137	H1	1	MTP	0.0998	0.0000	****
13	H3'2	-2.454304	1.016378	1.386112	H1	1	MTP	0.0998	0.0000	****
14	N4'	-1.239246	0.566306	-0.248807	N	1	MTP	-0.1318	0.0000	****
15	C5'	-1.823333	1.401828	-1.291874	CT	1	MTP	-0.1445	0.0000	****
16	H5'1	-1.199862	1.338949	-2.168107	H1	1	MTP	0.0645	0.0000	****
17	H5'2	-2.806879	1.036260	-1.535584	H1	1	MTP	0.0645	0.0000	****

18	C'	-1.973998	2.841207	-0.811415	C	1	MTP	0.6951	0.0000	****
19	O1'	-3.011624	3.225254	-0.334363	O	1	MTP	-0.6015	0.0000	****
20	C7'	0.113600	0.464041	-0.230592	C	1	MTP	0.5167	0.0000	****
21	O7'	0.800996	0.961161	-1.086483	O	1	MTP	-0.5280	0.0000	****
22	C8'	0.766505	-0.312212	0.914680	CT	1	MTP	-0.0520	0.0000	****
23	H8'1	0.372818	-1.315880	0.955700	H1	1	MTP	0.1003	0.0000	****
24	H8'2	0.557977	0.180680	1.851259	H1	1	MTP	0.1003	0.0000	****
25	C9'	-4.041595	-1.251222	1.610553	CT	1	MTP	0.1301	0.0000	****
26	H9'1	-4.210855	-0.477758	2.356256	H1	1	MTP	0.0590	0.0000	****
27	H9'2	-3.411778	-2.018791	2.048887	H1	1	MTP	0.0590	0.0000	****
28	O10'	-5.257488	-1.789156	1.163293	OH	1	MTP	-0.6713	0.0000	****
29	H10'	-5.716379	-2.193251	1.886156	HO	1	MTP	0.4373	0.0000	****
30	N1	2.199095	-0.369228	0.746125	N*	1	MTP	-0.0755	0.0000	****
31	C2	2.931817	0.717666	1.150562	C	1	MTP	0.5342	0.0000	****
32	O2	2.457803	1.640760	1.749217	O	1	MTP	-0.5545	0.0000	****
33	N3	4.262930	0.649281	0.840538	NA	1	MTP	-0.4337	0.0000	****
34	H3	4.813971	1.424838	1.142231	H	1	MTP	0.3287	0.0000	****
35	C4	4.918363	-0.329957	0.112682	C	1	MTP	0.5866	0.0000	****
36	O4	6.092091	-0.245355	-0.101714	O	1	MTP	-0.5593	0.0000	****
37	C5	4.055888	-1.425692	-0.339984	CM	1	MTP	0.0243	0.0000	****
38	C7	4.680287	-2.529774	-1.145733	CT	1	MTP	-0.2381	0.0000	****
39	H73	5.127731	-2.138991	-2.052623	HC	1	MTP	0.0821	0.0000	****
40	H72	5.468182	-3.019676	-0.584610	HC	1	MTP	0.0821	0.0000	****
41	H71	3.939973	-3.272938	-1.419789	HC	1	MTP	0.0821	0.0000	****
42	C6	2.767463	-1.371598	-0.014734	CM	1	MTP	-0.2703	0.0000	****
43	H6	2.077851	-2.136183	-0.317510	H4	1	MTP	0.2387	0.0000	****
44	N	-0.876798	3.606040	-0.936233	N	1	MTP	-0.5053	0.0000	****
45	H	-0.026485	3.156773	-1.197597	H	1	MTP	0.3271	0.0000	****
46	CH3	-0.819546	4.943626	-0.386512	CT	1	MTP	-0.0976	0.0000	****
47	HH31	-0.760908	4.933562	0.696658	H1	1	MTP	0.0853	0.0000	****
48	HH32	0.055735	5.442353	-0.779939	H1	1	MTP	0.0853	0.0000	****
49	HH33	-1.701278	5.495998	-0.677284	H1	1	MTP	0.0853	0.0000	****

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@<TRIPOS>SUBSTRUCTURE

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GCP

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GCP

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

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2	H1'1	2.788920	-2.419137	-0.055916	H	1	GCP	0.2784	0.0000	****
3	C2'	2.630417	-0.523116	-0.917345	CT	1	GCP	0.0528	0.0000	****
4	H2'1	3.388500	0.232783	-0.767886	H1	1	GCP	0.1363	0.0000	****
5	C3'	1.248910	0.134907	-1.024890	CT	1	GCP	-0.1234	0.0000	****
6	H3'1	0.504790	-0.599531	-1.287396	H1	1	GCP	0.0809	0.0000	****
7	H3'2	1.269210	0.861886	-1.828337	H1	1	GCP	0.0809	0.0000	****
8	N4'	0.830830	0.844382	0.178566	N	1	GCP	-0.1034	0.0000	****
9	C5'	1.388083	2.179326	0.377348	CT	1	GCP	-0.1424	0.0000	****
10	H5'1	1.407954	2.392945	1.433406	H1	1	GCP	0.0802	0.0000	****
11	H5'2	2.399076	2.188070	0.005038	H1	1	GCP	0.0802	0.0000	****
12	C'	0.595847	3.228539	-0.395664	C	1	GCP	0.6199	0.0000	****
13	O1'	0.943714	3.590385	-1.490504	O	1	GCP	-0.5806	0.0000	****
14	C7'	-0.107239	0.425715	1.042028	C	1	GCP	0.5312	0.0000	****
15	O7'	-0.583920	1.152502	1.889173	O	1	GCP	-0.5649	0.0000	****
16	C8'	-0.596562	-1.021874	1.014113	CT	1	GCP	-0.1542	0.0000	****
17	H8'1	-0.531795	-1.362822	2.039178	H1	1	GCP	0.0926	0.0000	****
18	H8'2	-0.016645	-1.675714	0.396835	H1	1	GCP	0.0926	0.0000	****
19	C9'	2.930310	-1.251338	-2.222117	CT	1	GCP	0.1284	0.0000	****
20	H9'1	2.924591	-0.531221	-3.035294	H1	1	GCP	0.0497	0.0000	****
21	H9'2	2.152720	-1.986351	-2.424248	H1	1	GCP	0.0497	0.0000	****
22	O10'	4.181628	-1.873440	-2.105175	OH	1	GCP	-0.6690	0.0000	****

23	H10'	4.406673	-2.291285	-2.924595	HO	1	GCP	0.4382	0.0000	****
24	N1	-1.984179	-1.106290	0.576026	N*	1	GCP	-0.0388	0.0000	****
25	C2	-2.267068	-1.655975	-0.689648	C	1	GCP	0.7490	0.0000	****
26	O2	-1.351038	-2.022043	-1.380657	O	1	GCP	-0.5921	0.0000	****
27	N3	-3.567503	-1.740644	-1.060912	NC	1	GCP	-0.7700	0.0000	****
28	C4	-4.512582	-1.323108	-0.274271	CA	1	GCP	0.8543	0.0000	****
29	N4	-5.780838	-1.459083	-0.697812	N2	1	GCP	-0.9738	0.0000	****
30	H41	-6.533134	-1.019636	-0.222212	H	1	GCP	0.4235	0.0000	****
31	H42	-5.926363	-1.766910	-1.632749	H	1	GCP	0.4235	0.0000	****
32	C5	-4.265886	-0.749157	1.019776	CM	1	GCP	-0.5173	0.0000	****
33	H5	-5.058468	-0.412818	1.657783	HA	1	GCP	0.2029	0.0000	****
34	C6	-2.980078	-0.667278	1.385305	CM	1	GCP	-0.0185	0.0000	****
35	H6	-2.667692	-0.247377	2.320001	H4	1	GCP	0.1899	0.0000	****

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@<TRIPOS>SUBSTRUCTURE

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C' 1

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GNC

@<TRIPOS>MOLECULE

GNC

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

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2	H1'1	-0.372149	0.940540	-0.000000	H	1	GNC	0.2683	0.0000	****
3	H1'2	-0.372151	-0.470283	-0.814524	H	1	GNC	0.2683	0.0000	****
4	H1'3	-0.372144	-0.470264	0.814532	H	1	GNC	0.2683	0.0000	****
5	C2'	1.507376	-0.000000	0.000000	CT	1	GNC	0.3376	0.0000	****
6	H2'1	1.833250	0.534506	-0.881204	HP	1	GNC	0.0685	0.0000	****
7	C3'	2.094850	-1.417038	0.000000	CT	1	GNC	-0.0833	0.0000	****
8	H3'1	1.868310	-1.915294	0.928678	H1	1	GNC	0.0545	0.0000	****
9	H3'2	3.174494	-1.348210	-0.063790	H1	1	GNC	0.0545	0.0000	****
10	N4'	1.655874	-2.244745	-1.117431	N	1	GNC	-0.0870	0.0000	****
11	C5'	2.327958	-2.027648	-2.395434	CT	1	GNC	-0.1820	0.0000	****
12	H5'1	1.656952	-2.305509	-3.191569	H1	1	GNC	0.0902	0.0000	****
13	H5'2	2.557570	-0.979129	-2.488622	H1	1	GNC	0.0902	0.0000	****
14	C'	3.639927	-2.801990	-2.467311	C	1	GNC	0.6203	0.0000	****
15	O1'	4.688139	-2.279399	-2.186654	O	1	GNC	-0.5801	0.0000	****
16	C7'	0.783308	-3.261095	-1.036904	C	1	GNC	0.5146	0.0000	****
17	O7'	0.631240	-4.062069	-1.936023	O	1	GNC	-0.5620	0.0000	****
18	C8'	-0.095758	-3.432499	0.201480	CT	1	GNC	-0.1478	0.0000	****
19	H8'1	-1.101927	-3.550372	-0.179215	H1	1	GNC	0.1032	0.0000	****
20	H8'2	-0.078678	-2.603269	0.877445	H1	1	GNC	0.1032	0.0000	****
21	C9'	2.018809	0.743539	1.228112	CT	1	GNC	0.0891	0.0000	****
22	H9'1	3.103039	0.791350	1.183189	H1	1	GNC	0.0461	0.0000	****
23	H9'2	1.747766	0.199052	2.131337	H1	1	GNC	0.0461	0.0000	****
24	O10'	1.455016	2.027503	1.235743	OH	1	GNC	-0.6783	0.0000	****
25	H10'	1.799371	2.521511	1.966541	HO	1	GNC	0.4459	0.0000	****
26	N1	0.267856	-4.628625	0.950876	N*	1	GNC	-0.0588	0.0000	****
27	C2	0.898955	-4.483583	2.201791	C	1	GNC	0.7677	0.0000	****
28	O2	1.137076	-3.374256	2.605897	O	1	GNC	-0.6116	0.0000	****
29	N3	1.206124	-5.612925	2.884705	NC	1	GNC	-0.7639	0.0000	****
30	C4	0.929717	-6.783087	2.394161	CA	1	GNC	0.8431	0.0000	****
31	N4	1.241958	-7.859097	3.136494	N2	1	GNC	-0.9719	0.0000	****
32	H41	1.205773	-8.775500	2.756818	H	1	GNC	0.4235	0.0000	****
33	H42	1.767425	-7.711138	3.968380	H	1	GNC	0.4235	0.0000	****
34	C5	0.294687	-6.974159	1.119381	CM	1	GNC	-0.5099	0.0000	****
35	H5	0.070586	-7.947538	0.731212	HA	1	GNC	0.2007	0.0000	****
36	C6	-0.009016	-5.856781	0.446441	CM	1	GNC	-0.0133	0.0000	****
37	H6	-0.472308	-5.868348	-0.519366	H4	1	GNC	0.1904	0.0000	****

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@<TRIPOS>SUBSTRUCTURE

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0 **** ****

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C' 1

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GCC

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GCC

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	-3.099827	1.846246	-1.417676	N	1	GCC	-0.3796	0.0000	****
2	H1'1	-3.671574	2.574750	-1.784973	H	1	GCC	0.2738	0.0000	****
3	C2'	-1.848261	1.613523	-2.122096	CT	1	GCC	0.0724	0.0000	****
4	H2'1	-1.797784	0.559705	-2.357321	H1	1	GCC	0.1339	0.0000	****
5	C3'	-0.623438	2.024668	-1.295119	CT	1	GCC	-0.1382	0.0000	****
6	H3'1	-0.609107	3.093376	-1.154832	H1	1	GCC	0.0801	0.0000	****
7	H3'2	0.273367	1.768719	-1.847087	H1	1	GCC	0.0801	0.0000	****
8	N4'	-0.528153	1.361314	0.000000	N	1	GCC	-0.0476	0.0000	****
9	C5'	0.000000	-0.000000	0.000000	CT	1	GCC	-0.5030	0.0000	****
10	H5'1	-0.398625	-0.526479	0.851590	H1	1	GCC	0.1684	0.0000	****
11	H5'2	-0.330444	-0.496262	-0.897402	H1	1	GCC	0.1684	0.0000	****
12	C7'	-0.766535	1.937298	1.188376	C	1	GCC	0.5677	0.0000	****
13	O7'	-0.448304	1.419482	2.238945	O	1	GCC	-0.5857	0.0000	****
14	C8'	-1.494072	3.278962	1.268118	CT	1	GCC	-0.1640	0.0000	****
15	H8'1	-2.281823	3.131485	1.995372	H1	1	GCC	0.0932	0.0000	****
16	H8'2	-1.932254	3.602882	0.347295	H1	1	GCC	0.0932	0.0000	****
17	C9'	-1.843699	2.397174	-3.429208	CT	1	GCC	0.1197	0.0000	****
18	H9'1	-0.933167	2.165351	-3.974230	H1	1	GCC	0.0523	0.0000	****
19	H9'2	-1.842978	3.466050	-3.221242	H1	1	GCC	0.0523	0.0000	****
20	O10'	-2.980946	2.036702	-4.166237	OH	1	GCC	-0.6738	0.0000	****

21	H10'	-2.969980	2.478239	-5.003861	HO	1	GCC	0.4408	0.0000	****
22	N1	-0.609300	4.337001	1.739561	N*	1	GCC	-0.0389	0.0000	****
23	C2	-0.199002	5.335840	0.835079	C	1	GCC	0.7445	0.0000	****
24	O2	-0.576436	5.274309	-0.307010	O	1	GCC	-0.5879	0.0000	****
25	N3	0.606440	6.317903	1.307256	NC	1	GCC	-0.7716	0.0000	****
26	C4	0.985900	6.327676	2.549121	CA	1	GCC	0.8616	0.0000	****
27	N4	1.765827	7.344614	2.953967	N2	1	GCC	-0.9778	0.0000	****
28	H41	2.217869	7.327563	3.837497	H	1	GCC	0.4244	0.0000	****
29	H42	2.103523	7.973587	2.260860	H	1	GCC	0.4244	0.0000	****
30	C5	0.596715	5.323905	3.500865	CM	1	GCC	-0.5198	0.0000	****
31	H5	0.916436	5.349404	4.523358	HA	1	GCC	0.2025	0.0000	****
32	C6	-0.200141	4.354734	3.032524	CM	1	GCC	-0.0194	0.0000	****
33	H6	-0.547291	3.543921	3.640494	H4	1	GCC	0.1940	0.0000	****
34	C'	1.525134	0.000000	-0.000000	C	1	GCC	0.8129	0.0000	****
35	O1'	2.036814	-1.124430	-0.000000	O2	1	GCC	-0.8266	0.0000	****
36	OXT	2.065904	1.108429	-0.000000	O2	1	GCC	-0.8266	0.0000	****

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MCP

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MCP

47 47 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

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2	H1	2.926850	-3.040248	2.254910	HC	1	MCP	0.0460	0.0000	****
3	H2	4.621422	-2.690784	1.946091	HC	1	MCP	0.0460	0.0000	****
4	H3	3.800560	-1.891206	3.281362	HC	1	MCP	0.0460	0.0000	****
5	C	3.323224	-1.104279	1.363367	C	1	MCP	0.5575	0.0000	****
6	O	3.558932	0.034592	1.656503	O	1	MCP	-0.5770	0.0000	****
7	N1'	2.717500	-1.457508	0.194461	N	1	MCP	-0.3361	0.0000	****
8	H1'1	2.788920	-2.419137	-0.055916	H	1	MCP	0.2690	0.0000	****
9	C2'	2.630417	-0.523116	-0.917345	CT	1	MCP	0.1463	0.0000	****
10	H2'1	3.388500	0.232783	-0.767886	H1	1	MCP	0.1139	0.0000	****
11	C3'	1.248910	0.134907	-1.024890	CT	1	MCP	-0.1738	0.0000	****
12	H3'1	0.504790	-0.599531	-1.287396	H1	1	MCP	0.0712	0.0000	****
13	H3'2	1.269210	0.861886	-1.828337	H1	1	MCP	0.0712	0.0000	****
14	N4'	0.830830	0.844382	0.178566	N	1	MCP	0.0073	0.0000	****
15	C5'	1.388083	2.179326	0.377348	CT	1	MCP	-0.0141	0.0000	****
16	H5'1	1.407954	2.392945	1.433406	H1	1	MCP	0.0549	0.0000	****
17	H5'2	2.399076	2.188070	0.005038	H1	1	MCP	0.0549	0.0000	****
18	C'	0.595847	3.228539	-0.395664	C	1	MCP	0.5472	0.0000	****
19	O1'	0.943714	3.590385	-1.490504	O	1	MCP	-0.5600	0.0000	****
20	C7'	-0.107239	0.425715	1.042028	C	1	MCP	0.4267	0.0000	****
21	O7'	-0.583920	1.152502	1.889173	O	1	MCP	-0.5674	0.0000	****
22	C8'	-0.596562	-1.021874	1.014113	CT	1	MCP	-0.1183	0.0000	****
23	H8'1	-0.531795	-1.362822	2.039178	H1	1	MCP	0.0559	0.0000	****
24	H8'2	-0.016645	-1.675714	0.396835	H1	1	MCP	0.0559	0.0000	****
25	C9'	2.930310	-1.251338	-2.222117	CT	1	MCP	0.0338	0.0000	****
26	H9'1	2.924591	-0.531221	-3.035294	H1	1	MCP	0.0785	0.0000	****
27	H9'2	2.152720	-1.986351	-2.424248	H1	1	MCP	0.0785	0.0000	****
28	O10'	4.181628	-1.873440	-2.105175	OH	1	MCP	-0.6968	0.0000	****
29	H10'	4.406673	-2.291285	-2.924595	HO	1	MCP	0.4539	0.0000	****
30	N1	-1.984179	-1.106290	0.576026	N*	1	MCP	-0.0010	0.0000	****
31	C2	-2.267068	-1.655975	-0.689648	C	1	MCP	0.7905	0.0000	****
32	O2	-1.351038	-2.022043	-1.380657	O	1	MCP	-0.6190	0.0000	****
33	N3	-3.567503	-1.740644	-1.060912	NC	1	MCP	-0.7781	0.0000	****
34	C4	-4.512582	-1.323108	-0.274271	CA	1	MCP	0.8462	0.0000	****
35	N4	-5.780838	-1.459083	-0.697812	N2	1	MCP	-0.9721	0.0000	****
36	H41	-6.533134	-1.019636	-0.222212	H	1	MCP	0.4244	0.0000	****
37	H42	-5.926363	-1.766910	-1.632749	H	1	MCP	0.4244	0.0000	****
38	C5	-4.265886	-0.749157	1.019776	CM	1	MCP	-0.5185	0.0000	****
39	H5	-5.058468	-0.412818	1.657783	HA	1	MCP	0.2030	0.0000	****
40	C6	-2.980078	-0.667278	1.385305	CM	1	MCP	-0.0348	0.0000	****
41	H6	-2.667692	-0.247377	2.320001	H4	1	MCP	0.2055	0.0000	****
42	N	-0.501691	3.694204	0.227672	N	1	MCP	-0.4405	0.0000	****
43	H	-0.796491	3.215623	1.050367	H	1	MCP	0.2932	0.0000	****
44	CH3	-1.416689	4.596588	-0.433545	CT	1	MCP	-0.1192	0.0000	****
45	HH31	-1.983923	4.101447	-1.215571	H1	1	MCP	0.0853	0.0000	****
46	HH32	-2.104773	4.994056	0.301041	H1	1	MCP	0.0853	0.0000	****
47	HH33	-0.868289	5.414008	-0.879122	H1	1	MCP	0.0853	0.0000	****

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GGP

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GGP

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	3.885223	-0.880910	-0.808808	N	1	GGP	-0.3871	0.0000	****
2	H1'1	4.692324	-0.821494	-1.390197	H	1	GGP	0.2784	0.0000	****
3	C2'	3.269966	0.403563	-0.501697	CT	1	GGP	0.0528	0.0000	****
4	H2'1	3.627542	0.743034	0.463926	H1	1	GGP	0.1363	0.0000	****
5	C3'	1.722821	0.356073	-0.484518	CT	1	GGP	-0.1234	0.0000	****
6	H3'1	1.388455	-0.566907	-0.917081	H1	1	GGP	0.0809	0.0000	****
7	H3'2	1.327562	1.143279	-1.113853	H1	1	GGP	0.0809	0.0000	****
8	N4'	1.148909	0.539995	0.839740	N	1	GGP	-0.1034	0.0000	****
9	C5'	1.193173	1.896219	1.365522	CT	1	GGP	-0.1424	0.0000	****
10	H5'1	1.269349	1.861910	2.440148	H1	1	GGP	0.0802	0.0000	****
11	H5'2	2.066925	2.395857	0.973812	H1	1	GGP	0.0802	0.0000	****
12	C'	-0.022711	2.704404	0.913884	C	1	GGP	0.6199	0.0000	****
13	O1'	-0.060654	3.200901	-0.191064	O	1	GGP	-0.5806	0.0000	****
14	C7'	0.331682	-0.315533	1.497025	C	1	GGP	0.5312	0.0000	****
15	O7'	-0.229491	0.010256	2.514325	O	1	GGP	-0.5649	0.0000	****
16	C8'	0.084256	-1.731059	0.973989	CT	1	GGP	-0.3588	0.0000	****
17	H8'1	0.323946	-2.395413	1.790117	H1	1	GGP	0.1686	0.0000	****
18	H8'2	0.695268	-2.010694	0.137551	H1	1	GGP	0.1686	0.0000	****
19	C9'	3.729343	1.392596	-1.566309	CT	1	GGP	0.1284	0.0000	****
20	H9'1	3.365225	2.382666	-1.309302	H1	1	GGP	0.0497	0.0000	****
21	H9'2	3.300096	1.116687	-2.527664	H1	1	GGP	0.0497	0.0000	****
22	O10'	5.131334	1.367779	-1.625421	OH	1	GGP	-0.6690	0.0000	****
23	H10'	5.437384	1.965740	-2.292987	HO	1	GGP	0.4382	0.0000	****
24	N9	-1.318472	-1.899626	0.647736	N*	1	GGP	0.0925	0.0000	****
25	C8	-2.151295	-2.929493	1.019778	CK	1	GGP	0.0810	0.0000	****
26	H8	-1.810756	-3.699205	1.681946	H5	1	GGP	0.1655	0.0000	****
27	N7	-3.319112	-2.853317	0.504598	NB	1	GGP	-0.5385	0.0000	****
28	C6	-4.289069	-1.079564	-1.071869	C	1	GGP	0.4329	0.0000	****
29	O6	-5.412616	-1.396765	-1.323070	O	1	GGP	-0.5353	0.0000	****
30	C5	-3.286072	-1.707024	-0.258674	CB	1	GGP	0.2010	0.0000	****
31	C4	-2.056538	-1.116212	-0.172703	CB	1	GGP	0.0655	0.0000	****
32	N3	-1.620990	0.032575	-0.741352	NC	1	GGP	-0.3538	0.0000	****
33	C2	-2.523556	0.622336	-1.451609	CA	1	GGP	0.4952	0.0000	****
34	N2	-2.229830	1.791259	-2.076673	N2	1	GGP	-0.9092	0.0000	****
35	H21	-1.438697	2.273128	-1.694739	H	1	GGP	0.4099	0.0000	****
36	H22	-2.988062	2.385710	-2.327274	H	1	GGP	0.4099	0.0000	****
37	N1	-3.772306	0.116500	-1.633233	NA	1	GGP	-0.3450	0.0000	****
38	H1	-4.414685	0.588749	-2.232978	H	1	GGP	0.3137	0.0000	****

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	0.000000	0.000000	-0.000000	N3	1	GNG	-0.0679	0.0000	****
2	H1'1	-0.372149	0.940540	-0.000000	H	1	GNG	0.2683	0.0000	****
3	H1'2	-0.372151	-0.470283	-0.814524	H	1	GNG	0.2683	0.0000	****
4	H1'3	-0.372144	-0.470264	0.814532	H	1	GNG	0.2683	0.0000	****
5	C2'	1.507376	-0.000000	0.000000	CT	1	GNG	0.3376	0.0000	****
6	H2'1	1.859200	0.541465	-0.870956	HP	1	GNG	0.0685	0.0000	****
7	C3'	2.122471	-1.420516	-0.000000	CT	1	GNG	-0.0833	0.0000	****
8	H3'1	1.358780	-2.140998	0.220192	H1	1	GNG	0.0545	0.0000	****
9	H3'2	2.850738	-1.501736	0.796879	H1	1	GNG	0.0545	0.0000	****
10	N4'	2.806114	-1.759431	-1.238802	N	1	GNG	-0.0870	0.0000	****
11	C5'	4.093915	-1.114611	-1.447430	CT	1	GNG	-0.1820	0.0000	****
12	H5'1	4.258019	-0.974729	-2.503510	H1	1	GNG	0.0902	0.0000	****
13	H5'2	4.082859	-0.146317	-0.969098	H1	1	GNG	0.0902	0.0000	****
14	C'	5.224675	-1.916774	-0.804444	C	1	GNG	0.6203	0.0000	****
15	O1'	5.445504	-1.832517	0.384251	O	1	GNG	-0.5801	0.0000	****
16	C7'	2.535524	-2.803226	-2.056778	C	1	GNG	0.5146	0.0000	****

17	O7'	3.274157	-3.096001	-2.964885	O	1	GNG	-0.5620	0.0000	****
18	C8'	1.281814	-3.656573	-1.860538	CT	1	GNG	-0.3590	0.0000	****
19	H8'1	0.766924	-3.650548	-2.809082	H1	1	GNG	0.1808	0.0000	****
20	H8'2	0.600949	-3.285147	-1.119171	H1	1	GNG	0.1808	0.0000	****
21	C9'	1.960922	0.742656	1.251148	CT	1	GNG	0.0891	0.0000	****
22	H9'1	3.041717	0.844064	1.229432	H1	1	GNG	0.0461	0.0000	****
23	H9'2	1.696300	0.162963	2.133482	H1	1	GNG	0.0461	0.0000	****
24	O10'	1.334538	1.998280	1.278344	OH	1	GNG	-0.6783	0.0000	****
25	H10'	1.591751	2.471051	2.057572	HO	1	GNG	0.4459	0.0000	****
26	N9	1.656789	-5.023667	-1.555662	N*	1	GNG	0.0604	0.0000	****
27	C8	1.178959	-6.176564	-2.134578	CK	1	GNG	0.0966	0.0000	****
28	H8	0.496144	-6.135073	-2.958551	H5	1	GNG	0.1596	0.0000	****
29	N7	1.630678	-7.244108	-1.594848	NB	1	GNG	-0.5410	0.0000	****
30	C6	3.271740	-7.511572	0.356470	C	1	GNG	0.4206	0.0000	****
31	O6	3.413603	-8.681415	0.549914	O	1	GNG	-0.5339	0.0000	****
32	C5	2.466422	-6.798686	-0.594451	CB	1	GNG	0.2116	0.0000	****
33	C4	2.486193	-5.432275	-0.567367	CB	1	GNG	0.0660	0.0000	****
34	N3	3.195186	-4.605619	0.236837	NC	1	GNG	-0.3192	0.0000	****
35	C2	3.946556	-5.234987	1.077222	CA	1	GNG	0.4407	0.0000	****
36	N2	4.721299	-4.532969	1.943424	N2	1	GNG	-0.8765	0.0000	****
37	H21	4.892542	-3.585672	1.665455	H	1	GNG	0.4011	0.0000	****
38	H22	5.512743	-4.993186	2.334455	H	1	GNG	0.4011	0.0000	****
39	N1	3.989653	-6.591107	1.162750	NA	1	GNG	-0.3243	0.0000	****
40	H1	4.550843	-7.024708	1.864476	H	1	GNG	0.3125	0.0000	****

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1 GNG

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C' 1

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GCG

@<TRIPOS>MOLECULE

GCG

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SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	N1'	-2.967890	2.379092	-2.284478	N	1	GCG	-0.3796	0.0000	****
2	H1'1	-3.406790	2.348313	-3.178555	H	1	GCG	0.2738	0.0000	****
3	C2'	-1.889871	1.414513	-2.110720	CT	1	GCG	0.0724	0.0000	****
4	H2'1	-2.280208	0.537709	-1.606357	H1	1	GCG	0.1339	0.0000	****
5	C3'	-0.689136	1.971916	-1.308382	CT	1	GCG	-0.1382	0.0000	****
6	H3'1	-0.783380	3.036598	-1.216876	H1	1	GCG	0.0801	0.0000	****
7	H3'2	0.227624	1.797618	-1.857158	H1	1	GCG	0.0801	0.0000	****
8	N4'	-0.526617	1.356622	-0.000000	N	1	GCG	-0.0476	0.0000	****
9	C5'	0.000000	-0.000000	0.000000	CT	1	GCG	-0.5030	0.0000	****
10	H5'1	-0.396333	-0.538526	0.845406	H1	1	GCG	0.1684	0.0000	****
11	H5'2	-0.315180	-0.498534	-0.904789	H1	1	GCG	0.1684	0.0000	****
12	C7'	-0.523101	1.985187	1.198627	C	1	GCG	0.5677	0.0000	****
13	O7'	-0.204978	1.407021	2.208813	O	1	GCG	-0.5857	0.0000	****
14	C8'	-0.920253	3.456914	1.320235	CT	1	GCG	-0.3124	0.0000	****
15	H8'1	-1.703475	3.492915	2.061955	H1	1	GCG	0.1489	0.0000	****
16	H8'2	-1.307070	3.890551	0.418297	H1	1	GCG	0.1489	0.0000	****
17	C9'	-1.417699	1.006679	-3.501165	CT	1	GCG	0.1197	0.0000	****
18	H9'1	-0.680373	0.215390	-3.405851	H1	1	GCG	0.0523	0.0000	****
19	H9'2	-0.937988	1.855318	-3.985181	H1	1	GCG	0.0523	0.0000	****
20	O10'	-2.528794	0.577347	-4.243345	OH	1	GCG	-0.6738	0.0000	****
21	H10'	-2.258784	0.333116	-5.117595	HO	1	GCG	0.4408	0.0000	****
22	N9	0.203046	4.233690	1.807389	N*	1	GCG	0.0894	0.0000	****
23	C8	0.211072	5.149352	2.834099	CK	1	GCG	0.0757	0.0000	****
24	H8	-0.662604	5.311947	3.431714	H5	1	GCG	0.1679	0.0000	****
25	N7	1.332735	5.746379	2.977069	NB	1	GCG	-0.5384	0.0000	****
26	C6	3.508356	5.460320	1.651509	C	1	GCG	0.4327	0.0000	****
27	O6	4.308753	6.222144	2.104350	O	1	GCG	-0.5358	0.0000	****
28	C5	2.138216	5.207838	1.997867	CB	1	GCG	0.2036	0.0000	****
29	C4	1.447019	4.275452	1.275989	CB	1	GCG	0.0617	0.0000	****
30	N3	1.876062	3.499727	0.252783	NC	1	GCG	-0.3583	0.0000	****
31	C2	3.115941	3.696160	-0.048918	CA	1	GCG	0.4921	0.0000	****
32	N2	3.685140	2.981204	-1.053015	N2	1	GCG	-0.9048	0.0000	****
33	H21	3.197662	2.136408	-1.282466	H	1	GCG	0.4085	0.0000	****
34	H22	4.676823	2.893746	-1.054811	H	1	GCG	0.4085	0.0000	****
35	N1	3.895632	4.615646	0.579462	NA	1	GCG	-0.3428	0.0000	****
36	H1	4.833699	4.761078	0.272470	H	1	GCG	0.3130	0.0000	****
37	C'	1.528239	0.000000	-0.000000	C	1	GCG	0.8129	0.0000	****
38	O1'	2.039919	-1.124430	-0.000000	O2	1	GCG	-0.8266	0.0000	****

39 OXT 2.069009 1.108429 0.000000 O2 1 GCG -0.8266 0.0000 ****

@<TRIPOS>BOND

1	1	2	1
2	1	3	1
3	3	4	1
4	3	5	1
5	3	17	1
6	5	6	1
7	5	7	1
8	5	8	1
9	8	9	1
10	8	12	1
11	9	10	1
12	9	11	1
13	9	37	1
14	12	13	1
15	12	14	1
16	14	15	1
17	14	16	1
18	14	22	1
19	17	18	1
20	17	19	1
21	17	20	1
22	20	21	1
23	22	23	1
24	22	29	1
25	23	24	1
26	23	25	1
27	25	28	1
28	26	27	1
29	26	28	1
30	26	35	1
31	28	29	1
32	29	30	1
33	30	31	1
34	31	32	1
35	31	35	1
36	32	33	1
37	32	34	1
38	35	36	1
39	37	38	1
40	37	39	1

@<TRIPOS>SUBSTRUCTURE

1 GCG

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

N1' 1

0 0

@<TRIPOS>RESIDUECONNECT

1 N1' 0 0 0 0 0

MGP

@<TRIPOS>MOLECULE

MGP

50 51 1 0 1

SMALL

USER_CHARGES

@<TRIPOS>ATOM

1	C1	4.797332	-3.055821	-0.240181	CT	1	MGP	-0.2105	0.0000	****
2	H2	4.398571	-3.973878	0.166687	HC	1	MGP	0.0680	0.0000	****
3	H3	4.983696	-3.174554	-1.301012	HC	1	MGP	0.0680	0.0000	****
4	H4	5.742392	-2.845956	0.251674	HC	1	MGP	0.0680	0.0000	****
5	C	3.832568	-1.927072	0.049755	C	1	MGP	0.6718	0.0000	****
6	O	3.073211	-1.962992	0.982695	O	1	MGP	-0.6062	0.0000	****
7	N1'	3.885223	-0.880910	-0.808808	N	1	MGP	-0.5466	0.0000	****
8	H1'1	4.692324	-0.821494	-1.390197	H	1	MGP	0.3188	0.0000	****
9	C2'	3.269966	0.403563	-0.501697	CT	1	MGP	0.0879	0.0000	****
10	H2'1	3.627542	0.743034	0.463926	H1	1	MGP	0.1240	0.0000	****
11	C3'	1.722821	0.356073	-0.484518	CT	1	MGP	-0.0561	0.0000	****
12	H3'1	1.388455	-0.566907	-0.917081	H1	1	MGP	0.0476	0.0000	****
13	H3'2	1.327562	1.143279	-1.113853	H1	1	MGP	0.0476	0.0000	****
14	N4'	1.148909	0.539995	0.839740	N	1	MGP	-0.1749	0.0000	****
15	C5'	1.193173	1.896219	1.365522	CT	1	MGP	-0.1384	0.0000	****
16	H5'1	1.269349	1.861910	2.440148	H1	1	MGP	0.0884	0.0000	****
17	H5'2	2.066925	2.395857	0.973812	H1	1	MGP	0.0884	0.0000	****
18	C'	-0.022711	2.704404	0.913884	C	1	MGP	0.5906	0.0000	****
19	O1'	-0.060654	3.200901	-0.191064	O	1	MGP	-0.5690	0.0000	****
20	C7'	0.331682	-0.315533	1.497025	C	1	MGP	0.5935	0.0000	****
21	O7'	-0.229491	0.010256	2.514325	O	1	MGP	-0.5839	0.0000	****
22	C8'	0.084256	-1.731059	0.973989	CT	1	MGP	-0.1737	0.0000	****
23	H8'1	0.323946	-2.395413	1.790117	H1	1	MGP	0.1426	0.0000	****
24	H8'2	0.695268	-2.010694	0.137551	H1	1	MGP	0.1426	0.0000	****
25	C9'	3.729343	1.392596	-1.566309	CT	1	MGP	0.1164	0.0000	****
26	H9'1	3.365225	2.382666	-1.309302	H1	1	MGP	0.0568	0.0000	****
27	H9'2	3.300096	1.116687	-2.527664	H1	1	MGP	0.0568	0.0000	****
28	O10'	5.131334	1.367779	-1.625421	OH	1	MGP	-0.6703	0.0000	****
29	H10'	5.437384	1.965740	-2.292987	HO	1	MGP	0.4425	0.0000	****
30	N9	-1.318472	-1.899626	0.647736	N*	1	MGP	-0.0583	0.0000	****
31	C8	-2.151295	-2.929493	1.019778	CK	1	MGP	0.1204	0.0000	****
32	H8	-1.810756	-3.699205	1.681946	H5	1	MGP	0.1610	0.0000	****
33	N7	-3.319112	-2.853317	0.504598	NB	1	MGP	-0.5544	0.0000	****
34	C6	-4.289069	-1.079564	-1.071869	C	1	MGP	0.3876	0.0000	****
35	O6	-5.412616	-1.396765	-1.323070	O	1	MGP	-0.5301	0.0000	****
36	C5	-3.286072	-1.707024	-0.258674	CB	1	MGP	0.2799	0.0000	****
37	C4	-2.056538	-1.116212	-0.172703	CB	1	MGP	-0.0067	0.0000	****
38	N3	-1.620990	0.032575	-0.741352	NC	1	MGP	-0.2318	0.0000	****
39	C2	-2.523556	0.622336	-1.451609	CA	1	MGP	0.3686	0.0000	****
40	N2	-2.229830	1.791259	-2.076673	N2	1	MGP	-0.8561	0.0000	****
41	H21	-1.438697	2.273128	-1.694739	H	1	MGP	0.4002	0.0000	****
42	H22	-2.988062	2.385710	-2.327274	H	1	MGP	0.4002	0.0000	****
43	N1	-3.772306	0.116500	-1.633233	NA	1	MGP	-0.2892	0.0000	****
44	H1	-4.414685	0.588749	-2.232978	H	1	MGP	0.3076	0.0000	****
45	N	-0.997952	2.830308	1.823108	N	1	MGP	-0.5123	0.0000	****
46	H	-0.983172	2.174325	2.574093	H	1	MGP	0.3597	0.0000	****
47	CH3	-2.250717	3.484390	1.517669	CT	1	MGP	-0.0655	0.0000	****
48	HH31	-2.913524	2.846151	0.942320	H1	1	MGP	0.0763	0.0000	****
49	HH32	-2.742007	3.747994	2.444593	H1	1	MGP	0.0763	0.0000	****
50	HH33	-2.057496	4.385125	0.954315	H1	1	MGP	0.0763	0.0000	****

@<TRIPOS>BOND

1	1	2	1
2	1	3	1
3	1	4	1
4	1	5	1
5	5	6	1
6	5	7	1
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8	7	9	1
9	9	10	1
10	9	11	1
11	9	25	1

12	11	12	1
13	11	13	1
14	11	14	1
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16	14	20	1
17	15	16	1
18	15	17	1
19	15	18	1
20	18	19	1
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23	20	22	1
24	22	23	1
25	22	24	1
26	22	30	1
27	25	26	1
28	25	27	1
29	25	28	1
30	28	29	1
31	30	31	1
32	30	37	1
33	31	32	1
34	31	33	1
35	33	36	1
36	34	35	1
37	34	36	1
38	34	43	1
39	36	37	1
40	37	38	1
41	38	39	1
42	39	40	1
43	39	43	1
44	40	41	1
45	40	42	1
46	43	44	1
47	45	46	1
48	45	47	1
49	47	48	1
50	47	49	1
51	47	50	1

@<TRIPOS>SUBSTRUCTURE

1 MGP

1 ****

0 **** ****

@<TRIPOS>HEADTAIL

0 0

0 0

@<TRIPOS>RESIDUECONNECT

1 0 0 0 0 0 0

frcmod.pna

PNA frcmod file: frcmod.pna
MASS

BOND

CQ-NA 502.0 1.324 by analogy to CQ-NC, parm99.dat

ANGL

C -CT-N* 50.00 110.13 FF cst by analogy to CT-CT-N*; Eq.val. from
QM geo.opt.

CQ-NA-C 70.0 118.60 by analogy to CA-NC-CQ, parm99.dat

H5-CQ-NA 50.0 115.45 by analogy to H5-CQ-NC, parm99.dat

NC-CQ-NA 70.0 129.10 by analogy to NC-CQ-NC, parm99.dat

CQ-NA-H 50.0 118.00 by analogy to CA-NA-H, parm99.dat

DIHEDRAL

X -CQ-NA-X 4 6.00 180.0 2.0 by analogy to X -CA-NA-X,
parm99.dat

NONB

8.3. Modified NAB program

8.3.1. fd_helix.nab file

```
// put_tmp is a helper function to simplify the code somewhat -  
// all it does is output the proper elements to a temporary  
// pdb file.  
  
int put_tmp( float x, float y, float z, int count, int rescount,  
string atomname, string resname, file outfile, int nres ) {  
  
// check if 5' end  
  
if( rescount == 1 || rescount == ( nres / 2 ) + 1 )  
    if( atomname =~ "P" )  
        return 1;  
  
if( atomname == "O1' " )  
    atomname = "O4'";  
  
// I have no idea why this is necessary.. would be just as easy  
// to change O1' to O4' in all data files, but they must have had  
// some reason for naming it O1' in the data.  
  
fprintf(outfile, "ATOM %5d %-4s %3s%6d    %8.3lf%8.3lf%8.3lf\n",  
        count, atomname, resname, rescount, x, y, z );  
  
return 1;  
  
};  
  
// fd_helix is a function designed to build helices of varying types
```

```

// from fiber diffraction data. The current available types are:

// arna          Right Handed A-RNA (Arnott)
// aprna         Right Handed A-PRIME RNA (Arnott)
// lbdna         Right Handed B-DNA (Langridge)
// abdna         Right Handed B-DNA (Arnott)
// sbdna         Left Handed B-DNA (Sasisekharan)
// adna          Right Handed A-DNA (Arnott)

// fd_helix will create a temporary file in your working directory
// called nab_tmp.pdb. This file contains the helix_before_the
// addition of hydrogens. This is ordinarily deleted, but could be
// examined if things go wrong.

molecule fd_helix( string helix_type, string seq, string acid_type )
{
// ----- //

// Data Segment

molecule m;

// These arrays are used to store the r, phi, zz values of the
// various atom types within each residue:

float ade_r[ hashed ];
float ade_phi[ hashed ];
float ade_zz[ hashed ];

float gua_r[ hashed ];
float gua_phi[ hashed ];
float gua_zz[ hashed ];

float thy_r[ hashed ];
float thy_phi[ hashed ];
float thy_zz[ hashed ];

float ura_r[ hashed ];
float ura_phi[ hashed ];
float ura_zz[ hashed ];

float cyt_r[ hashed ];
float cyt_phi[ hashed ];
float cyt_zz[ hashed ];

float apn_r[ hashed ];
float apn_phi[ hashed ];
float apn_zz[ hashed ];

float tpn_r[ hashed ];
float tpn_phi[ hashed ];
float tpn_zz[ hashed ];

float cpn_r[ hashed ];
float cpn_phi[ hashed ];
float cpn_zz[ hashed ];

float gpn_r[ hashed ];
float gpn_phi[ hashed ];
float gpn_zz[ hashed ];

```

```

float dim_r[ hashed ];
float dim_phi[ hashed ];
float dim_zz[ hashed ];

float nbc_r[ hashed ];
float nbc_phi[ hashed ];
float nbc_zz[ hashed ];

// These are simply temp variables used to aid in filling the arrays.

float temp_r, temp_phi, temp_zz;

// These are used to calculate and output coordinates to the temporary pdb file.

float x, y, z, yyr, xrad;

// height values are angstroms, rotation; values are degrees.

float current_height, current_rotation;
float height_increment, rotation_increment;

float hxht[ hashed ];
float hxrep[ hashed ];

string tempname, cseq, fullseq, buffer, restype;
string temp, amberhome;
string resout;

int nres, nresh, i, hxmul, count, chain, begin, end;

file infile, outfile;

// -----

// Code Segment

hxht[ "arna" ] = 3.30; hxht[ "aprna" ] = 3.00; hxht[ "lbdna" ] = 3.38;
hxht[ "abdna" ] = 3.38; hxht[ "sbdna" ] = -3.38; hxht[ "adna" ] = 2.56;
hxht[ "apna" ] = 3.30;

hxrep["arna"] = 23.2; hxrep["aprna"] = 30.0; hxrep["lbdna"] = 36.0;
hxrep["abdna"] = 36.0; hxrep["sbdna"] = 36.0; hxrep["adna"] = 32.7;
hxrep["apna"] = 23.2;

temp = wc_complement( seq, acid_type + "amber94.rlb", acid_type );
cseq = "";
for( i = length( temp ); i >= 1; i-- ) {
    cseq += substr( temp, i, 1 );
}
fullseq = seq + cseq;

nresh = length( seq );

nres = length( fullseq );

if( !( amberhome = getenv( "AMBERHOME" ) ) ){
    fprintf( stderr, "AMBERHOME not defined.\n" );
    exit( 1 );
}

```

```

temp = amberhome + "/dat/fd_data/" + helix_type + ".dat";
printf("Data file: %s\n", temp);

infile = fopen( temp, "r" );
if( infile == NULL ){
    fprintf( stderr, "Unable to open data file %s; exiting\n", temp );
    exit(1);
}

outfile = fopen( "nab_tmp.pdb", "w" );

// Read lines from data file and store r, phi, zz values as appropriate
// residue type.

while( buffer = getline( infile ) ) {
    sscanf( buffer, "%s %lf %lf %lf %s", tempname, temp_r, temp_phi,
            temp_zz, restype );

    if( restype == "A" || restype == "a" ) {
        ade_r[ tempname ] = temp_r;
        ade_phi[ tempname ] = temp_phi;
        ade_zz[ tempname ] = temp_zz;
    }
    else if( restype == "G" || restype == "g" ) {
        gua_r[ tempname ] = temp_r;
        gua_phi[ tempname ] = temp_phi;
        gua_zz[ tempname ] = temp_zz;
    }
    else if( restype == "T" || restype == "t" ) {
        thy_r[ tempname ] = temp_r;
        thy_phi[ tempname ] = temp_phi;
        thy_zz[ tempname ] = temp_zz;
    }
    else if( restype == "U" || restype == "u" ) {
        ura_r[ tempname ] = temp_r;
        ura_phi[ tempname ] = temp_phi;
        ura_zz[ tempname ] = temp_zz;
    }
    else if( restype == "C" || restype == "c" ) {
        cyt_r[ tempname ] = temp_r;
        cyt_phi[ tempname ] = temp_phi;
        cyt_zz[ tempname ] = temp_zz;
    }
    else if( restype == "X" || restype == "x" ) {
        apn_r[ tempname ] = temp_r;
        apn_phi[ tempname ] = temp_phi;
        apn_zz[ tempname ] = temp_zz;
    }
    else if( restype == "Y" || restype == "y" ) {
        tpn_r[ tempname ] = temp_r;
        tpn_phi[ tempname ] = temp_phi;
        tpn_zz[ tempname ] = temp_zz;
    }
    else if( restype == "J" || restype == "j" ) {
        cpn_r[ tempname ] = temp_r;
        cpn_phi[ tempname ] = temp_phi;
        cpn_zz[ tempname ] = temp_zz;
    }
    else if( restype == "K" || restype == "k" ) {
        gpn_r[ tempname ] = temp_r;
        gpn_phi[ tempname ] = temp_phi;
        gpn_zz[ tempname ] = temp_zz;
    }
}

```

```

    }
    else if( restype == "W" || restype == "w" ) {
        dim_r[ tempname ] = temp_r;
        dim_phi[ tempname ] = temp_phi;
        dim_zz[ tempname ] = temp_zz;
    }
    else if( restype == "Q" || restype == "q" ) {
        nbc_r[ tempname ] = temp_r;
        nbc_phi[ tempname ] = temp_phi;
        nbc_zz[ tempname ] = temp_zz;
    }
}

height_increment = hxht[ helix_type ];
rotation_increment = hxrep[ helix_type ];
current_height = 0;
current_rotation = 0;
count = 0;

// Here we build the actual molecule - it is output to a temporary
// pdb file in order to allow the addition of hydrogens as a final
// stage.

for( chain = 1; chain <= 2; chain++ ) {

    if(chain == 1) {
        begin = 1;
        end = nresh;
        hxmul = -1;
    }

    else if( chain == 2) {
        begin = nresh + 1;
        end = nres;
        hxmul = 1;
    }

    for( i = begin; i <= end; i++ ) {
        restype = substr( fullseq, i, 1 );
        if( restype == "A" || restype == "a" ) {
            resout = "DA"; if( acid_type == "rna" ) resout = "A";
            for( tempname in ade_r ) {
                count++;
                yyr = (hxmul * ade_phi[ tempname ] + current_rotation);
                xrad = ade_r[ tempname ];
                x = xrad * cos(yyr);
                y = xrad * sin(yyr);
                z = hxmul * ade_zz[ tempname ] + current_height;
                put_tmp( x, y, z, count, i, tempname, resout, outfile,
nres);
            }

        }

        else if( restype == "G" || restype == "g" ) {
            resout = "DG"; if( acid_type == "rna" ) resout = "G";
            for( tempname in gua_r ) {
                count++;
                yyr = (hxmul * gua_phi[ tempname ] + current_rotation);
                xrad = gua_r[ tempname ];
                x = xrad * cos(yyr);
                y = xrad * sin(yyr);

```

```

        z = hxmul * gua_zz[ tempname ] + current_height;
        put_tmp( x, y, z, count, i, tempname, resout, outfile,
nres);
    }
}

else if( restype == "T" || restype == "t" ) {
    resout = "DT";
    for( tempname in thy_r ) {
        count++;
        yyr = (hxmul * thy_phi[ tempname ] + current_rotation);
        xrad = thy_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * thy_zz[ tempname ] + current_height;
        put_tmp( x, y, z, count, i, tempname, resout, outfile,
nres);
    }
}

else if( restype == "U" || restype == "u" ) {
    resout = "U";
    for( tempname in ura_r ) {
        count++;
        yyr = (hxmul * ura_phi[ tempname ] + current_rotation);
        xrad = ura_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * ura_zz[ tempname ] + current_height;
        put_tmp( x, y, z, count, i, tempname, resout, outfile,
nres);
    }
}

else if( restype == "C" || restype == "c" ) {
    resout = "DC"; if( acid_type == "rna" ) resout = "C";
    for( tempname in cyt_r ) {
        count++;
        yyr = (hxmul * cyt_phi[ tempname ] + current_rotation);
        xrad = cyt_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * cyt_zz[ tempname ] + current_height;
        put_tmp( x, y, z, count, i, tempname, resout, outfile,
nres);
    }
}

else if( restype == "X" || restype == "x" ) {
    resout = "APN"; if( acid_type == "rna" ) resout = "C";
    for( tempname in apn_r ) {
        count++;
        yyr = (hxmul * apn_phi[ tempname ] +
current_rotation);
        xrad = apn_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * apn_zz[ tempname ] + current_height;

```

```

                                put_tmp( x, y, z, count, i, tempname, resout,
outfile, nres);
                                }
                                }
else if( restype == "Y" || restype == "y" ) {
    resout = "TPN"; if( acid_type == "rna" ) resout = "C";
    for( tempname in tpn_r ) {
        count++;
        yyr = (hxmul * tpn_phi[ tempname ] +
current_rotation);
        xrad = tpn_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * tpn_zz[ tempname ] + current_height;
        put_tmp( x, y, z, count, i, tempname, resout,
outfile, nres);
    }
}
else if( restype == "J" || restype == "j" ) {
    resout = "CPN"; if( acid_type == "rna" ) resout = "C";
    for( tempname in cpn_r ) {
        count++;
        yyr = (hxmul * cpn_phi[ tempname ] +
current_rotation);
        xrad = cpn_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * cpn_zz[ tempname ] + current_height;
        put_tmp( x, y, z, count, i, tempname, resout,
outfile, nres);
    }
}
else if( restype == "K" || restype == "k" ) {
    resout = "GPN"; if( acid_type == "rna" ) resout = "C";
    for( tempname in gpn_r ) {
        count++;
        yyr = (hxmul * gpn_phi[ tempname ] +
current_rotation);
        xrad = gpn_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * gpn_zz[ tempname ] + current_height;
        put_tmp( x, y, z, count, i, tempname, resout,
outfile, nres);
    }
}
}
else if( restype == "W" || restype == "w" ) {
    resout = "DIM"; if( acid_type == "rna" ) resout = "C";
    for( tempname in dim_r ) {
        count++;
        yyr = (hxmul * dim_phi[ tempname ] +
current_rotation);
        xrad = dim_r[ tempname ];
        x = xrad * cos(yyr);
        y = xrad * sin(yyr);
        z = hxmul * dim_zz[ tempname ] + current_height;

```

```

                                put_tmp( x, y, z, count, i, tempname, resout,
outfile, nres);
                                }

                                }
else if( restype == "Q" || restype == "q" ) {
    resout = "NBC"; if( acid_type == "rna" ) resout = "C";
    for( tempname in nbc_r ) {
        count++;
        yyr = (hxmul * nbc_phi[ tempname ] +
current_rotation);
                                xrad = nbc_r[ tempname ];
                                x = xrad * cos(yyr);
                                y = xrad * sin(yyr);
                                z = hxmul * nbc_zz[ tempname ] + current_height;
                                put_tmp( x, y, z, count, i, tempname, resout,
outfile, nres);
                                }

                                }

                                // Increase unit twist and height

                                current_height += height_increment;
                                current_rotation += rotation_increment;
                                }

height_increment = -height_increment;
rotation_increment = -rotation_increment;

current_rotation += rotation_increment;
current_height += height_increment;

if(chain == 1)
    fprintf( outfile, "TER\n" ); // Need TER card for getpdb_prm to
                                // function properly
}

fclose( infile );

// Close outfile in order to flush output stream.
// Otherwise nab_tmp.pdb would be incomplete when
// read by getpdb_prm

fclose( outfile );

m = getpdb_prm( "nab_tmp.pdb", "leaprc.pna", "", 0 );
unlink( "nab_tmp.pdb" );

return m;

};

```

8.3.2. wc_complement.nab

```
// wc_complement() - create W/C string to seq.
string      wc_complement( string seq, string rlib, string rlt )
//      (note the rlib is unused: just there for backwards compatibility)
{
    string      acbase, base, wcbase, wcseq;
    int      i, len;

    if( rlt == "dna" )
        acbase = "t";
    else if( rlt == "rna" )
        acbase = "u";
    else if( rlt == "pna" )
        acbase = "t";
    else{
        fprintf( stderr,
            "wc_complement: rlt (%s) is not dna/rna, has no W/C complement\n",
                rlt );
        exit( 1 );
    }

    len = length( seq );
    wcseq = NULL;
    for( i = 1; i <= len; i = i + 1 ){
        base = substr( seq, i, 1 );
        if( base == "a" || base == "A" )
            wcbase = acbase;
        else if( base == "c" || base == "C" )
            wcbase = "g";
        else if( base == "g" || base == "G" )
            wcbase = "c";
        else if( base == "t" || base == "T" )
            wcbase = "a";
        else if( base == "u" || base == "U" )
            wcbase = "a";
        else if( base == "x" || base == "APN" )
            wcbase = "acbase";
        else if( base == "y" || base == "TPN" )
            wcbase = "a";
        else if( base == "j" || base == "CPN" )
            wcbase = "g";
        else if( base == "k" || base == "GPN" )
            wcbase = "c";
        else if( base == "w" || base == "DIM" )
            wcbase = "a";
        else if( base == "q" || base == "NBC" )
            wcbase = "a";

        else{
            fprintf( stderr, "wc_complement: unknown base %s\n",
                base );
            exit( 1 );
        }
        wcseq = wcseq + wcbase;
    }
    return( wcseq );
};
```

8.3.3. pna.amber94.rlb file

```
#
#      bpna single bases
#
pdb      pna.amber94/pna.amber94.pdb
bnd      pna.amber94/pna.amber94.bnd
qr       pna.amber94/pna.amber94.qr
chi      pna.amber94/pna.amber94.chi
type     pna
atomtype      all
```

8.3.4. pna.amber94.pdb file

```
ATOM      1  P      ADE      1      -2.946  -5.462  -1.138
ATOM      2  O1P    ADE      1      -2.555  -6.846  -0.805
ATOM      3  O2P    ADE      1      -4.059  -4.892  -0.340
ATOM      4  O5'    ADE      1      -1.679  -4.498  -1.044
ATOM      5  C5'    ADE      1      -0.625  -4.632  -2.031
ATOM      6  H5'1    ADE      1      -0.795  -3.931  -2.848
ATOM      7  H5'2    ADE      1      -0.617  -5.647  -2.425
ATOM      8  C4'    ADE      1       0.712  -4.336  -1.394
ATOM      9  H4'    ADE      1       1.480  -4.833  -1.986
ATOM     10  O4'    ADE      1       0.889  -2.884  -1.330
ATOM     11  C1'    ADE      1       0.731  -2.441   0.004
ATOM     12  H1'    ADE      1       1.684  -2.029   0.336
ATOM     13  N9     ADE      1      -0.275  -1.345   0.002
ATOM     14  C8     ADE      1      -1.640  -1.447   0.001
ATOM     15  H8     ADE      1      -2.167  -2.401   0.007
ATOM     16  N7     ADE      1      -2.248  -0.298  -0.007
ATOM     17  C5     ADE      1      -1.215   0.623   0.003
ATOM     18  C6     ADE      1      -1.198   2.033  -0.001
ATOM     19  N6     ADE      1      -2.314   2.780  -0.005
ATOM     20  H61    ADE      1      -2.246   3.807  -0.008
ATOM     21  H62    ADE      1      -3.238   2.324  -0.005
ATOM     22  N1     ADE      1       0.000   2.637   0.000
ATOM     23  C2     ADE      1       1.098   1.887   0.005
ATOM     24  H2     ADE      1       2.038   2.438   0.011
ATOM     25  N3     ADE      1       1.216   0.576   0.003
ATOM     26  C4     ADE      1      -0.000   0.000  -0.000
ATOM     27  C3'    ADE      1       0.894  -4.801   0.050
ATOM     28  H3'    ADE      1       0.352  -5.728   0.235
ATOM     29  C2'    ADE      1       0.330  -3.640   0.853
ATOM     30  H2'1    ADE      1      -0.750  -3.724   0.966
ATOM     31  H2'2    ADE      1       0.763  -3.602   1.853
ATOM     32  O3'    ADE      1       2.267  -4.981   0.388
ATOM      1  P      GUA      1      -2.846  -5.524  -1.132
ATOM      2  O1P    GUA      1      -2.426  -6.899  -0.800
ATOM      3  O2P    GUA      1      -3.969  -4.977  -0.332
ATOM      4  O5'    GUA      1      -1.599  -4.534  -1.040
ATOM      5  C5'    GUA      1      -0.545  -4.646  -2.029
ATOM      6  H5'1    GUA      1      -0.731  -3.949  -2.846
ATOM      7  H5'2    GUA      1      -0.516  -5.661  -2.424
ATOM      8  C4'    GUA      1       0.787  -4.322  -1.395
ATOM      9  H4'    GUA      1       1.564  -4.803  -1.989
ATOM     10  O4'    GUA      1       0.935  -2.868  -1.331
ATOM     11  C1'    GUA      1       0.770  -2.428   0.003
ATOM     12  H1'    GUA      1       1.715  -1.996   0.333
ATOM     13  N9     GUA      1      -0.259  -1.353   0.003
```

ATOM	14	C8	GUA	1	-1.634	-1.459	0.003
ATOM	15	H8	GUA	1	-2.154	-2.417	0.003
ATOM	16	N7	GUA	1	-2.259	-0.310	0.005
ATOM	17	C5	GUA	1	-1.230	0.634	0.002
ATOM	18	C6	GUA	1	-1.287	2.046	0.000
ATOM	19	O6	GUA	1	-2.273	2.783	0.000
ATOM	20	N1	GUA	1	-0.000	2.612	-0.000
ATOM	21	H1	GUA	1	0.067	3.641	-0.007
ATOM	22	C2	GUA	1	1.180	1.898	0.008
ATOM	23	N2	GUA	1	2.306	2.619	0.011
ATOM	24	H21	GUA	1	2.258	3.648	0.008
ATOM	25	H22	GUA	1	3.220	2.145	0.017
ATOM	26	N3	GUA	1	1.235	0.568	0.002
ATOM	27	C4	GUA	1	0.000	-0.000	0.000
ATOM	28	C3'	GUA	1	0.982	-4.784	0.048
ATOM	29	H3'	GUA	1	0.459	-5.722	0.235
ATOM	30	C2'	GUA	1	0.396	-3.635	0.853
ATOM	31	H2'1	GUA	1	-0.683	-3.741	0.968
ATOM	32	H2'2	GUA	1	0.829	-3.588	1.852
ATOM	33	O3'	GUA	1	2.359	-4.936	0.384
ATOM	1	P	THY	1	-0.049	-4.059	-1.130
ATOM	2	O1P	THY	1	0.760	-5.248	-0.796
ATOM	3	O2P	THY	1	-1.284	-3.869	-0.332
ATOM	4	O5'	THY	1	0.849	-2.745	-1.038
ATOM	5	C5'	THY	1	1.890	-2.540	-2.026
ATOM	6	H5'1	THY	1	1.507	-1.930	-2.844
ATOM	7	H5'2	THY	1	2.219	-3.501	-2.419
ATOM	8	C4'	THY	1	3.066	-1.836	-1.390
ATOM	9	H4'	THY	1	3.952	-2.066	-1.983
ATOM	10	O4'	THY	1	2.776	-0.403	-1.328
ATOM	11	C1'	THY	1	2.486	-0.030	0.005
ATOM	12	H1'	THY	1	3.261	0.662	0.336
ATOM	13	N1	THY	1	1.186	0.692	0.003
ATOM	14	C6	THY	1	-0.000	-0.000	-0.000
ATOM	15	H6	THY	1	0.015	-1.090	-0.005
ATOM	16	C5	THY	1	-1.181	0.648	0.003
ATOM	17	C7	THY	1	-2.504	-0.058	-0.006
ATOM	18	H71	THY	1	-2.630	-0.606	0.928
ATOM	19	H72	THY	1	-3.306	0.673	-0.109
ATOM	20	H73	THY	1	-2.539	-0.756	-0.843
ATOM	21	C4	THY	1	-1.234	2.090	-0.001
ATOM	22	O4	THY	1	-2.272	2.762	-0.003
ATOM	23	N3	THY	1	-0.000	2.699	-0.000
ATOM	24	H3	THY	1	0.007	3.729	-0.003
ATOM	25	C2	THY	1	1.230	2.071	0.004
ATOM	26	O2	THY	1	2.275	2.697	0.007
ATOM	27	C3'	THY	1	3.387	-2.218	0.053
ATOM	28	H3'	THY	1	3.165	-3.268	0.241
ATOM	29	C2'	THY	1	2.485	-1.294	0.856
ATOM	30	H2'1	THY	1	1.487	-1.714	0.970
ATOM	31	H2'2	THY	1	2.884	-1.120	1.856
ATOM	32	O3'	THY	1	4.747	-1.955	0.391
ATOM	1	P	CYT	1	-0.006	-4.082	-1.120
ATOM	2	O1P	CYT	1	0.817	-5.261	-0.785
ATOM	3	O2P	CYT	1	-1.243	-3.906	-0.321
ATOM	4	O5'	CYT	1	0.876	-2.757	-1.031
ATOM	5	C5'	CYT	1	1.913	-2.541	-2.021
ATOM	6	H5'1	CYT	1	1.521	-1.937	-2.840
ATOM	7	H5'2	CYT	1	2.253	-3.499	-2.413
ATOM	8	C4'	CYT	1	3.081	-1.821	-1.388
ATOM	9	H4'	CYT	1	3.968	-2.042	-1.982
ATOM	10	O4'	CYT	1	2.773	-0.392	-1.329

ATOM	11	C1'	CYT	1	2.481	-0.021	0.005
ATOM	12	H1'	CYT	1	3.247	0.682	0.333
ATOM	13	N1	CYT	1	1.171	0.686	0.002
ATOM	14	C6	CYT	1	-0.000	0.000	-0.000
ATOM	15	H6	CYT	1	0.020	-1.090	-0.004
ATOM	16	C5	CYT	1	-1.199	0.644	0.002
ATOM	17	H5	CYT	1	-2.142	0.097	0.009
ATOM	18	C4	CYT	1	-1.161	2.073	-0.005
ATOM	19	N4	CYT	1	-2.286	2.770	-0.010
ATOM	20	H41	CYT	1	-3.194	2.283	-0.013
ATOM	21	H42	CYT	1	-2.255	3.800	-0.010
ATOM	22	N3	CYT	1	-0.000	2.741	-0.000
ATOM	23	C2	CYT	1	1.183	2.079	-0.000
ATOM	24	O2	CYT	1	2.271	2.668	0.004
ATOM	25	C3'	CYT	1	3.408	-2.197	0.055
ATOM	26	H3'	CYT	1	3.199	-3.250	0.245
ATOM	27	C2'	CYT	1	2.496	-1.282	0.858
ATOM	28	H2'1	CYT	1	1.503	-1.715	0.974
ATOM	29	H2'2	CYT	1	2.894	-1.101	1.857
ATOM	30	O3'	CYT	1	4.765	-1.917	0.391
ATOM	1	P	APU	1	-0.049	-4.059	-1.130
ATOM	2	O1P	APU	1	0.760	-5.248	-0.796
ATOM	3	O2P	APU	1	-1.284	-3.869	-0.332
ATOM	4	O5'	APU	1	0.849	-2.745	-1.038
ATOM	5	C5'	APU	1	1.890	-2.540	-2.026
ATOM	6	H5'1	APU	1	1.507	-1.930	-2.844
ATOM	7	H5'2	APU	1	2.219	-3.501	-2.419
ATOM	8	C4'	APU	1	3.066	-1.836	-1.390
ATOM	9	H4'	APU	1	3.952	-2.066	-1.983
ATOM	10	O4'	APU	1	2.776	-0.403	-1.328
ATOM	11	C1'	APU	1	2.486	-0.030	0.005
ATOM	12	H1'	APU	1	3.261	0.662	0.336
ATOM	13	O1'	APU	1	1.186	0.692	0.003
ATOM	14	HO1'	APU	1	-0.000	-0.000	-0.000
ATOM	15	C3'	APU	1	3.387	-2.218	0.053
ATOM	16	H3'	APU	1	3.165	-3.268	0.241
ATOM	17	C2'	APU	1	2.485	-1.294	0.856
ATOM	18	H2'1	APU	1	1.487	-1.714	0.970
ATOM	19	H2'2	APU	1	2.884	-1.120	1.856
ATOM	20	O3'	APU	1	4.747	-1.955	0.391
ATOM	1	N1'	APN	1	3.621	-1.450	-0.069
ATOM	2	H1'1	APN	1	4.246	-1.272	0.683
ATOM	3	C2'	APN	1	3.458	-0.420	-1.077
ATOM	4	H2'1	APN	1	3.628	-0.848	-2.057
ATOM	5	H2'2	APN	1	4.227	0.322	-0.906
ATOM	6	C3'	APN	1	2.080	0.241	-1.080
ATOM	7	H3'1	APN	1	1.324	-0.471	-1.363
ATOM	8	H3'2	APN	1	2.071	1.016	-1.836
ATOM	9	N4'	APN	1	1.741	0.868	0.196
ATOM	10	C5'	APN	1	2.299	2.197	0.430
ATOM	11	H5'1	APN	1	2.392	2.357	1.493
ATOM	12	H5'2	APN	1	3.276	2.251	-0.025
ATOM	13	C'	APN	1	1.450	3.289	-0.217
ATOM	14	O1'	APN	1	1.768	3.771	-1.273
ATOM	15	C7'	APN	1	0.858	0.403	1.100
ATOM	16	O7'	APN	1	0.506	1.058	2.054
ATOM	17	C8'	APN	1	0.288	-1.011	0.978
ATOM	18	H8'1	APN	1	0.638	-1.548	1.851
ATOM	19	H8'2	APN	1	0.593	-1.558	0.104
ATOM	20	N9	APN	1	-1.153	-0.974	1.005
ATOM	21	C8	APN	1	-1.982	-0.899	2.098
ATOM	22	H8	APN	1	-1.577	-0.897	3.089

ATOM	23	N7	APN	1	-3.226	-0.831	1.805
ATOM	24	C6	APN	1	-4.279	-0.782	-0.521
ATOM	25	N6	APN	1	-5.576	-0.734	-0.172
ATOM	26	H61	APN	1	-5.821	-0.568	0.777
ATOM	27	H62	APN	1	-6.246	-0.541	-0.880
ATOM	28	C5	APN	1	-3.248	-0.843	0.423
ATOM	29	C4	APN	1	-1.972	-0.921	-0.084
ATOM	30	N3	APN	1	-1.628	-0.930	-1.367
ATOM	31	C2	APN	1	-2.681	-0.862	-2.150
ATOM	32	H2	APN	1	-2.486	-0.862	-3.207
ATOM	33	N1	APN	1	-3.961	-0.790	-1.811
ATOM	1	C'	CAP	1	0.000	0.000	0.000
ATOM	2	O1'	CAP	1	-0.512	-1.124	0.000
ATOM	3	OXT	CAP	1	-0.541	1.108	0.000
ATOM	4	N1'	CAP	1	4.659	2.044	1.148
ATOM	5	H1'1	CAP	1	5.197	1.232	0.950
ATOM	6	C2'	CAP	1	3.404	1.871	1.856
ATOM	7	H2'1	CAP	1	3.389	2.519	2.723
ATOM	8	H2'2	CAP	1	3.374	0.849	2.212
ATOM	9	C3'	CAP	1	2.164	2.173	1.015
ATOM	10	H3'1	CAP	1	2.133	3.217	0.756
ATOM	11	H3'2	CAP	1	1.284	1.978	1.616
ATOM	12	N4'	CAP	1	2.069	1.344	-0.185
ATOM	13	C5'	CAP	1	1.527	0.000	0.000
ATOM	14	H5'1	CAP	1	1.916	-0.644	-0.773
ATOM	15	H5'2	CAP	1	1.839	-0.372	0.964
ATOM	16	C7'	CAP	1	2.293	1.748	-1.450
ATOM	17	O7'	CAP	1	2.033	1.053	-2.405
ATOM	18	C8'	CAP	1	2.936	3.107	-1.732
ATOM	19	H8'1	CAP	1	3.884	2.893	-2.208
ATOM	20	H8'2	CAP	1	3.126	3.716	-0.867
ATOM	21	N9	CAP	1	2.119	3.866	-2.647
ATOM	22	C8	CAP	1	2.068	3.777	-4.016
ATOM	23	H8	CAP	1	2.712	3.103	-4.543
ATOM	24	N7	CAP	1	1.204	4.551	-4.557
ATOM	25	C6	CAP	1	-0.402	6.153	-3.385
ATOM	26	N6	CAP	1	-1.010	6.679	-4.463
ATOM	27	H61	CAP	1	-0.863	6.269	-5.356
ATOM	28	H62	CAP	1	-1.821	7.236	-4.319
ATOM	29	C5	CAP	1	0.614	5.198	-3.487
ATOM	30	C4	CAP	1	1.164	4.775	-2.299
ATOM	31	N3	CAP	1	0.818	5.173	-1.081
ATOM	32	C2	CAP	1	-0.148	6.063	-1.118
ATOM	33	H2	CAP	1	-0.487	6.434	-0.168
ATOM	34	N1	CAP	1	-0.767	6.567	-2.177
ATOM	1	C'	CCP	1	0.000	0.000	0.000
ATOM	2	O1'	CCP	1	-0.512	-1.124	0.000
ATOM	3	OXT	CCP	1	-0.541	1.108	0.000
ATOM	4	N1'	CCP	1	4.548	2.222	1.204
ATOM	5	H1'1	CCP	1	5.114	1.414	1.093
ATOM	6	C2'	CCP	1	3.247	2.062	1.829
ATOM	7	H2'1	CCP	1	3.151	2.775	2.637
ATOM	8	H2'2	CCP	1	3.221	1.070	2.263
ATOM	9	C3'	CCP	1	2.066	2.253	0.881
ATOM	10	H3'1	CCP	1	2.020	3.268	0.530
ATOM	11	H3'2	CCP	1	1.151	2.079	1.435
ATOM	12	N4'	CCP	1	2.080	1.327	-0.251
ATOM	13	C5'	CCP	1	1.528	0.000	0.000
ATOM	14	H5'1	CCP	1	1.912	-0.684	-0.741
ATOM	15	H5'2	CCP	1	1.839	-0.327	0.981
ATOM	16	C7'	CCP	1	2.319	1.666	-1.533
ATOM	17	O7'	CCP	1	2.067	0.925	-2.456

ATOM	18	C8'	CCP	1	3.021	2.983	-1.865
ATOM	19	H8'1	CCP	1	4.035	2.720	-2.140
ATOM	20	H8'2	CCP	1	3.064	3.687	-1.056
ATOM	21	N1	CCP	1	2.387	3.609	-3.006
ATOM	22	C2	CCP	1	1.123	4.184	-2.781
ATOM	23	O2	CCP	1	0.673	4.116	-1.668
ATOM	24	N3	CCP	1	0.498	4.768	-3.831
ATOM	25	C4	CCP	1	1.022	4.716	-5.017
ATOM	26	N4	CCP	1	0.366	5.334	-6.017
ATOM	27	H41	CCP	1	0.615	5.181	-6.965
ATOM	28	H42	CCP	1	-0.543	5.689	-5.820
ATOM	29	C5	CCP	1	2.263	4.051	-5.307
ATOM	30	H5	CCP	1	2.665	3.994	-6.298
ATOM	31	C6	CCP	1	2.891	3.508	-4.254
ATOM	32	H6	CCP	1	3.822	2.984	-4.349
ATOM	1	C'	CGP	1	0.000	0.000	0.000
ATOM	2	O1'	CGP	1	-0.512	-1.124	0.000
ATOM	3	OXT	CGP	1	-0.541	1.108	0.000
ATOM	4	N1'	CGP	1	4.733	2.038	1.113
ATOM	5	H1'1	CGP	1	5.293	1.308	0.738
ATOM	6	C2'	CGP	1	3.569	1.675	1.899
ATOM	7	H2'1	CGP	1	3.614	2.178	2.857
ATOM	8	H2'2	CGP	1	3.630	0.610	2.086
ATOM	9	C3'	CGP	1	2.237	2.023	1.239
ATOM	10	H3'1	CGP	1	2.152	3.088	1.129
ATOM	11	H3'2	CGP	1	1.428	1.718	1.889
ATOM	12	N4'	CGP	1	2.057	1.357	-0.047
ATOM	13	C5'	CGP	1	1.529	0.000	0.000
ATOM	14	H5'1	CGP	1	1.923	-0.570	-0.827
ATOM	15	H5'2	CGP	1	1.845	-0.462	0.924
ATOM	16	C7'	CGP	1	2.035	1.955	-1.260
ATOM	17	O7'	CGP	1	1.713	1.358	-2.258
ATOM	18	C8'	CGP	1	2.465	3.417	-1.408
ATOM	19	H8'1	CGP	1	3.222	3.422	-2.180
ATOM	20	H8'2	CGP	1	2.885	3.852	-0.518
ATOM	21	N9	CGP	1	1.356	4.220	-1.882
ATOM	22	C8	CGP	1	1.365	5.151	-2.897
ATOM	23	H8	CGP	1	2.241	5.306	-3.493
ATOM	24	N7	CGP	1	0.255	5.768	-3.030
ATOM	25	C6	CGP	1	-1.923	5.510	-1.702
ATOM	26	O6	CGP	1	-2.709	6.291	-2.147
ATOM	27	C5	CGP	1	-0.560	5.233	-2.056
ATOM	28	C4	CGP	1	0.114	4.280	-1.346
ATOM	29	N3	CGP	1	-0.327	3.501	-0.332
ATOM	30	C2	CGP	1	-1.560	3.725	-0.016
ATOM	31	N2	CGP	1	-2.134	3.020	0.987
ATOM	32	H21	CGP	1	-1.655	2.176	1.234
ATOM	33	H22	CGP	1	-3.127	2.961	1.014
ATOM	34	N1	CGP	1	-2.322	4.667	-0.633
ATOM	35	H1	CGP	1	-3.253	4.834	-0.316
ATOM	1	N1'	CPN	1	3.313	-1.543	0.213
ATOM	2	H1'1	CPN	1	3.703	-1.508	1.125
ATOM	3	C2'	CPN	1	3.410	-0.355	-0.616
ATOM	4	H2'1	CPN	1	3.831	-0.625	-1.576
ATOM	5	H2'2	CPN	1	4.106	0.317	-0.131
ATOM	6	C3'	CPN	1	2.079	0.351	-0.862
ATOM	7	H3'1	CPN	1	1.419	-0.270	-1.440
ATOM	8	H3'2	CPN	1	2.266	1.236	-1.458
ATOM	9	N4'	CPN	1	1.418	0.779	0.370
ATOM	10	C5'	CPN	1	1.868	2.049	0.931
ATOM	11	H5'1	CPN	1	1.604	2.082	1.976
ATOM	12	H5'2	CPN	1	2.942	2.113	0.832

ATOM	13	C'	CPN	1	1.286	3.249	0.184
ATOM	14	O1'	CPN	1	1.962	3.877	-0.589
ATOM	15	C7'	CPN	1	0.290	0.243	0.878
ATOM	16	O7'	CPN	1	-0.353	0.791	1.744
ATOM	17	C8'	CPN	1	-0.169	-1.147	0.438
ATOM	18	H8'1	CPN	1	0.068	-1.813	1.258
ATOM	19	H8'2	CPN	1	0.309	-1.521	-0.447
ATOM	20	N1	CPN	1	-1.603	-1.156	0.242
ATOM	21	C2	CPN	1	-2.082	-0.490	-0.899
ATOM	22	O2	CPN	1	-1.272	0.028	-1.621
ATOM	23	N3	CPN	1	-3.422	-0.471	-1.105
ATOM	24	C4	CPN	1	-4.228	-0.981	-0.225
ATOM	25	N4	CPN	1	-5.545	-0.953	-0.495
ATOM	26	H41	CPN	1	-6.217	-1.160	0.206
ATOM	27	H42	CPN	1	-5.840	-0.435	-1.292
ATOM	28	C5	CPN	1	-3.781	-1.582	1.003
ATOM	29	H5	CPN	1	-4.464	-1.973	1.730
ATOM	30	C6	CPN	1	-2.454	-1.629	1.179
ATOM	31	H6	CPN	1	-2.006	-2.050	2.057
ATOM	1	C'	CTP	1	0.000	0.000	0.000
ATOM	2	O1'	CTP	1	-0.512	-1.124	0.000
ATOM	3	OXT	CTP	1	-0.541	1.108	0.000
ATOM	4	N1'	CTP	1	4.595	2.187	1.178
ATOM	5	H1'1	CTP	1	5.185	1.393	1.088
ATOM	6	C2'	CTP	1	3.310	2.006	1.830
ATOM	7	H2'1	CTP	1	3.231	2.691	2.665
ATOM	8	H2'2	CTP	1	3.294	1.000	2.230
ATOM	9	C3'	CTP	1	2.105	2.230	0.919
ATOM	10	H3'1	CTP	1	2.064	3.255	0.597
ATOM	11	H3'2	CTP	1	1.205	2.051	1.494
ATOM	12	N4'	CTP	1	2.076	1.333	-0.235
ATOM	13	C5'	CTP	1	1.528	0.000	0.000
ATOM	14	H5'1	CTP	1	1.915	-0.674	-0.748
ATOM	15	H5'2	CTP	1	1.839	-0.336	0.977
ATOM	16	C7'	CTP	1	2.306	1.688	-1.514
ATOM	17	O7'	CTP	1	2.032	0.967	-2.446
ATOM	18	C8'	CTP	1	3.017	3.004	-1.830
ATOM	19	H8'1	CTP	1	4.031	2.741	-2.101
ATOM	20	H8'2	CTP	1	3.063	3.701	-1.016
ATOM	21	N1	CTP	1	2.401	3.654	-2.970
ATOM	22	C2	CTP	1	1.246	4.351	-2.728
ATOM	23	O2	CTP	1	0.772	4.453	-1.631
ATOM	24	N3	CTP	1	0.681	4.920	-3.834
ATOM	25	H3	CTP	1	-0.150	5.450	-3.671
ATOM	26	C4	CTP	1	1.094	4.802	-5.153
ATOM	27	O4	CTP	1	0.491	5.348	-6.029
ATOM	28	C5	CTP	1	2.285	3.968	-5.341
ATOM	29	C6	CTP	1	2.849	3.439	-4.258
ATOM	30	H6	CTP	1	3.711	2.806	-4.326
ATOM	31	C7	CTP	1	2.782	3.753	-6.743
ATOM	32	H71	CTP	1	2.018	3.288	-7.356
ATOM	33	H72	CTP	1	3.037	4.698	-7.210
ATOM	34	H73	CTP	1	3.660	3.117	-6.748
ATOM	1	N1'	GPN	1	4.122	-0.011	-0.354
ATOM	2	H1'1	GPN	1	4.686	0.388	0.360
ATOM	3	C2'	GPN	1	3.375	0.883	-1.218
ATOM	4	H2'1	GPN	1	3.603	0.654	-2.252
ATOM	5	H2'2	GPN	1	3.728	1.888	-1.020
ATOM	6	C3'	GPN	1	1.861	0.807	-1.034
ATOM	7	H3'1	GPN	1	1.509	-0.169	-1.309
ATOM	8	H3'2	GPN	1	1.386	1.504	-1.712
ATOM	9	N4'	GPN	1	1.442	1.143	0.323

ATOM	10	C5'	GPN	1	1.275	2.561	0.612
ATOM	11	H5'1	GPN	1	1.508	2.749	1.648
ATOM	12	H5'2	GPN	1	1.958	3.123	-0.007
ATOM	13	C'	GPN	1	-0.137	3.034	0.262
ATOM	14	O1'	GPN	1	-0.437	3.292	-0.881
ATOM	15	C7'	GPN	1	0.909	0.291	1.228
ATOM	16	O7'	GPN	1	0.435	0.681	2.266
ATOM	17	C8'	GPN	1	0.931	-1.219	0.974
ATOM	18	H8'1	GPN	1	1.388	-1.656	1.851
ATOM	19	H8'2	GPN	1	1.499	-1.519	0.110
ATOM	20	N9	GPN	1	-0.420	-1.732	0.886
ATOM	21	C8	GPN	1	-0.938	-2.848	1.506
ATOM	22	H8	GPN	1	-0.351	-3.414	2.200
ATOM	23	N7	GPN	1	-2.143	-3.104	1.168
ATOM	24	C6	GPN	1	-3.681	-1.852	-0.458
ATOM	25	O6	GPN	1	-4.721	-2.438	-0.491
ATOM	26	C5	GPN	1	-2.471	-2.112	0.269
ATOM	27	C4	GPN	1	-1.413	-1.265	0.092
ATOM	28	N3	GPN	1	-1.328	-0.155	-0.676
ATOM	29	C2	GPN	1	-2.418	0.102	-1.321
ATOM	30	N2	GPN	1	-2.477	1.183	-2.135
ATOM	31	H21	GPN	1	-1.767	1.871	-1.970
ATOM	32	H22	GPN	1	-3.371	1.552	-2.367
ATOM	33	N1	GPN	1	-3.526	-0.682	-1.245
ATOM	34	H1	GPN	1	-4.324	-0.466	-1.804
ATOM	1	N1'	NAP	1	0.000	0.000	0.000
ATOM	2	H1'1	NAP	1	-0.372	-0.941	0.000
ATOM	3	H1'2	NAP	1	-0.372	0.470	0.815
ATOM	4	H1'3	NAP	1	-0.372	0.470	-0.815
ATOM	5	C2'	NAP	1	1.451	0.000	0.000
ATOM	6	H2'1	NAP	1	1.809	0.729	-0.716
ATOM	7	H2'2	NAP	1	1.772	-0.977	-0.339
ATOM	8	C3'	NAP	1	2.077	0.319	1.357
ATOM	9	H3'1	NAP	1	1.854	1.331	1.644
ATOM	10	H3'2	NAP	1	3.155	0.256	1.263
ATOM	11	N4'	NAP	1	1.673	-0.613	2.409
ATOM	12	C5'	NAP	1	2.392	-1.884	2.456
ATOM	13	H5'1	NAP	1	1.756	-2.630	2.906
ATOM	14	H5'2	NAP	1	2.636	-2.184	1.449
ATOM	15	C'	NAP	1	3.713	-1.759	3.213
ATOM	16	O1'	NAP	1	4.753	-1.642	2.619
ATOM	17	C7'	NAP	1	0.814	-0.353	3.413
ATOM	18	O7'	NAP	1	0.656	-1.115	4.340
ATOM	19	C8'	NAP	1	-0.041	0.915	3.400
ATOM	20	H8'1	NAP	1	-1.068	0.578	3.334
ATOM	21	H8'2	NAP	1	0.144	1.587	2.583
ATOM	22	N9	NAP	1	0.129	1.645	4.632
ATOM	23	C8	NAP	1	-0.484	1.427	5.841
ATOM	24	H8	NAP	1	-1.217	0.655	5.949
ATOM	25	N7	NAP	1	-0.093	2.213	6.773
ATOM	26	C6	NAP	1	1.678	4.047	6.623
ATOM	27	N6	NAP	1	1.615	4.514	7.882
ATOM	28	H61	NAP	1	1.102	4.007	8.566
ATOM	29	H62	NAP	1	2.320	5.146	8.184
ATOM	30	C5	NAP	1	0.862	3.006	6.165
ATOM	31	C4	NAP	1	1.016	2.659	4.844
ATOM	32	N3	NAP	1	1.863	3.200	3.975
ATOM	33	C2	NAP	1	2.574	4.158	4.526
ATOM	34	H2	NAP	1	3.286	4.647	3.886
ATOM	35	N1	NAP	1	2.533	4.605	5.774
ATOM	1	N1'	NCP	1	0.000	0.000	0.000
ATOM	2	H1'1	NCP	1	-0.372	-0.941	0.000

ATOM	3	H1'2	NCP	1	-0.372	0.470	0.815
ATOM	4	H1'3	NCP	1	-0.372	0.470	-0.815
ATOM	5	C2'	NCP	1	1.452	0.000	0.000
ATOM	6	H2'1	NCP	1	1.807	0.707	-0.739
ATOM	7	H2'2	NCP	1	1.772	-0.987	-0.311
ATOM	8	C3'	NCP	1	2.081	0.359	1.344
ATOM	9	H3'1	NCP	1	1.859	1.376	1.609
ATOM	10	H3'2	NCP	1	3.158	0.297	1.245
ATOM	11	N4'	NCP	1	1.683	-0.543	2.425
ATOM	12	C5'	NCP	1	2.433	-1.791	2.526
ATOM	13	H5'1	NCP	1	1.846	-2.507	3.080
ATOM	14	H5'2	NCP	1	2.613	-2.171	1.532
ATOM	15	C'	NCP	1	3.802	-1.592	3.175
ATOM	16	O1'	NCP	1	4.802	-1.581	2.506
ATOM	17	C7'	NCP	1	0.880	-0.217	3.457
ATOM	18	O7'	NCP	1	0.791	-0.905	4.448
ATOM	19	C8'	NCP	1	-0.037	1.002	3.365
ATOM	20	H8'1	NCP	1	-1.035	0.614	3.201
ATOM	21	H8'2	NCP	1	0.194	1.681	2.567
ATOM	22	N1	NCP	1	-0.029	1.724	4.620
ATOM	23	C2	NCP	1	1.136	2.454	4.913
ATOM	24	O2	NCP	1	2.026	2.428	4.105
ATOM	25	N3	NCP	1	1.179	3.132	6.086
ATOM	26	C4	NCP	1	0.205	3.031	6.938
ATOM	27	N4	NCP	1	0.294	3.745	8.075
ATOM	28	H41	NCP	1	-0.320	3.576	8.836
ATOM	29	H42	NCP	1	1.154	4.212	8.258
ATOM	30	C5	NCP	1	-0.958	2.214	6.718
ATOM	31	H5	NCP	1	-1.738	2.122	7.446
ATOM	32	C6	NCP	1	-1.007	1.578	5.540
ATOM	33	H6	NCP	1	-1.824	0.939	5.267
ATOM	1	N1'	NGP	1	0.000	0.000	0.000
ATOM	2	H1'1	NGP	1	-0.372	-0.941	0.000
ATOM	3	H1'2	NGP	1	-0.372	0.470	0.815
ATOM	4	H1'3	NGP	1	-0.372	0.470	-0.815
ATOM	5	C2'	NGP	1	1.451	0.000	0.000
ATOM	6	H2'1	NGP	1	1.809	0.668	-0.774
ATOM	7	H2'2	NGP	1	1.770	-1.002	-0.259
ATOM	8	C3'	NGP	1	2.074	0.428	1.327
ATOM	9	H3'1	NGP	1	1.817	1.449	1.538
ATOM	10	H3'2	NGP	1	3.152	0.398	1.239
ATOM	11	N4'	NGP	1	1.689	-0.440	2.435
ATOM	12	C5'	NGP	1	2.476	-1.652	2.607
ATOM	13	H5'1	NGP	1	1.855	-2.432	3.018
ATOM	14	H5'2	NGP	1	2.840	-1.969	1.640
ATOM	15	C'	NGP	1	3.703	-1.396	3.483
ATOM	16	O1'	NGP	1	4.698	-0.886	3.020
ATOM	17	C7'	NGP	1	0.899	-0.098	3.479
ATOM	18	O7'	NGP	1	0.765	-0.818	4.437
ATOM	19	C8'	NGP	1	0.108	1.213	3.453
ATOM	20	H8'1	NGP	1	-0.919	0.939	3.649
ATOM	21	H8'2	NGP	1	0.145	1.739	2.515
ATOM	22	N9	NGP	1	0.540	2.080	4.529
ATOM	23	C8	NGP	1	-0.250	2.777	5.417
ATOM	24	H8	NGP	1	-1.315	2.662	5.412
ATOM	25	N7	NGP	1	0.413	3.533	6.204
ATOM	26	C6	NGP	1	2.946	3.911	6.350
ATOM	27	O6	NGP	1	3.140	4.708	7.217
ATOM	28	C5	NGP	1	1.729	3.345	5.839
ATOM	29	C4	NGP	1	1.812	2.452	4.808
ATOM	30	N3	NGP	1	2.910	1.974	4.180
ATOM	31	C2	NGP	1	4.014	2.460	4.644

ATOM	32	N2	NGP	1	5.195	2.073	4.106
ATOM	33	H21	NGP	1	5.155	1.226	3.572
ATOM	34	H22	NGP	1	6.021	2.187	4.649
ATOM	35	N1	NGP	1	4.056	3.374	5.649
ATOM	36	H1	NGP	1	4.933	3.754	5.935
ATOM	1	N1'	NTP	1	0.000	0.000	0.000
ATOM	2	H1'1	NTP	1	-0.372	-0.941	0.000
ATOM	3	H1'2	NTP	1	-0.372	0.470	0.815
ATOM	4	H1'3	NTP	1	-0.372	0.470	-0.815
ATOM	5	C2'	NTP	1	1.452	0.000	0.000
ATOM	6	H2'1	NTP	1	1.811	0.710	-0.734
ATOM	7	H2'2	NTP	1	1.772	-0.986	-0.314
ATOM	8	C3'	NTP	1	2.081	0.353	1.346
ATOM	9	H3'1	NTP	1	1.845	1.367	1.616
ATOM	10	H3'2	NTP	1	3.158	0.304	1.243
ATOM	11	N4'	NTP	1	1.700	-0.561	2.422
ATOM	12	C5'	NTP	1	2.457	-1.807	2.505
ATOM	13	H5'1	NTP	1	1.863	-2.543	3.023
ATOM	14	H5'2	NTP	1	2.663	-2.156	1.504
ATOM	15	C'	NTP	1	3.809	-1.612	3.190
ATOM	16	O1'	NTP	1	4.817	-1.511	2.541
ATOM	17	C7'	NTP	1	0.878	-0.269	3.448
ATOM	18	O7'	NTP	1	0.793	-0.972	4.429
ATOM	19	C8'	NTP	1	-0.057	0.938	3.365
ATOM	20	H8'1	NTP	1	-1.043	0.540	3.162
ATOM	21	H8'2	NTP	1	0.181	1.643	2.592
ATOM	22	N1	NTP	1	-0.104	1.632	4.637
ATOM	23	C2	NTP	1	0.939	2.477	4.914
ATOM	24	O2	NTP	1	1.838	2.665	4.143
ATOM	25	N3	NTP	1	0.871	3.086	6.135
ATOM	26	H3	NTP	1	1.613	3.721	6.343
ATOM	27	C4	NTP	1	-0.072	2.884	7.132
ATOM	28	O4	NTP	1	0.000	3.480	8.165
ATOM	29	C5	NTP	1	-1.106	1.900	6.795
ATOM	30	C6	NTP	1	-1.053	1.330	5.594
ATOM	31	H6	NTP	1	-1.767	0.591	5.291
ATOM	32	C7	NTP	1	-2.148	1.589	7.832
ATOM	33	H71	NTP	1	-1.690	1.211	8.739
ATOM	34	H72	NTP	1	-2.702	2.482	8.102
ATOM	35	H73	NTP	1	-2.848	0.847	7.466
ATOM	1	N1'	TPN	1	-3.164	-2.077	-0.367
ATOM	2	H1'1	TPN	1	-3.506	-2.157	-1.296
ATOM	3	C2'	TPN	1	-3.574	-0.919	0.408
ATOM	4	H2'1	TPN	1	-4.043	-1.249	1.326
ATOM	5	H2'2	TPN	1	-4.324	-0.395	-0.172
ATOM	6	C3'	TPN	1	-2.435	0.029	0.773
ATOM	7	H3'1	TPN	1	-1.736	-0.458	1.430
ATOM	8	H3'2	TPN	1	-2.848	0.863	1.327
ATOM	9	N4'	TPN	1	-1.740	0.577	-0.391
ATOM	10	C5'	TPN	1	-2.365	1.732	-1.029
ATOM	11	H5'1	TPN	1	-2.030	1.790	-2.053
ATOM	12	H5'2	TPN	1	-3.437	1.599	-1.013
ATOM	13	C'	TPN	1	-2.071	3.030	-0.281
ATOM	14	O1'	TPN	1	-2.886	3.507	0.466
ATOM	15	C7'	TPN	1	-0.494	0.253	-0.788
ATOM	16	O7'	TPN	1	0.117	0.907	-1.602
ATOM	17	C8'	TPN	1	0.167	-1.027	-0.277
ATOM	18	H8'1	TPN	1	0.113	-1.739	-1.091
ATOM	19	H8'2	TPN	1	-0.303	-1.469	0.580
ATOM	20	N1	TPN	1	1.566	-0.793	0.023
ATOM	21	C2	TPN	1	1.843	-0.187	1.221
ATOM	22	O2	TPN	1	0.986	0.112	2.005

ATOM	23	N3	TPN	1	3.170	0.045	1.450
ATOM	24	H3	TPN	1	3.390	0.466	2.328
ATOM	25	C4	TPN	1	4.231	-0.185	0.587
ATOM	26	O4	TPN	1	5.349	0.081	0.916
ATOM	27	C5	TPN	1	3.845	-0.752	-0.709
ATOM	28	C6	TPN	1	2.555	-1.003	-0.917
ATOM	29	H6	TPN	1	2.204	-1.401	-1.848
ATOM	30	C7	TPN	1	4.928	-0.998	-1.722
ATOM	31	H71	TPN	1	5.446	-0.077	-1.964
ATOM	32	H72	TPN	1	5.668	-1.689	-1.335
ATOM	33	H73	TPN	1	4.515	-1.411	-2.635
ATOM	1	C4"	DIM	1	4.038	2.897	-0.899
ATOM	2	C5"	DIM	1	5.423	3.134	-0.514
ATOM	3	C6"	DIM	1	5.906	4.407	-0.572
ATOM	4	C2"	DIM	1	3.834	5.358	-1.353
ATOM	5	H3"	DIM	1	2.391	3.938	-1.567
ATOM	6	H6"	DIM	1	6.939	4.582	-0.287
ATOM	7	C7"	DIM	1	6.378	2.073	-0.052
ATOM	8	N2L	DIM	1	5.877	0.811	-0.017
ATOM	9	C0L	DIM	1	6.675	-0.323	0.404
ATOM	10	H0L1	DIM	1	7.574	-0.400	-0.224
ATOM	11	H0L2	DIM	1	7.018	-0.097	1.419
ATOM	12	C7L	DIM	1	5.941	-1.653	0.377
ATOM	13	C6L	DIM	1	4.568	-1.748	0.309
ATOM	14	C8L	DIM	1	6.713	-2.845	0.450
ATOM	15	C5L	DIM	1	3.911	-3.009	0.314
ATOM	16	H6L	DIM	1	3.966	-0.846	0.247
ATOM	17	C9L	DIM	1	6.111	-4.080	0.464
ATOM	18	H8L	DIM	1	7.797	-2.771	0.496
ATOM	19	C4L	DIM	1	2.497	-3.120	0.241
ATOM	20	C10L	DIM	1	4.698	-4.201	0.397
ATOM	21	H9L	DIM	1	6.714	-4.983	0.520
ATOM	22	C3L	DIM	1	1.871	-4.349	0.248
ATOM	23	H4L	DIM	1	1.904	-2.213	0.175
ATOM	24	C11L	DIM	1	4.033	-5.455	0.402
ATOM	25	C12L	DIM	1	2.662	-5.527	0.327
ATOM	26	H11L	DIM	1	4.627	-6.364	0.459
ATOM	27	H12L	DIM	1	2.170	-6.496	0.324
ATOM	28	C2L	DIM	1	0.360	-4.495	0.208
ATOM	29	H2L1	DIM	1	0.104	-5.416	-0.334
ATOM	30	H2L2	DIM	1	-0.044	-4.602	1.220
ATOM	31	N1L	DIM	1	-0.329	-3.366	-0.387
ATOM	32	C7	DIM	1	-1.680	-3.324	-0.506
ATOM	33	C5	DIM	1	-2.233	-2.089	-1.159
ATOM	34	C4	DIM	1	-1.431	-0.975	-1.646
ATOM	35	C6	DIM	1	-3.583	-2.033	-1.313
ATOM	36	H6	DIM	1	-4.179	-2.871	-0.969
ATOM	37	C2	DIM	1	-3.560	0.132	-2.382
ATOM	38	H3	DIM	1	-1.667	0.848	-2.576
ATOM	39	C8'	DIM	1	-5.679	-1.036	-2.106
ATOM	40	H8'1	DIM	1	-6.163	-1.539	-1.265
ATOM	41	H8'2	DIM	1	-6.045	-0.009	-2.161
ATOM	42	C7'	DIM	1	-5.957	-1.769	-3.437
ATOM	43	N4'	DIM	1	-7.266	-1.955	-3.781
ATOM	44	C3'	DIM	1	-8.416	-1.477	-3.004
ATOM	45	H3'1	DIM	1	-9.162	-1.123	-3.719
ATOM	46	H3'2	DIM	1	-8.130	-0.617	-2.396
ATOM	47	C5'	DIM	1	-7.549	-2.630	-5.055
ATOM	48	H5'1	DIM	1	-6.675	-3.217	-5.323
ATOM	49	H5'2	DIM	1	-8.401	-3.301	-4.916
ATOM	50	C'	DIM	1	-7.974	-1.615	-6.130
ATOM	51	C2'	DIM	1	-9.024	-2.575	-2.104

ATOM	52	H2'1	DIM	1	-8.293	-2.908	-1.364
ATOM	53	H2'2	DIM	1	-9.266	-3.453	-2.713
ATOM	54	N1'	DIM	1	-10.223	-2.125	-1.425
ATOM	55	H1'1	DIM	1	-11.071	-2.033	-1.971
ATOM	56	CC1	DIM	1	5.742	6.838	-1.014
ATOM	57	HC1	DIM	1	5.683	7.233	-2.030
ATOM	58	HC2	DIM	1	5.181	7.503	-0.354
ATOM	59	HC3	DIM	1	6.782	6.789	-0.695
ATOM	60	O2"	DIM	1	3.161	6.308	-1.708
ATOM	61	O4"	DIM	1	3.445	1.819	-0.908
ATOM	62	N3"	DIM	1	3.358	4.059	-1.290
ATOM	63	O7"	DIM	1	7.532	2.370	0.263
ATOM	64	H2L	DIM	1	4.914	0.685	-0.320
ATOM	65	H1L	DIM	1	0.196	-2.579	-0.762
ATOM	66	O7	DIM	1	-2.440	-4.214	-0.117
ATOM	67	O4	DIM	1	-0.207	-0.869	-1.607
ATOM	68	N3	DIM	1	-2.191	0.061	-2.210
ATOM	69	O2	DIM	1	-4.125	1.068	-2.917
ATOM	70	N1	DIM	1	-4.249	-0.973	-1.853
ATOM	71	O7'	DIM	1	-5.017	-2.157	-4.125
ATOM	72	O1'	DIM	1	-9.075	-1.072	-6.047
ATOM	73	N1"	DIM	1	5.176	5.487	-0.967
ATOM	1	C4"	NBC	1	4.195	3.338	-0.909
ATOM	2	C5"	NBC	1	4.837	3.370	0.399
ATOM	3	C6"	NBC	1	4.623	4.449	1.203
ATOM	4	C2"	NBC	1	3.178	5.569	-0.369
ATOM	5	H3"	NBC	1	2.943	4.485	-2.075
ATOM	6	H6"	NBC	1	5.102	4.473	2.176
ATOM	7	C7"	NBC	1	5.731	2.295	0.942
ATOM	8	N2L	NBC	1	5.940	1.237	0.116
ATOM	9	C0L	NBC	1	6.781	0.119	0.495
ATOM	10	H0L1	NBC	1	7.687	0.105	-0.128
ATOM	11	H0L2	NBC	1	7.108	0.315	1.521
ATOM	12	C7L	NBC	1	6.102	-1.237	0.405
ATOM	13	C6L	NBC	1	4.735	-1.385	0.322
ATOM	14	C8L	NBC	1	6.922	-2.399	0.434
ATOM	15	C5L	NBC	1	4.130	-2.670	0.268
ATOM	16	H6L	NBC	1	4.097	-0.506	0.294
ATOM	17	C9L	NBC	1	6.370	-3.657	0.390
ATOM	18	H8L	NBC	1	8.001	-2.283	0.492
ATOM	19	C4L	NBC	1	2.722	-2.835	0.179
ATOM	20	C10L	NBC	1	4.964	-3.832	0.307
ATOM	21	H9L	NBC	1	7.009	-4.536	0.413
ATOM	22	C3L	NBC	1	2.146	-4.088	0.129
ATOM	23	H4L	NBC	1	2.093	-1.951	0.147
ATOM	24	C11L	NBC	1	4.351	-5.111	0.253
ATOM	25	C12L	NBC	1	2.984	-5.235	0.164
ATOM	26	H11L	NBC	1	4.981	-5.997	0.276
ATOM	27	H12L	NBC	1	2.532	-6.222	0.116
ATOM	28	C2L	NBC	1	0.643	-4.293	0.071
ATOM	29	H2L1	NBC	1	0.428	-5.199	-0.512
ATOM	30	H2L2	NBC	1	0.237	-4.459	1.074
ATOM	31	N1L	NBC	1	-0.088	-3.168	-0.481
ATOM	32	C7	NBC	1	-1.129	-2.591	0.170
ATOM	33	C5	NBC	1	-1.786	-1.447	-0.546
ATOM	34	C4	NBC	1	-1.379	-0.937	-1.849
ATOM	35	C6	NBC	1	-2.848	-0.867	0.074
ATOM	36	H6	NBC	1	-3.165	-1.260	1.035
ATOM	37	C2	NBC	1	-3.239	0.740	-1.684
ATOM	38	H3	NBC	1	-1.911	0.517	-3.207
ATOM	39	C8'	NBC	1	-4.740	0.685	0.233
ATOM	40	H8'1	NBC	1	-4.587	0.663	1.315

ATOM	41	H8'2	NBC	1	-4.886	1.721	-0.081
ATOM	42	C7'	NBC	1	-5.958	-0.170	-0.182
ATOM	43	N4'	NBC	1	-7.155	0.133	0.398
ATOM	44	C3'	NBC	1	-7.377	1.229	1.349
ATOM	45	H3'1	NBC	1	-8.345	1.673	1.109
ATOM	46	H3'2	NBC	1	-6.628	2.011	1.205
ATOM	47	C5'	NBC	1	-8.329	-0.655	-0.002
ATOM	48	H5'1	NBC	1	-7.975	-1.603	-0.399
ATOM	49	H5'2	NBC	1	-8.943	-0.846	0.883
ATOM	50	C'	NBC	1	-9.215	0.138	-0.977
ATOM	51	C2'	NBC	1	-7.360	0.756	2.818
ATOM	52	H2'1	NBC	1	-6.382	0.343	3.075
ATOM	53	H2'2	NBC	1	-8.081	-0.061	2.947
ATOM	54	N1'	NBC	1	-7.687	1.824	3.743
ATOM	55	H1'1	NBC	1	-8.653	2.125	3.798
ATOM	56	O2"	NBC	1	2.472	6.507	-0.690
ATOM	57	O4"	NBC	1	4.281	2.449	-1.755
ATOM	58	N3"	NBC	1	3.407	4.466	-1.174
ATOM	59	O7"	NBC	1	6.223	2.404	2.067
ATOM	60	H2L	NBC	1	5.496	1.267	-0.798
ATOM	61	H1L	NBC	1	0.149	-2.798	-1.399
ATOM	62	O7	NBC	1	-1.539	-2.949	1.278
ATOM	63	O4	NBC	1	-0.461	-1.351	-2.554
ATOM	64	N3	NBC	1	-2.153	0.145	-2.296
ATOM	65	O2	NBC	1	-3.869	1.660	-2.174
ATOM	66	N1	NBC	1	-3.536	0.202	-0.420
ATOM	67	O7'	NBC	1	-5.813	-1.077	-0.997
ATOM	68	O1'	NBC	1	-9.866	1.096	-0.560
ATOM	69	N1"	NBC	1	3.839	5.511	0.867
ATOM	70	C1A	NBC	1	3.682	6.620	1.820
ATOM	71	H1A1	NBC	1	4.102	6.339	2.763
ATOM	72	C2A	NBC	1	4.404	7.871	1.293
ATOM	73	H2A1	NBC	1	4.397	8.658	2.049
ATOM	74	H2A2	NBC	1	3.890	8.254	0.401
ATOM	75	N3A	NBC	1	5.798	7.606	0.987
ATOM	76	H3A1	NBC	1	5.881	7.317	0.033
ATOM	77	H1A2	NBC	1	2.642	6.836	1.944
ATOM	78	CCC	NBC	1	6.588	8.827	1.199
ATOM	79	C2C	NBC	1	8.061	8.726	1.647
ATOM	80	OCC	NBC	1	6.062	9.956	1.014
ATOM	81	N0C	NBC	1	8.805	9.971	1.355
ATOM	82	H2C1	NBC	1	8.049	8.566	2.731
ATOM	83	H2C2	NBC	1	8.542	7.848	1.200
ATOM	84	C1C	NBC	1	9.448	9.969	0.005
ATOM	85	C3C	NBC	1	9.691	10.451	2.460
ATOM	86	C9C	NBC	1	9.775	11.389	-0.501
ATOM	87	H1C1	NBC	1	10.367	9.368	-0.001
ATOM	88	H1C2	NBC	1	8.744	9.491	-0.686
ATOM	89	C4C	NBC	1	9.010	11.521	3.338
ATOM	90	H3C1	NBC	1	10.604	10.851	2.016
ATOM	91	H3C2	NBC	1	10.010	9.623	3.101
ATOM	92	N8C	NBC	1	8.623	12.350	-0.348
ATOM	93	H9C1	NBC	1	10.638	11.800	0.027
ATOM	94	H9C2	NBC	1	10.063	11.321	-1.554
ATOM	95	N5C	NBC	1	8.354	12.604	2.522
ATOM	96	H4C1	NBC	1	9.754	11.948	4.020
ATOM	97	H4C2	NBC	1	8.224	11.072	3.953
ATOM	98	C7C	NBC	1	8.979	13.658	0.297
ATOM	99	H8C1	NBC	1	8.238	12.553	-1.270
ATOM	100	C6C	NBC	1	9.320	13.499	1.791
ATOM	101	H5C1	NBC	1	7.796	13.170	3.159
ATOM	102	H7C1	NBC	1	8.114	14.317	0.172

ATOM	103	H7C2	NBC	1	9.826	14.130	-0.213
ATOM	104	H6C1	NBC	1	9.328	14.491	2.250
ATOM	105	H6C2	NBC	1	10.327	13.095	1.915
ATOM	106	Zn	NBC	1	7.301	11.490	1.059

8.3.5. pna.amber94.bnd

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8.3.6. pna.amber94.qr

ATOM	1	P	ADE	1	1.166	2.000
ATOM	2	O1P	ADE	1	-0.776	1.400
ATOM	3	O2P	ADE	1	-0.776	1.400
ATOM	4	O5'	ADE	1	-0.495	1.400
ATOM	5	C5'	ADE	1	-0.007	1.700
ATOM	6	H5'1	ADE	1	0.075	1.000
ATOM	7	H5'2	ADE	1	0.075	1.000
ATOM	8	C4'	ADE	1	0.163	1.700
ATOM	9	H4'	ADE	1	0.118	1.000
ATOM	10	O4'	ADE	1	-0.369	1.400

ATOM	11	C1'	ADE	1	0.043	1.700
ATOM	12	H1'	ADE	1	0.184	1.000
ATOM	13	N9	ADE	1	-0.027	1.500
ATOM	14	C8	ADE	1	0.161	1.700
ATOM	15	H8	ADE	1	0.188	1.000
ATOM	16	N7	ADE	1	-0.618	1.500
ATOM	17	C5	ADE	1	0.072	1.700
ATOM	18	C6	ADE	1	0.690	1.700
ATOM	19	N6	ADE	1	-0.912	1.500
ATOM	20	H61	ADE	1	0.417	1.000
ATOM	21	H62	ADE	1	0.417	1.000
ATOM	22	N1	ADE	1	-0.762	1.500
ATOM	23	C2	ADE	1	0.572	1.700
ATOM	24	H2	ADE	1	0.060	1.000
ATOM	25	N3	ADE	1	-0.742	1.500
ATOM	26	C4	ADE	1	0.380	1.700
ATOM	27	C3'	ADE	1	0.071	1.700
ATOM	28	H3'	ADE	1	0.099	1.000
ATOM	29	C2'	ADE	1	-0.085	1.700
ATOM	30	H2'1	ADE	1	0.072	1.000
ATOM	31	H2'2	ADE	1	0.072	1.000
ATOM	32	O3'	ADE	1	-0.523	1.400
ATOM	1	P	GUA	1	1.166	2.000
ATOM	2	O1P	GUA	1	-0.776	1.400
ATOM	3	O2P	GUA	1	-0.776	1.400
ATOM	4	O5'	GUA	1	-0.495	1.400
ATOM	5	C5'	GUA	1	-0.007	1.700
ATOM	6	H5'1	GUA	1	0.075	1.000
ATOM	7	H5'2	GUA	1	0.075	1.000
ATOM	8	C4'	GUA	1	0.163	1.700
ATOM	9	H4'	GUA	1	0.118	1.000
ATOM	10	O4'	GUA	1	-0.369	1.400
ATOM	11	C1'	GUA	1	0.036	1.700
ATOM	12	H1'	GUA	1	0.175	1.000
ATOM	13	N9	GUA	1	0.058	1.500
ATOM	14	C8	GUA	1	0.074	1.700
ATOM	15	H8	GUA	1	0.200	1.000
ATOM	16	N7	GUA	1	-0.573	1.500
ATOM	17	C5	GUA	1	0.199	1.700
ATOM	18	C6	GUA	1	0.492	1.700
ATOM	19	O6	GUA	1	-0.570	1.400
ATOM	20	N1	GUA	1	-0.505	1.500
ATOM	21	H1	GUA	1	0.352	1.000
ATOM	22	C2	GUA	1	0.743	1.700
ATOM	23	N2	GUA	1	-0.923	1.500
ATOM	24	H21	GUA	1	0.423	1.000
ATOM	25	H22	GUA	1	0.423	1.000
ATOM	26	N3	GUA	1	-0.664	1.500
ATOM	27	C4	GUA	1	0.181	1.700
ATOM	28	C3'	GUA	1	0.071	1.700
ATOM	29	H3'	GUA	1	0.099	1.000
ATOM	30	C2'	GUA	1	-0.085	1.700
ATOM	31	H2'1	GUA	1	0.072	1.000
ATOM	32	H2'2	GUA	1	0.072	1.000
ATOM	33	O3'	GUA	1	-0.523	1.400
ATOM	1	P	THY	1	1.166	2.000
ATOM	2	O1P	THY	1	-0.776	1.400
ATOM	3	O2P	THY	1	-0.776	1.400
ATOM	4	O5'	THY	1	-0.495	1.400
ATOM	5	C5'	THY	1	-0.007	1.700
ATOM	6	H5'1	THY	1	0.075	1.000
ATOM	7	H5'2	THY	1	0.075	1.000

ATOM	8	C4'	THY	1	0.163	1.700
ATOM	9	H4'	THY	1	0.118	1.000
ATOM	10	O4'	THY	1	-0.369	1.400
ATOM	11	C1'	THY	1	0.068	1.700
ATOM	12	H1'	THY	1	0.180	1.000
ATOM	13	N1	THY	1	-0.024	1.500
ATOM	14	C6	THY	1	-0.221	1.700
ATOM	15	H6	THY	1	0.261	1.000
ATOM	16	C5	THY	1	0.003	1.700
ATOM	17	C7	THY	1	-0.227	1.700
ATOM	18	H71	THY	1	0.077	1.000
ATOM	19	H72	THY	1	0.077	1.000
ATOM	20	H73	THY	1	0.077	1.000
ATOM	21	C4	THY	1	0.519	1.700
ATOM	22	O4	THY	1	-0.556	1.400
ATOM	23	N3	THY	1	-0.434	1.500
ATOM	24	H3	THY	1	0.342	1.000
ATOM	25	C2	THY	1	0.568	1.700
ATOM	26	O2	THY	1	-0.588	1.400
ATOM	27	C3'	THY	1	0.071	1.700
ATOM	28	H3'	THY	1	0.099	1.000
ATOM	29	C2'	THY	1	-0.085	1.700
ATOM	30	H2'1	THY	1	0.072	1.000
ATOM	31	H2'2	THY	1	0.072	1.000
ATOM	32	O3'	THY	1	-0.523	1.400
ATOM	1	P	CYT	1	1.166	2.000
ATOM	2	O1P	CYT	1	-0.776	1.400
ATOM	3	O2P	CYT	1	-0.776	1.400
ATOM	4	O5'	CYT	1	-0.495	1.400
ATOM	5	C5'	CYT	1	-0.007	1.700
ATOM	6	H5'1	CYT	1	0.075	1.000
ATOM	7	H5'2	CYT	1	0.075	1.000
ATOM	8	C4'	CYT	1	0.163	1.700
ATOM	9	H4'	CYT	1	0.118	1.000
ATOM	10	O4'	CYT	1	-0.369	1.400
ATOM	11	C1'	CYT	1	-0.012	1.700
ATOM	12	H1'	CYT	1	0.196	1.000
ATOM	13	N1	CYT	1	-0.034	1.500
ATOM	14	C6	CYT	1	-0.018	1.700
ATOM	15	H6	CYT	1	0.229	1.000
ATOM	16	C5	CYT	1	-0.522	1.700
ATOM	17	H5	CYT	1	0.186	1.000
ATOM	18	C4	CYT	1	0.844	1.700
ATOM	19	N4	CYT	1	-0.977	1.500
ATOM	20	H41	CYT	1	0.431	1.000
ATOM	21	H42	CYT	1	0.431	1.000
ATOM	22	N3	CYT	1	-0.775	1.500
ATOM	23	C2	CYT	1	0.796	1.700
ATOM	24	O2	CYT	1	-0.655	1.400
ATOM	25	C3'	CYT	1	0.071	1.700
ATOM	26	H3'	CYT	1	0.099	1.000
ATOM	27	C2'	CYT	1	-0.085	1.700
ATOM	28	H2'1	CYT	1	0.072	1.000
ATOM	29	H2'2	CYT	1	0.072	1.000
ATOM	30	O3'	CYT	1	-0.523	1.400
ATOM	1	P	APU	1	1.166	2.000
ATOM	2	O1P	APU	1	-0.776	1.400
ATOM	3	O2P	APU	1	-0.776	1.400
ATOM	4	O5'	APU	1	-0.495	1.400
ATOM	5	C5'	APU	1	-0.007	1.700
ATOM	6	H5'1	APU	1	0.075	1.000
ATOM	7	H5'2	APU	1	0.075	1.000

ATOM	8	C4'	APU	1	0.163	1.700
ATOM	9	H4'	APU	1	0.118	1.000
ATOM	10	O4'	APU	1	-0.369	1.400
ATOM	11	C1'	APU	1	0.068	1.700
ATOM	12	H1'	APU	1	0.180	1.000
ATOM	13	O1'	APU	1	-0.600	1.400
ATOM	14	HO1'	APU	1	0.452	1.000
ATOM	27	C3'	APU	1	0.071	1.700
ATOM	28	H3'	APU	1	0.099	1.000
ATOM	29	C2'	APU	1	-0.085	1.700
ATOM	30	H2'1	APU	1	0.072	1.000
ATOM	31	H2'2	APU	1	0.072	1.000
ATOM	32	O3'	APU	1	-0.523	1.400

8.3.7. pna.amber94.chi

ADE

N9 O4' H1' C2'
 C1' H2'2 H2'1 C3'
 C2' O3' H3' C4'
 C3' O4' H4' C5'
 C4' H5'1 H5'2 O5'
 N9 C1' C8 C6
 C8 N9 N7 H8
 C4 N9 N3 C5
 C5 N7 C4 C6
 C2 N3 N1 H2
 C6 C5 N1 N6
 N6 C6 H61 H62

CYT

N1 O4' H1' C2'
 C1' H2'2 H2'1 C3'
 C2' O3' H3' C4'
 C3' O4' H4' C5'
 C4' H5'1 H5'2 O5'
 N1 C1' C6 C2
 C2 N1 N3 O2
 C4 N3 C5 N4
 N4 C4 H41 H42
 C5 C4 C6 H5
 C6 C5 N1 H6

GUA

N9 O4' H1' C2'
 C1' H2'2 H2'1 C3'
 C2' O3' H3' C4'
 C3' O4' H4' C5'
 C4' H5'1 H5'2 O5'
 N9 C1' C8 C6
 C8 N9 N7 H8
 C4 N9 N3 C5
 C5 N7 C4 C6
 C2 N3 N1 N2
 N2 C2 H21 H22
 C6 C5 N1 O6

THY

N1 O4' H1' C2'
 C1' H2'2 H2'1 C3'
 C2' O3' H3' C4'

C3' O4' H4' C5'
 C4' H5'1 H5'2 O5'
 N1 C1' C6 C2
 C2 N1 N3 O2
 N3 C2 C4 H3
 C4 N3 C5 O4
 C5 C4 C6 C7
 C6 C5 N1 H6
 # APU

8.3.8. apna.dat file

P	8.92	69.20	-4.0600	A
O1P	7.69	73.90	-4.5300	A
O2P	9.96	67.90	-5.1000	A
O3'	9.58	41.43	-5.4300	A
O5'	8.70	60.10	-3.3300	A
C1'	8.59	35.9	-2.04	A
C2'	8.81	33.4	-3.50	A
C3'	8.91	42.2	-4.17	A
C4'	9.77	46.6	-3.13	A
C5'	9.91	55.4	-3.1900	A
O1'	9.2200	44.00	-1.8600	A
N6	3.19	14.8	-0.13	A
C6	4.49	16.7	-0.43	A
C5	5.22	29.8	-0.91	A
N7	5.18	44.8	-1.19	A
C8	6.40	45.7	-1.61	A
N9	7.16	36.1	-1.62	A
C4	6.52	26.7	-1.17	A
N3	7.25	17.3	-1.02	A
C2	6.65	8.4	-0.56	A
N1	5.40	5.3	-0.26	A
P	8.92	69.20	-4.0600	G
O1P	7.69	73.90	-4.5300	G
O2P	9.96	67.90	-5.1000	G
O3'	9.58	41.43	-5.4300	G
O5'	8.70	60.10	-3.3300	G
C1'	8.59	35.9	-2.04	G
C2'	8.81	33.4	-3.50	G
C3'	8.91	42.2	-4.17	G
C4'	9.77	46.6	-3.13	G
C5'	9.91	55.4	-3.1900	G
O1'	9.2200	44.00	-1.8600	G
N2	7.71	0.7	-0.37	G
O6	3.27	13.2	-0.11	G
C6	4.44	16.4	-0.41	G
C5	5.20	29.4	-0.90	G
N7	5.14	44.6	-1.17	G
C8	6.38	45.7	-1.60	G
N9	7.16	36.1	-1.62	G
C4	6.52	26.6	-1.17	G
N3	7.30	17.4	-1.03	G
C2	6.78	8.1	-0.57	G
N1	5.46	5.3	-0.27	G
P	8.92	69.20	-4.0600	T
O1P	7.69	73.90	-4.5300	T
O2P	9.96	67.90	-5.1000	T

O3'	9.58	41.43	-5.4300	T
O5'	8.70	60.10	-3.3300	T
C1'	8.59	35.9	-2.04	T
C2'	8.81	33.4	-3.50	T
C3'	8.91	42.2	-4.17	T
C4'	9.77	46.6	-3.13	T
C5'	9.91	55.4	-3.1900	T
O1'	9.2200	44.00	-1.8600	T
O4	3.30	35.5	-0.47	T
C4	4.48	36.4	-0.83	T
N3	5.39	24.3	-0.82	T
C2	6.71	26.0	-1.20	T
O2	7.56	19.0	-1.15	T
N1	7.16	36.1	-1.62	T
C6	6.55	46.4	-1.67	T
C5	5.30	49.3	-1.30	T
C7	4.98	65.7	-1.33	T
P	8.92	69.20	-4.0600	C
O1P	7.69	73.90	-4.5300	C
O2P	9.96	67.90	-5.1000	C
O3'	9.58	41.43	-5.4300	C
O5'	8.70	60.10	-3.3300	C
C1'	8.59	35.9	-2.04	C
C2'	8.81	33.4	-3.50	C
C3'	8.91	42.2	-4.17	C
C4'	9.77	46.6	-3.13	C
C5'	9.91	55.4	-3.1900	C
O1'	9.2200	44.00	-1.8600	C
N4	3.30	34.5	-0.46	C
C4	4.57	35.5	-0.84	C
N3	5.41	23.4	-0.80	C
C2	6.69	25.9	-1.19	C
O2	7.61	19.2	-1.17	C
N1	7.16	36.1	-1.62	C
C6	6.55	46.3	-1.66	C
C5	5.28	49.1	-1.29	C
N6	3.19	14.8	-0.1300	X
C6	4.49	16.7	-0.4300	X
C5	5.22	29.8	-0.9100	X
N7	5.18	44.8	-1.1900	X
C8	6.40	45.7	-1.6100	X
N9	7.16	36.1	-1.6200	X
C4	6.52	26.7	-1.1700	X
N3	7.25	17.3	-1.0200	X
C2	6.65	8.4	-0.5600	X
N1	5.40	5.3	-0.2600	X
O4	3.30	35.5	-0.4700	Y
C4	4.48	36.4	-0.8300	Y
N3	5.39	24.3	-0.8200	Y
C2	6.71	26.0	-1.2000	Y
O2	7.56	19.0	-1.1500	Y
N1	7.16	36.1	-1.6200	Y
C6	6.55	46.4	-1.6700	Y
C5	5.30	49.3	-1.3000	Y
C7	4.98	65.7	-1.3300	Y
N4	3.30	34.5	-0.4600	J
C4	4.57	35.5	-0.8400	J
N3	5.41	23.4	-0.8000	J
C2	6.69	25.9	-1.1900	J
O2	7.61	19.2	-1.1700	J
N1	7.16	36.1	-1.6200	J
C6	6.55	46.3	-1.6600	J

C5	5.28	49.1	-1.2900	J
N2	7.71	0.7	-0.3700	K
O6	3.27	13.2	-0.1100	K
C6	4.44	16.4	-0.4100	K
C5	5.20	29.4	-0.9000	K
N7	5.14	44.6	-1.1700	K
C8	6.38	45.7	-1.6000	K
N9	7.16	36.1	-1.6200	K
C4	6.52	26.6	-1.1700	K
N3	7.30	17.4	-1.0300	K
C2	6.78	8.1	-0.5700	K
N1	5.46	5.3	-0.2700	K
O4	3.30	35.5	-0.4700	W
C4	4.48	36.4	-0.8300	W
N3	5.39	24.3	-0.8200	W
C2	6.71	26.0	-1.2000	W
O2	7.56	19.0	-1.1500	W
N1	7.16	36.1	-1.6200	W
C6	6.55	46.4	-1.6700	W
C5	5.30	49.3	-1.3000	W
C7	4.98	65.7	-1.3300	W
O4	3.30	35.5	-0.4700	Q
C4	4.48	36.4	-0.8300	Q
N3	5.39	24.3	-0.8200	Q
C2	6.71	26.0	-1.2000	Q
O2	7.56	19.0	-1.1500	Q
N1	7.16	36.1	-1.6200	Q
C6	6.55	46.4	-1.6700	Q
C5	5.30	49.3	-1.3000	Q
C7	4.98	65.7	-1.3300	Q

8.3.9. leaprc.pna

```

logfile leap.log
#
# ----- leaprc for loading the ff10 force field
# ----- NOTE: this is designed for PDB format 3!
# ff10 = ff99SB for proteins; ff99bsc0 for DNA; ff99sbsc_chiOL3 for RNA
# phosphoaa parms from N. Homeyer, A.H.C. Horn, H. Lanig, H. Sticht
# J. Mol. Model. 2006, 12, 281-289. OP vdW parameters modified
# by T. Steinbrecher and J. Latzer.
#
# load atom type hybridizations
#
addAtomTypes {
  { "H" "H" "sp3" }
  { "HO" "H" "sp3" }
  { "HS" "H" "sp3" }
  { "H1" "H" "sp3" }
  { "H2" "H" "sp3" }
  { "H3" "H" "sp3" }
  { "H4" "H" "sp3" }
  { "H5" "H" "sp3" }
  { "HW" "H" "sp3" }
  { "HC" "H" "sp3" }
  { "HA" "H" "sp3" }
  { "HP" "H" "sp3" }
  { "OH" "O" "sp3" }
}

```

```

{ "OS" "O" "sp3" }
{ "O" "O" "sp2" }
{ "O2" "O" "sp2" }
{ "OW" "O" "sp3" }
{ "CT" "C" "sp3" }
{ "CH" "C" "sp3" }
{ "C2" "C" "sp3" }
{ "C3" "C" "sp3" }
{ "C" "C" "sp2" }
{ "C*" "C" "sp2" }
{ "CA" "C" "sp2" }
{ "CB" "C" "sp2" }
{ "CC" "C" "sp2" }
{ "CN" "C" "sp2" }
{ "CM" "C" "sp2" }
{ "CK" "C" "sp2" }
{ "CQ" "C" "sp2" }
{ "CD" "C" "sp2" }
{ "CE" "C" "sp2" }
{ "CF" "C" "sp2" }
{ "CP" "C" "sp2" }
{ "CI" "C" "sp2" }
{ "CJ" "C" "sp2" }
{ "CW" "C" "sp2" }
{ "CV" "C" "sp2" }
{ "CR" "C" "sp2" }
{ "CA" "C" "sp2" }
{ "CY" "C" "sp2" }
{ "CO" "C" "sp2" }
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{ "N" "N" "sp2" }
{ "NA" "N" "sp2" }
{ "N2" "N" "sp2" }
{ "N*" "N" "sp2" }
{ "NP" "N" "sp2" }
{ "NQ" "N" "sp2" }
{ "NB" "N" "sp2" }
{ "NC" "N" "sp2" }
{ "NT" "N" "sp3" }
{ "N3" "N" "sp3" }
{ "S" "S" "sp3" }
{ "SH" "S" "sp3" }
{ "P" "P" "sp3" }
{ "LP" "" "sp3" }
{ "F" "F" "sp3" }
{ "CL" "Cl" "sp3" }
{ "BR" "Br" "sp3" }
{ "I" "I" "sp3" }
{ "FE" "Fe" "sp3" }
{ "EP" "" "sp3" }

```

```

}
```

```

#
# Load the main parameter set.
#
parm10 = loadamberparams parm10.dat
FRCMOD = loadAmberParams frcmmod.pna

```

```

source leaprc.ff99bsc0
frcmod14SB = loadamberparams frcmod.ff14SB
source leaprc.gaff
#
#      Load main chain and terminating amino acid libraries, nucleic acids
#
loadOff amino10.lib
loadOff aminoct10.lib
loadOff aminont10.lib
loadOff phosphoaa10.lib
loadOff nucleic10.lib
loadOff RNA_CI.lib

#
#      Load water and ions
#
loadOff ions94.lib
loadOff solvents.lib
loadOff atomic_ions.lib
loadOff solvents.lib
HOH = TP3
WAT = TP3

# PNA residue

APN = loadmol3 tripos1-mol3.mol2
NAP = loadmol3 tripos2-mol3.mol2
CAP = loadmol3 tripos3-mol3.mol2
MAPN = loadmol3 tripos4-mol3.mol2

CPN = loadmol3 tripos5-mol3.mol2
NCP = loadmol3 tripos6-mol3.mol2
CCP = loadmol3 tripos7-mol3.mol2
MCPN = loadmol3 tripos8-mol3.mol2

GPN = loadmol3 tripos9-mol3.mol2
NGP = loadmol3 tripos10-mol3.mol2
CGP = loadmol3 tripos11-mol3.mol2
MGPN = loadmol3 tripos12-mol3.mol2

IPN = loadmol3 tripos13-mol3.mol2
NIP = loadmol3 tripos14-mol3.mol2
CIP = loadmol3 tripos15-mol3.mol2
MIPN = loadmol3 tripos16-mol3.mol2

TPN = loadmol3 tripos17-mol3.mol2
NTP = loadmol3 tripos18-mol3.mol2
CTP = loadmol3 tripos19-mol3.mol2
MTP = loadmol3 tripos20-mol3.mol2

UPN = loadmol3 tripos21-mol3.mol2
NUP = loadmol3 tripos22-mol3.mol2
CUP = loadmol3 tripos23-mol3.mol2
MUPN = loadmol3 tripos24-mol3.mol2

DIM = loadmol3 dimeric_charged.mol2

NBC = loadmol3 NBC.mol2

charge APN
charge CPN

```

charge GPN
charge IPN
charge TPN
charge UPN

charge NAP
charge NCP
charge NGP
charge NIP
charge NTP
charge NUP

charge CAP
charge CCP
charge CGP
charge CIP
charge CTP
charge CUP

charge MAPN
charge MCPN
charge MGPN
charge MIPN
charge MTPN
charge MUPN

charge DIM

charge NBC

Define the PDB name map for the amino acids and nucleic acids
#

```
addPdbResMap {  
  { 0 "ALA" "NALA" } { 1 "ALA" "CALA" }  
  { 0 "ARG" "NARG" } { 1 "ARG" "CARG" }  
  { 0 "ASN" "NASN" } { 1 "ASN" "CASN" }  
  { 0 "ASP" "NASP" } { 1 "ASP" "CASP" }  
  { 0 "CYS" "NCYS" } { 1 "CYS" "CCYS" }  
  { 0 "CYX" "NCYX" } { 1 "CYX" "CCYX" }  
  { 0 "GLN" "NGLN" } { 1 "GLN" "CGLN" }  
  { 0 "GLU" "NGLU" } { 1 "GLU" "CGLU" }  
  { 0 "GLY" "NGLY" } { 1 "GLY" "CGLY" }  
  { 0 "HID" "NHID" } { 1 "HID" "CHID" }  
  { 0 "HIE" "NHIE" } { 1 "HIE" "CHIE" }  
  { 0 "HIP" "NHIP" } { 1 "HIP" "CHIP" }  
  { 0 "ILE" "NILE" } { 1 "ILE" "CILE" }  
  { 0 "LEU" "NLEU" } { 1 "LEU" "CLEU" }  
  { 0 "LYS" "NLYS" } { 1 "LYS" "CLYS" }  
  { 0 "MET" "NMET" } { 1 "MET" "CMET" }  
  { 0 "PHE" "NPHE" } { 1 "PHE" "CPHE" }  
  { 0 "PRO" "NPRO" } { 1 "PRO" "CPRO" }  
  { 0 "SER" "NSER" } { 1 "SER" "CSER" }  
  { 0 "THR" "NTHR" } { 1 "THR" "CTHR" }  
  { 0 "TRP" "NTRP" } { 1 "TRP" "CTRP" }  
  { 0 "TYR" "NTYR" } { 1 "TYR" "CTYR" }  
  { 0 "VAL" "NVAL" } { 1 "VAL" "CVAL" }  
  { 0 "HIS" "NHIS" } { 1 "HIS" "CHIS" }  
  { 0 "G" "G5" } { 1 "G" "G3" }  
  { 0 "A" "A5" } { 1 "A" "A3" }  
  { 0 "C" "C5" } { 1 "C" "C3" }  
  { 0 "U" "U5" } { 1 "U" "U3" }
```

```

{ 0 "DG" "DG5" } { 1 "DG" "DG3" }
{ 0 "DA" "DA5" } { 1 "DA" "DA3" }
{ 0 "DC" "DC5" } { 1 "DC" "DC3" }
{ 0 "DT" "DT5" } { 1 "DT" "DT3" }
# some old Amber residue names for RNA:
{ 0 "RA5" "A5" } { 1 "RA3" "A3" } {"RA" "A" }
{ 0 "RC5" "C5" } { 1 "RC3" "C3" } {"RC" "C" }
{ 0 "RG5" "G5" } { 1 "RG3" "G3" } {"RG" "G" }
{ 0 "RU5" "U5" } { 1 "RU3" "U3" } {"RU" "U" }
# some really old Amber residue names, assuming DNA:
{ 0 "GUA" "DG5" } { 1 "GUA" "DG3" } { "GUA" "DG" }
{ 0 "ADE" "DA5" } { 1 "ADE" "DA3" } { "ADE" "DA" }
{ 0 "CYT" "DC5" } { 1 "CYT" "DC3" } { "CYT" "DC" }
{ 0 "THY" "DT5" } { 1 "THY" "DT3" } { "THY" "DT" }
# uncomment out the following if you have this old style RNA files:
# { 0 "GUA" "G5" } { 1 "GUA" "G3" } { "GUA" "G" }
# { 0 "ADE" "A5" } { 1 "ADE" "A3" } { "ADE" "A" }
# { 0 "CYT" "C5" } { 1 "CYT" "C3" } { "CYT" "C" }
# { 0 "THY" "T5" } { 1 "THY" "T3" } { "THY" "T" }

{ 0 "APN" "NAP" } { 1 "APN" "CAP" }
{ 0 "CPN" "NCP" } { 1 "CPN" "CCP" }
{ 0 "GPN" "NGP" } { 1 "GPN" "CGP" }
{ 0 "IPN" "NIP" } { 1 "IPN" "CIP" }
{ 0 "TPN" "NTP" } { 1 "TPN" "CTP" }
{ 0 "UPN" "NUP" } { 1 "UPN" "CUP" }

}

# try to be good about reading in really old atom names as well:
addPdbAtomMap {
{ "O5*" "O5' " }
{ "C5*" "C5' " }
{ "C4*" "C4' " }
{ "O4*" "O4' " }
{ "C3*" "C3' " }
{ "O3*" "O3' " }
{ "C2*" "C2' " }
{ "O2*" "O2' " }
{ "C1*" "C1' " }
{ "C5M" "C7" }
{ "H1*" "H1' " }
{ "H2*1" "H2' " }
{ "H2*2" "H2' " }
{ "H2'1" "H2' " }
{ "H2'2" "H2' " }
{ "H3*" "H3' " }
{ "H4*" "H4' " }
{ "H5*1" "H5' " }
{ "H5*2" "H5' " }
{ "H5'1" "H5' " }
{ "H5'2" "H5' " }
{ "HO'2" "HO2' " }
{ "H5T" "HO5' " }
{ "H3T" "HO3' " }
{ "O1' " "O4' " }
{ "OA" "OP1" }
{ "OB" "OP2" }
{ "O1P" "OP1" }
{ "O2P" "OP2" }
}

```

```

#
# assume that most often proteins use HIE
#
NHIS = NHIE
HIS = HIE
CHIS = CHIE

```

8.3.10.script.sh

```
logfile 0PNA-from-mol3-files.log
```

```
verbosity 2
```

```

addAtomTypes {
  { "H" "H" "sp3" }
  { "HO" "H" "sp3" }
  { "HS" "H" "sp3" }
  { "H1" "H" "sp3" }
  { "H2" "H" "sp3" }
  { "H3" "H" "sp3" }
  { "H4" "H" "sp3" }
  { "H5" "H" "sp3" }
  { "HW" "H" "sp3" }
  { "HC" "H" "sp3" }
  { "HA" "H" "sp3" }
  { "HP" "H" "sp3" }
  { "OH" "O" "sp3" }
  { "OS" "O" "sp3" }
  { "O" "O" "sp2" }
  { "O2" "O" "sp2" }
  { "OW" "O" "sp3" }
  { "CT" "C" "sp3" }
  { "CH" "C" "sp3" }
  { "C2" "C" "sp3" }
  { "C3" "C" "sp3" }
  { "C" "C" "sp2" }
  { "C*" "C" "sp2" }
  { "CA" "C" "sp2" }
  { "CB" "C" "sp2" }
  { "CC" "C" "sp2" }
  { "CN" "C" "sp2" }
  { "CM" "C" "sp2" }
  { "CK" "C" "sp2" }
  { "CQ" "C" "sp2" }
  { "CD" "C" "sp2" }
  { "CE" "C" "sp2" }
  { "CF" "C" "sp2" }
  { "CP" "C" "sp2" }
  { "CI" "C" "sp2" }
  { "CJ" "C" "sp2" }
  { "CW" "C" "sp2" }
}

```

```

    { "CV" "C" "sp2" }
    { "CR" "C" "sp2" }
    { "CA" "C" "sp2" }
    { "CY" "C" "sp2" }
    { "C0" "C" "sp2" }
    { "MG" "Mg" "sp3" }
    { "N" "N" "sp2" }
    { "NA" "N" "sp2" }
    { "N2" "N" "sp2" }
    { "N*" "N" "sp2" }
    { "NP" "N" "sp2" }
    { "NQ" "N" "sp2" }
    { "NB" "N" "sp2" }
    { "NC" "N" "sp2" }
    { "NT" "N" "sp3" }
    { "N3" "N" "sp3" }
    { "S" "S" "sp3" }
    { "SH" "S" "sp3" }
    { "P" "P" "sp3" }
    { "LP" "" "sp3" }
    { "F" "F" "sp3" }
    { "CL" "Cl" "sp3" }
    { "BR" "Br" "sp3" }
    { "I" "I" "sp3" }
    { "FE" "Fe" "sp3" }
    { "EP" "" "sp3" }
}

```

```

addPdbResMap {
  { 0 "APN" "NAP" } { 1 "APN" "CAP" }
  { 0 "CPN" "NCP" } { 1 "CPN" "CCP" }
  { 0 "GPN" "NGP" } { 1 "GPN" "CGP" }
  { 0 "IPN" "NIP" } { 1 "IPN" "CIP" }
  { 0 "TPN" "NTP" } { 1 "TPN" "CTP" }
  { 0 "UPN" "NUP" } { 1 "UPN" "CUP" }
}

```

```

#parm99 = loadamberparams parm99.dat
#frcmod99SB = loadamberparams frcmod.ff99SB
#FRCMOD = loadAmberParams frcmod.pna
FRCMOD = loadAmberParams dimeric.frcmod
source leaprc.ff99bsc0
source leaprc.gaff
source leaprc.pna
#frcmodbsc0 = loadamberparams frcmod.parmbsc0

```

```

loadOff ions94.lib
loadOff solvents.lib
loadOff RNA_CI.lib
HOH = TP3
WAT = TP3

```

```

# See http://q4md-forcefieldtools.org/Tutorial/leap-mol3.php
APN = loadmol3 tripos1-mol3.mol2

```

```
NAP = loadmol3 tripos2-mol3.mol2
CAP = loadmol3 tripos3-mol3.mol2
MAPN = loadmol3 tripos4-mol3.mol2

CPN = loadmol3 tripos5-mol3.mol2
NCP = loadmol3 tripos6-mol3.mol2
CCP = loadmol3 tripos7-mol3.mol2
MCPN = loadmol3 tripos8-mol3.mol2

GPN = loadmol3 tripos9-mol3.mol2
NGP = loadmol3 tripos10-mol3.mol2
CGP = loadmol3 tripos11-mol3.mol2
MGPN = loadmol3 tripos12-mol3.mol2

IPN = loadmol3 tripos13-mol3.mol2
NIP = loadmol3 tripos14-mol3.mol2
CIP = loadmol3 tripos15-mol3.mol2
MIPN = loadmol3 tripos16-mol3.mol2

TPN = loadmol3 tripos17-mol3.mol2
NTP = loadmol3 tripos18-mol3.mol2
CTP = loadmol3 tripos19-mol3.mol2
MTP = loadmol3 tripos20-mol3.mol2

UPN = loadmol3 tripos21-mol3.mol2
NUP = loadmol3 tripos22-mol3.mol2
CUP = loadmol3 tripos23-mol3.mol2
MUPN = loadmol3 tripos24-mol3.mol2

DIM = loadmol3 dimeric_charged.mol2

charge APN
charge CPN
charge GPN
charge IPN
charge TPN
charge UPN

charge NAP
charge NCP
charge NGP
charge NIP
charge NTP
charge NUP

charge CAP
charge CCP
charge CGP
charge CIP
charge CTP
charge CUP

charge MAPN
charge MCPN
```

```

charge MGNP
charge MIPN
charge MTPN
charge MUPN

charge DIM

DUPLEX = loadpdb duplex.pdb
check DUPLEX
solvatebox DUPLEX TIP3PBOX 12.0
saveAmberParm DUPLEX DUPLEX.top DUPLEX.crd
savepdb DUPLEX DUPLEX.pdb
quit

```

8.3.11.script2.sh

```
logfile 0PNA-from-mol3-files.log
```

```
verbosity 2
```

```

addAtomTypes {
  { "H" "H" "sp3 " }
  { "HO" "H" "sp3 " }
  { "HS" "H" "sp3 " }
  { "H1" "H" "sp3 " }
  { "H2" "H" "sp3 " }
  { "H3" "H" "sp3 " }
  { "H4" "H" "sp3 " }
  { "H5" "H" "sp3 " }
  { "HW" "H" "sp3 " }
  { "HC" "H" "sp3 " }
  { "HA" "H" "sp3 " }
  { "HP" "H" "sp3 " }
  { "OH" "O" "sp3 " }
  { "OS" "O" "sp3 " }
  { "O" "O" "sp2 " }
  { "O2" "O" "sp2 " }
  { "OW" "O" "sp3 " }
  { "CT" "C" "sp3 " }
  { "CH" "C" "sp3 " }
  { "C2" "C" "sp3 " }
  { "C3" "C" "sp3 " }
  { "C" "C" "sp2 " }
  { "C*" "C" "sp2 " }
  { "CA" "C" "sp2 " }
  { "CB" "C" "sp2 " }
  { "CC" "C" "sp2 " }
  { "CN" "C" "sp2 " }
  { "CM" "C" "sp2 " }
  { "CK" "C" "sp2 " }
  { "CQ" "C" "sp2 " }
  { "CD" "C" "sp2 " }
  { "CE" "C" "sp2 " }
  { "CF" "C" "sp2 " }
  { "CP" "C" "sp2 " }
  { "CI" "C" "sp2 " }
  { "CJ" "C" "sp2 " }
}

```

```

{ "CW" "C" "sp2" }
{ "CV" "C" "sp2" }
{ "CR" "C" "sp2" }
{ "CA" "C" "sp2" }
{ "CY" "C" "sp2" }
{ "CO" "C" "sp2" }
{ "MG" "Mg" "sp3" }
{ "N" "N" "sp2" }
{ "NA" "N" "sp2" }
{ "N2" "N" "sp2" }
{ "N*" "N" "sp2" }
{ "NP" "N" "sp2" }
{ "NQ" "N" "sp2" }
{ "NB" "N" "sp2" }
{ "NC" "N" "sp2" }
{ "NT" "N" "sp3" }
{ "N3" "N" "sp3" }
{ "S" "S" "sp3" }
{ "SH" "S" "sp3" }
{ "P" "P" "sp3" }
{ "LP" "" "sp3" }
{ "F" "F" "sp3" }
{ "CL" "Cl" "sp3" }
{ "BR" "Br" "sp3" }
{ "I" "I" "sp3" }
{ "FE" "Fe" "sp3" }
{ "EP" "" "sp3" }
}

```

```

addPdbResMap {
  { 0 "APN" "NAP" } { 1 "APN" "CAP" }
  { 0 "CPN" "NCP" } { 1 "CPN" "CCP" }
  { 0 "GPN" "NGP" } { 1 "GPN" "CGP" }
  { 0 "IPN" "NIP" } { 1 "IPN" "CIP" }
  { 0 "TPN" "NTP" } { 1 "TPN" "CTP" }
  { 0 "UPN" "NUP" } { 1 "UPN" "CUP" }
}

```

```

#parm99 = loadamberparams parm99.dat
#frcmod99SB = loadamberparams frcmod.ff99SB
#FRCMOD = loadAmberParams frcmod.pna
FRCMOD = loadAmberParams dimeric.frcmod
source leaprc.ff99bsc0
source leaprc.gaff
source leaprc.pna
#frcmodbsc0 = loadamberparams frcmod.parmbsc0

```

```

loadOff ions94.lib
loadOff solvents.lib
loadOff RNA_CI.lib
HOH = TP3
WAT = TP3

```

```

# See http://q4md-forcefieldtools.org/Tutorial/leap-mol3.php
APN = loadmol3 tripos1-mol3.mol2
NAP = loadmol3 tripos2-mol3.mol2
CAP = loadmol3 tripos3-mol3.mol2
MAPN = loadmol3 tripos4-mol3.mol2

CPN = loadmol3 tripos5-mol3.mol2
NCP = loadmol3 tripos6-mol3.mol2
CCP = loadmol3 tripos7-mol3.mol2

```

```

MCPN = loadmol3 tripos8-mol3.mol2

GPN = loadmol3 tripos9-mol3.mol2
NGP = loadmol3 tripos10-mol3.mol2
CGP = loadmol3 tripos11-mol3.mol2
MGPN = loadmol3 tripos12-mol3.mol2

IPN = loadmol3 tripos13-mol3.mol2
NIP = loadmol3 tripos14-mol3.mol2
CIP = loadmol3 tripos15-mol3.mol2
MIPN = loadmol3 tripos16-mol3.mol2

TPN = loadmol3 tripos17-mol3.mol2
NTP = loadmol3 tripos18-mol3.mol2
CTP = loadmol3 tripos19-mol3.mol2
MTP = loadmol3 tripos20-mol3.mol2

UPN = loadmol3 tripos21-mol3.mol2
NUP = loadmol3 tripos22-mol3.mol2
CUP = loadmol3 tripos23-mol3.mol2
MUPN = loadmol3 tripos24-mol3.mol2

DIM = loadmol3 dimeric_charged.mol2

charge APN
charge CPN
charge GPN
charge IPN
charge TPN
charge UPN

charge NAP
charge NCP
charge NGP
charge NIP
charge NTP
charge NUP

charge CAP
charge CCP
charge CGP
charge CIP
charge CTP
charge CUP

charge MAPN
charge MCPN
charge MGPN
charge MIPN
charge MTPN
charge MUPN

charge DIM

DUPLEX = loadpdb duplex.pdb
check DUPLEX
#solvatebox DUPLEX TIP3PBOX 12.0
saveAmberParm DUPLEX DUPLEX.top DUPLEX.crd
savepdb DUPLEX DUPLEX.pdb
quit

```

8.3.12.script3.sh

```
logfile 0PNA-from-mol3-files.log
```

```
verbosity 2
```

```
addAtomTypes {  
  { "H" "H" "sp3" }  
  { "HO" "H" "sp3" }  
  { "HS" "H" "sp3" }  
  { "H1" "H" "sp3" }  
  { "H2" "H" "sp3" }  
  { "H3" "H" "sp3" }  
  { "H4" "H" "sp3" }  
  { "H5" "H" "sp3" }  
  { "HW" "H" "sp3" }  
  { "HC" "H" "sp3" }  
  { "HA" "H" "sp3" }  
  { "HP" "H" "sp3" }  
  { "OH" "O" "sp3" }  
  { "OS" "O" "sp3" }  
  { "O" "O" "sp2" }  
  { "O2" "O" "sp2" }  
  { "OW" "O" "sp3" }  
  { "CT" "C" "sp3" }  
  { "CH" "C" "sp3" }  
  { "C2" "C" "sp3" }  
  { "C3" "C" "sp3" }  
  { "C" "C" "sp2" }  
  { "C*" "C" "sp2" }  
  { "CA" "C" "sp2" }  
  { "CB" "C" "sp2" }  
  { "CC" "C" "sp2" }  
  { "CN" "C" "sp2" }  
  { "CM" "C" "sp2" }  
  { "CK" "C" "sp2" }  
  { "CQ" "C" "sp2" }  
  { "CD" "C" "sp2" }  
  { "CE" "C" "sp2" }  
  { "CF" "C" "sp2" }  
  { "CP" "C" "sp2" }  
  { "CI" "C" "sp2" }  
  { "CJ" "C" "sp2" }  
  { "CW" "C" "sp2" }  
  { "CV" "C" "sp2" }  
  { "CR" "C" "sp2" }  
  { "CA" "C" "sp2" }  
  { "CY" "C" "sp2" }  
  { "C0" "C" "sp2" }  
  { "MG" "Mg" "sp3" }  
  { "N" "N" "sp2" }  
  { "NA" "N" "sp2" }  
  { "N2" "N" "sp2" }  
  { "N*" "N" "sp2" }  
  { "NP" "N" "sp2" }  
  { "NQ" "N" "sp2" }  
  { "NB" "N" "sp2" }  
  { "NC" "N" "sp2" }  
  { "NT" "N" "sp3" }  
  { "N3" "N" "sp3" }  
  { "S" "S" "sp3" }  
}
```

```

    { "SH" "S" "sp3" }
    { "P" "P" "sp3" }
    { "LP" "" "sp3" }
    { "F" "F" "sp3" }
    { "CL" "Cl" "sp3" }
    { "BR" "Br" "sp3" }
    { "I" "I" "sp3" }
    { "FE" "Fe" "sp3" }
    { "EP" "" "sp3" }
}

addPdbResMap {
  { 0 "APN" "NAP" } { 1 "APN" "CAP" }
  { 0 "CPN" "NCP" } { 1 "CPN" "CCP" }
  { 0 "GPN" "NGP" } { 1 "GPN" "CGP" }
  { 0 "IPN" "NIP" } { 1 "IPN" "CIP" }
  { 0 "TPN" "NTP" } { 1 "TPN" "CTP" }
  { 0 "UPN" "NUP" } { 1 "UPN" "CUP" }
}

#parm99 = loadamberparams parm99.dat
#frcmod99SB = loadamberparams frcmod.ff99SB
#FRCMOD = loadAmberParams frcmod.pna
FRCMOD = loadAmberParams dimeric.frcmod
source leaprc.ff99bsc0
source leaprc.gaff
source leaprc.pna
#frcmodbsc0 = loadamberparams frcmod.parmbsc0

loadOff ions94.lib
loadOff solvents.lib
loadOff RNA_CI.lib
HOH = TP3
WAT = TP3

# See http://q4md-forcefieldtools.org/Tutorial/leap-mol3.php
APN = loadmol3 tripos1-mol3.mol2
NAP = loadmol3 tripos2-mol3.mol2
CAP = loadmol3 tripos3-mol3.mol2
MAPN = loadmol3 tripos4-mol3.mol2

CPN = loadmol3 tripos5-mol3.mol2
NCP = loadmol3 tripos6-mol3.mol2
CCP = loadmol3 tripos7-mol3.mol2
MCPN = loadmol3 tripos8-mol3.mol2

GPN = loadmol3 tripos9-mol3.mol2
NGP = loadmol3 tripos10-mol3.mol2
CGP = loadmol3 tripos11-mol3.mol2
MGPN = loadmol3 tripos12-mol3.mol2

IPN = loadmol3 tripos13-mol3.mol2
NIP = loadmol3 tripos14-mol3.mol2
CIP = loadmol3 tripos15-mol3.mol2
MIPN = loadmol3 tripos16-mol3.mol2

TPN = loadmol3 tripos17-mol3.mol2
NTP = loadmol3 tripos18-mol3.mol2
CTP = loadmol3 tripos19-mol3.mol2
MTP = loadmol3 tripos20-mol3.mol2

UPN = loadmol3 tripos21-mol3.mol2

```

```
NUP = loadmol3 tripos22-mol3.mol2
CUP = loadmol3 tripos23-mol3.mol2
MUPN = loadmol3 tripos24-mol3.mol2

DIM = loadmol3 dimeric_charged.mol2

charge APN
charge CPN
charge GPN
charge IPN
charge TPN
charge UPN

charge NAP
charge NCP
charge NGP
charge NIP
charge NTP
charge NUP

charge CAP
charge CCP
charge CGP
charge CIP
charge CTP
charge CUP

charge MAPN
charge MCPN
charge MGPN
charge MIPN
charge MTPN
charge MUPN

charge DIM

DUPLICATE = loadpdb DUPLICATE.pdb
check DUPLICATE
solvatebox DUPLICATE TIP3PBOX 12.0
saveAmberParm DUPLICATE DUPLICATE.top DUPLICATE.crd
savepdb DUPLICATE DUPLICATE.pdb
quit
```

8.3.13.generate.sh

```
#!/bin/bash

rm nuc.c a.out nab_tmp.pdb tprmtop tleap.out nab_tmp.pdb* leap.log pna.pdb tmp
duplex.pdb

nab nuc.nab
./a.out

nn=`cat nab_tmp.pdb.lpdb | wc -l`

echo $nn

kt=`echo "${nn} - 2" | bc -l`

echo $kt

head -${kt} nab_tmp.pdb.lpdb > pna.pdb

grep 'TER' -B 1000000 pna.pdb > tmp

mv tmp pna.pdb

rm nuc.c a.out nab_tmp.pdb tprmtop tleap.out nab_tmp.pdb* leap.log

nab nuc2.nab
./a.out

grep 'TER' -A 1000000 nuc.pdb > tmp

cat pna.pdb tmp > duplex.pdb

rm nuc.c a.out nab_tmp.pdb tprmtop tleap.out nab_tmp.pdb* leap.log pna.pdb tmp

tleap -f script.sh

sander -O -i min.in -o duplex.out -c DUPLEX.crd -p DUPLEX.top -r duplex.rst

ambpdb -aatm -p DUPLEX.top < duplex.rst > duplex.pdb

grep -v "WAT" duplex.pdb > tmp

mv tmp duplex.pdb

tleap -f script2.sh

rm leap.log 0PNA-from-mol3-files.log duplex.* mdinfo nuc.pdb
```

8.3.14.correction.sh

```
#!/bin/bash
echo "Which file is to minimize?"
read 'file'

sed "s/duplex.pdb/${file}/g" script.sh > script3.sh
tleap -f script3.sh

sander -O -i min.in -o duplex.out -c DUPLEX.crd -p DUPLEX.top -r duplex.rst
ambpdb -aatm -p DUPLEX.top < duplex.rst > duplex.pdb

grep -v "WAT" duplex.pdb > tmp
mv tmp duplex.pdb

tleap -f script2.sh

rm leap.log OPNA-from-mol3-files.log duplex.* mdinfo nuc.pdb
```

8.4. HPLC purification gradients

Eluents: water +0.1% TFA (A) and acetonitrile +0.1% TFA (B).

8.4.1. Unmodified PNA purification gradient

Time (min)	% A	% B
0	100	0
5	100	0
20	75	25
21	0	100
26	0	100
27	100	0
36	100	0

8.4.2. PNA1 purification gradients

Cycle 1:

Time (min)	% A	% B
0	100	0
5	100	0
50	62.5	37.5
51	0	100
56	0	100
57	100	0
66	100	0

Cycle 2:

Time (min)	% A	% B
0	100	0
5	100	0
41	70	30
42	0	100
47	0	100
48	100	0
57	100	0

8.4.3. PNA2 purification gradients

Cycle 1:

Time (min)	% A	% B
0	100	0
5	100	0
30	70	30
31	0	100
36	0	100
37	100	0
46	100	0

Cycle 2:

Time (min)	% A	% B
0	100	0
5	100	0
41	70	30
42	0	100
47	0	100
48	100	0
57	100	0

8.4.4. PNA3 purification gradients

Cycle 1:

Time (min)	% A	% B
0	100	0
5	100	0
41	70	30
42	0	100
47	0	100
48	100	0
57	100	0

Cycle 2:

Time (min)	% A	% B
0	100	0
5	100	0
31	78.3	21.7
32	0	100
37	0	100
38	100	0
47	100	0

8.5. UPLC gradient

Eluents: water +0.2% HCOOH (A) and acetonitrile +0.2% HCOOH (B)

Gradient:

Time (min)	% A	% B
0.0	100	0
0.9	100	0
6.6	50	50
7.0	50	50
7.2	0	100
8.0	0	100
8.2	100	0
11.0	100	0

8.6. Modified bases

8.6.1. Model 1

DIM

@<TRIPOS>MOLECULE

DIM

73 76 1 0 0

SMALL

resp

@<TRIPOS>ATOM

1	C4"	4.2100	2.9360	-1.1200	C	1	DIM	0.8982200137
2	C5"	4.8630	2.9980	0.1810	CM	1	DIM	-0.6087609863
3	C6"	4.6990	4.1180	0.9390	CM	1	DIM	0.0174510137
4	C2"	3.2880	5.2280	-0.6700	C	1	DIM	0.8899850137
5	H3"	2.9960	4.0820	-2.3270	H	1	DIM	0.4506100137
6	H6"	5.1870	4.1640	1.9070	H4	1	DIM	0.2148800137
7	C7"	5.7170	1.9110	0.7640	c	1	DIM	0.7989920137
8	N2L	5.8770	0.8110	-0.0170	n	1	DIM	-0.4741149863
9	C0L	6.6750	-0.3230	0.4040	c3	1	DIM	-0.2942059863
10	H0L1	7.5740	-0.4000	-0.2240	h1	1	DIM	0.2033580137
11	H0L2	7.0180	-0.0970	1.4190	h1	1	DIM	0.2033580137
12	C7L	5.9410	-1.6530	0.3770	ca	1	DIM	0.2966100137

13	C6L	4.5680	-1.7480	0.3090	ca	1	DIM	-0.5013839863
14	C8L	6.7130	-2.8450	0.4500	ca	1	DIM	-0.4221169863
15	C5L	3.9110	-3.0090	0.3140	ca	1	DIM	0.1981480137
16	H6L	3.9660	-0.8460	0.2470	ha	1	DIM	0.2768690137
17	C9L	6.1110	-4.0800	0.4640	ca	1	DIM	-0.3055369863
18	H8L	7.7970	-2.7710	0.4960	ha	1	DIM	0.2752420137
19	C4L	2.4970	-3.1200	0.2410	ca	1	DIM	-0.5118489863
20	C10L	4.6980	-4.2010	0.3970	ca	1	DIM	0.0629710137
21	H9L	6.7140	-4.9830	0.5200	ha	1	DIM	0.2741200137
22	C3L	1.8710	-4.3490	0.2480	ca	1	DIM	0.3130740137
23	H4L	1.9040	-2.2130	0.1750	ha	1	DIM	0.2839450137
24	C11L	4.0330	-5.4550	0.4020	ca	1	DIM	-0.2959469863
25	C12L	2.6620	-5.5270	0.3270	ca	1	DIM	-0.4382819863
26	H11L	4.6270	-6.3640	0.4590	ha	1	DIM	0.2723910137
27	H12L	2.1700	-6.4960	0.3240	ha	1	DIM	0.2794430137
28	C2L	0.3600	-4.4950	0.2080	c3	1	DIM	-0.2976539863
29	H2L1	0.1040	-5.4160	-0.3340	h1	1	DIM	0.1975590137
30	H2L2	-0.0440	-4.6020	1.2200	h1	1	DIM	0.1975590137
31	N1L	-0.3290	-3.3660	-0.3870	n	1	DIM	-0.4289249863
32	C7	-1.3410	-2.7200	0.2460	c	1	DIM	0.7467350137
33	C5	-1.9570	-1.5810	-0.5150	CM	1	DIM	-0.5279789863
34	C4	-1.5400	-1.1440	-1.8410	C	1	DIM	0.8602970137
35	C6	-2.9890	-0.9330	0.0870	CM	1	DIM	0.0158340137
36	H6	-3.3140	-1.2710	1.0650	H4	1	DIM	0.2182980137
37	C2	-3.3290	0.6130	-1.7360	C	1	DIM	0.9211980137
38	H3	-2.0240	0.2710	-3.2570	H	1	DIM	0.4410200137
39	C8'	-4.8160	0.7010	0.1910	CT	1	DIM	-0.8799629863
40	H8'1	-4.6550	0.7180	1.2720	H1	1	DIM	0.3446040137
41	H8'2	-4.9220	1.7270	-0.1660	H1	1	DIM	0.3446040137
42	C7'	-6.0700	-0.1210	-0.1800	C	1	DIM	1.0245680137
43	N4'	-7.2500	0.2550	0.3950	N	1	DIM	-0.3187549863
44	C3'	-7.4190	1.3990	1.2990	CT	1	DIM	-0.3469869863
45	H3'1	-8.3710	1.8710	1.0460	H1	1	DIM	0.2204430137
46	H3'2	-6.6410	2.1430	1.1170	H1	1	DIM	0.2204430137
47	C5'	-8.4580	-0.5010	0.0360	CT	1	DIM	-0.4152869863
48	H5'1	-8.1450	-1.4790	-0.3220	H1	1	DIM	0.2243340137
49	H5'2	-9.0720	-0.6290	0.9320	H1	1	DIM	0.2243340137
50	C'	-9.3190	0.2850	-0.9660	C	1	DIM	0.7478980137
51	C2'	-7.4100	0.9880	2.7870	CT	1	DIM	0.1274780137
52	H2'1	-6.4470	0.5470	3.0550	H1	1	DIM	0.1389980137
53	H2'2	-8.1620	0.2080	2.9550	H1	1	DIM	0.1389980137
54	N1'	-7.6860	2.1070	3.6670	N	1	DIM	-0.7980199863
55	H1'1	-8.6380	2.4490	3.7150	H	1	DIM	0.4787090137
56	CC1	3.8110	6.3810	1.4140	CT	1	DIM	-0.9552239863
57	HC1	4.1850	7.2640	0.8930	H1	1	DIM	0.3107190137
58	HC2	2.7580	6.5430	1.6540	H1	1	DIM	0.3107190137
59	HC3	4.3800	6.2210	2.3290	H1	1	DIM	0.3107190137
60	O2"	2.6180	6.1790	-1.0270	O	1	DIM	-0.7900779863
61	O4"	4.2530	2.0090	-1.9280	O	1	DIM	-0.7882199863
62	N3"	3.4660	4.0830	-1.4290	NA	1	DIM	-0.6127129863
63	O7"	6.2220	2.0480	1.8800	o	1	DIM	-0.7705439863
64	H2L	5.4270	0.8200	-0.9290	hn	1	DIM	0.4077910137
65	H1L	-0.0850	-3.0450	-1.3210	hn	1	DIM	0.3892720137
66	O7	-1.7560	-3.0130	1.3700	o	1	DIM	-0.7597729863
67	O4	-0.6450	-1.6250	-2.5330	O	1	DIM	-0.7739679863
68	N3	-2.2730	-0.0520	-2.3290	NA	1	DIM	-0.5804379863
69	O2	-3.9250	1.5360	-2.2610	O	1	DIM	-0.7950389863
70	N1	-3.6370	0.1410	-0.4490	N*	1	DIM	0.1742120137
71	O7'	-5.9690	-1.0670	-0.9560	O	1	DIM	-0.7750459863
72	O1'	-9.9270	1.2860	-0.5870	O	1	DIM	-0.7948819863
73	N1"	3.9560	5.1960	0.5630	N*	1	DIM	0.3146820137

@<TRIPOS>BOND

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60	43	44	1
61	43	47	1

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62  44  45  1
63  44  46  1
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70  51  53  1
71  51  54  1
72  54  55  1
73  56  57  1
74  56  58  1
75  56  59  1
76  56  73  1
@<TRIPOS>SUBSTRUCTURE
  1 DIM          1 TEMP          0 ***** 0 ROOT
@<TRIPOS>HEADTAIL
N1' 1
C' 1
@<TRIPOS>RESIDUECONNECT
1 N1' C' 0 0 0 0

```

frcmod.pna

PNA frcmod file: frcmod.pna
 MASS

BOND

CQ-NA	502.0	1.324	by analogy to CQ-NC, parm99.dat
CM-c	349.7	1.487	SOURCE1 from gaff.dat

ANGL

C -CT-N*	50.00	110.13	FF cst by analogy to CT-CT-N*; Eq.val. from
QM geo.opt.			
CQ-NA-C	70.0	118.60	by analogy to CA-NC-CQ, parm99.dat
H5-CQ-NA	50.0	115.45	by analogy to H5-CQ-NC, parm99.dat
NC-CQ-NA	70.0	129.10	by analogy to NC-CQ-NC, parm99.dat
CQ-NA-H	50.0	118.00	by analogy to CA-NA-H, parm99.dat
CM-c -o	68.670	123.44	SOURCE3 from gaff.dat
c -CM-C	62.650	120.000	SOURCE3 from gaff.dat
c -CM-CM	64.640	120.140	SOURCE3 from gaff.dat
n -c -CM	68.470	115.140	SOURCE4 from gaff.dat

DIHEDRAL

X -CQ-NA-X	4	6.00	180.0	2.0	by analogy to X -CA-NA-X,
parm99.dat					
CM-CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
CM-CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
C -CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
C -CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
n -c -CM-C	4	4.000	180.0	2.	optimized by Junmei Wang,
Jan-2013					
CM-CM-c -n	4	4.000	180.0	2.	optimized by Junmei Wang,
Jan-2013					

NONB

8.6.2. Model1 with cutter

NBC

@<TRIPOS>MOLECULE

NBC

106 109 1 0 0

SMALL

NO_CHARGES

Energy = 0

@<TRIPOS>ATOM

1	C4"	4.1950	3.3380	-0.9090	C	1	NBC	0.896168
2	C5"	4.8370	3.3700	0.3990	CM	1	NBC	-0.610813
3	C6"	4.6230	4.4490	1.2030	CM	1	NBC	0.015399
4	C2"	3.1780	5.5690	-0.3690	C	1	NBC	0.887933
5	H3"	2.9430	4.4850	-2.0750	H	1	NBC	0.448558
6	H6"	5.1020	4.4730	2.1760	H4	1	NBC	0.212828
7	C7"	5.7310	2.2950	0.9420	c	1	NBC	0.796940
8	N2L	5.9400	1.2370	0.1160	n	1	NBC	-0.476167
9	C0L	6.7810	0.1190	0.4950	c3	1	NBC	-0.296258
10	H0L1	7.6870	0.1050	-0.1280	h1	1	NBC	0.201306
11	H0L2	7.1080	0.3150	1.5210	h1	1	NBC	0.201306
12	C7L	6.1020	-1.2370	0.4050	ca	1	NBC	0.294558
13	C6L	4.7350	-1.3850	0.3220	ca	1	NBC	-0.503436
14	C8L	6.9220	-2.3990	0.4340	ca	1	NBC	-0.424169
15	C5L	4.1300	-2.6700	0.2680	ca	1	NBC	0.196096
16	H6L	4.0970	-0.5060	0.2940	ha	1	NBC	0.274817
17	C9L	6.3700	-3.6570	0.3900	ca	1	NBC	-0.307589
18	H8L	8.0010	-2.2830	0.4920	ha	1	NBC	0.273190
19	C4L	2.7220	-2.8350	0.1790	ca	1	NBC	-0.513901
20	C10L	4.9640	-3.8320	0.3070	ca	1	NBC	0.060919
21	H9L	7.0090	-4.5360	0.4130	ha	1	NBC	0.272068
22	C3L	2.1460	-4.0880	0.1290	ca	1	NBC	0.311022
23	H4L	2.0930	-1.9510	0.1470	ha	1	NBC	0.281893
24	C11L	4.3510	-5.1110	0.2530	ca	1	NBC	-0.297999
25	C12L	2.9840	-5.2350	0.1640	ca	1	NBC	-0.440334
26	H11L	4.9810	-5.9970	0.2760	ha	1	NBC	0.270339
27	H12L	2.5320	-6.2220	0.1160	ha	1	NBC	0.277391
28	C2L	0.6430	-4.2930	0.0710	c3	1	NBC	-0.299706
29	H2L1	0.4280	-5.1990	-0.5120	h1	1	NBC	0.195507
30	H2L2	0.2370	-4.4590	1.0740	h1	1	NBC	0.195507
31	N1L	-0.0880	-3.1680	-0.4810	n	1	NBC	-0.430977
32	C7	-1.1290	-2.5910	0.1700	c	1	NBC	0.744683
33	C5	-1.7860	-1.4470	-0.5460	CM	1	NBC	-0.530031
34	C4	-1.3790	-0.9370	-1.8490	C	1	NBC	0.858245
35	C6	-2.8480	-0.8670	0.0740	CM	1	NBC	0.013782
36	H6	-3.1650	-1.2600	1.0350	H4	1	NBC	0.216246
37	C2	-3.2390	0.7400	-1.6840	C	1	NBC	0.919146
38	H3	-1.9110	0.5170	-3.2070	H	1	NBC	0.438968
39	C8'	-4.7400	0.6850	0.2330	CT	1	NBC	-0.882015
40	H8'1	-4.5870	0.6630	1.3150	H1	1	NBC	0.342552
41	H8'2	-4.8860	1.7210	-0.0810	H1	1	NBC	0.342552
42	C7'	-5.9580	-0.1700	-0.1820	C	1	NBC	1.022516
43	N4'	-7.1550	0.1330	0.3980	N	1	NBC	-0.320807
44	C3'	-7.3770	1.2290	1.3490	CT	1	NBC	-0.349039
45	H3'1	-8.3450	1.6730	1.1090	H1	1	NBC	0.218391

46	H3'2	-6.6280	2.0110	1.2050	H1	1	NBC	0.218391
47	C5'	-8.3290	-0.6550	-0.0020	CT	1	NBC	-0.417339
48	H5'1	-7.9750	-1.6030	-0.3990	H1	1	NBC	0.222282
49	H5'2	-8.9430	-0.8460	0.8830	H1	1	NBC	0.222282
50	C'	-9.2150	0.1380	-0.9770	C	1	NBC	0.745846
51	C2'	-7.3600	0.7560	2.8180	CT	1	NBC	0.125426
52	H2'1	-6.3820	0.3430	3.0750	H1	1	NBC	0.136946
53	H2'2	-8.0810	-0.0610	2.9470	H1	1	NBC	0.136946
54	N1'	-7.6870	1.8240	3.7430	N	1	NBC	-0.800072
55	H1'1	-8.6530	2.1250	3.7980	H	1	NBC	0.476657
56	O2"	2.4720	6.5070	-0.6900	O	1	NBC	-0.792130
57	O4"	4.2810	2.4490	-1.7550	O	1	NBC	-0.790272
58	N3"	3.4070	4.4660	-1.1740	NA	1	NBC	-0.614765
59	O7"	6.2230	2.4040	2.0670	o	1	NBC	-0.772596
60	H2L	5.4960	1.2670	-0.7980	hn	1	NBC	0.405739
61	H1L	0.1490	-2.7980	-1.3990	hn	1	NBC	0.387220
62	O7	-1.5390	-2.9490	1.2780	o	1	NBC	-0.761825
63	O4	-0.4610	-1.3510	-2.5540	O	1	NBC	-0.776020
64	N3	-2.1530	0.1450	-2.2960	NA	1	NBC	-0.582490
65	O2	-3.8690	1.6600	-2.1740	O	1	NBC	-0.797091
66	N1	-3.5360	0.2020	-0.4200	N*	1	NBC	0.172160
67	O7'	-5.8130	-1.0770	-0.9970	O	1	NBC	-0.777098
68	O1'	-9.8660	1.0960	-0.5600	O	1	NBC	-0.796934
69	N1"	3.8390	5.5110	0.8670	N*	1	NBC	0.312630
70	C1A	3.6823	6.6196	1.8196	c3	1	NBC	0.020637
71	H1A1	4.1015	6.3386	2.7631	h1	1	NBC	0.110634
72	C2A	4.4039	7.8707	1.2934	c3	1	NBC	0.020637
73	H2A1	4.3974	8.6580	2.0495	h1	1	NBC	0.110634
74	H2A2	3.8900	8.2542	0.4007	h1	1	NBC	0.110634
75	N3A	5.7982	7.6056	0.9866	n	1	NBC	-0.721821
76	H3A1	5.8806	7.3169	0.0327	hn	1	NBC	0.478007
77	H1A2	2.6418	6.8361	1.9439	h1	1	NBC	0.110634
78	CCC	6.5876	8.8274	1.1987	c	1	NBC	0.920517
79	C2C	8.0609	8.7262	1.6474	c3	1	NBC	-0.985691
80	OCC	6.0621	9.9559	1.0144	o	1	NBC	-0.906523
81	N0C	8.8047	9.9709	1.3552	n3	1	NBC	0.485873
82	H2C1	8.0487	8.5664	2.7312	h1	1	NBC	0.348005
83	H2C2	8.5423	7.8482	1.2003	h1	1	NBC	0.348005
84	C1C	9.4481	9.9692	0.0048	c3	1	NBC	-0.690862
85	C3C	9.6909	10.4511	2.4600	c3	1	NBC	-0.690862
86	C9C	9.7751	11.3888	-0.5012	c3	1	NBC	-0.271161
87	H1C1	10.3666	9.3684	-0.0006	h1	1	NBC	0.295748
88	H1C2	8.7444	9.4912	-0.6855	h1	1	NBC	0.295748
89	C4C	9.0104	11.5214	3.3376	c3	1	NBC	-0.271161
90	H3C1	10.6040	10.8506	2.0155	h1	1	NBC	0.295748
91	H3C2	10.0096	9.6228	3.1008	h1	1	NBC	0.295748
92	N8C	8.6226	12.3497	-0.3482	n3	1	NBC	-0.626037
93	H9C1	10.6381	11.7995	0.0271	h1	1	NBC	0.238064
94	H9C2	10.0630	11.3208	-1.5537	h1	1	NBC	0.238064
95	N5C	8.3544	12.6035	2.5224	n3	1	NBC	-0.626037
96	H4C1	9.7540	11.9481	4.0198	h1	1	NBC	0.238064
97	H4C2	8.2241	11.0722	3.9533	h1	1	NBC	0.238064
98	C7C	8.9790	13.6578	0.2972	c3	1	NBC	-0.360158
99	H8C1	8.2380	12.5529	-1.2697	hn	1	NBC	0.468702
100	C6C	9.3200	13.4992	1.7910	c3	1	NBC	-0.360158
101	H5C1	7.7964	13.1702	3.1595	hn	1	NBC	0.468702
102	H7C1	8.1136	14.3166	0.1719	h1	1	NBC	0.250612
103	H7C2	9.8262	14.1296	-0.2125	h1	1	NBC	0.250612
104	H6C1	9.3279	14.4910	2.2503	h1	1	NBC	0.250612
105	H6C2	10.3270	13.0951	1.9154	h1	1	NBC	0.250612
106	Zn	7.3006	11.4904	1.0595	Zn	1	NBC	1.392676

@<TRIPOS>BOND

1	1	57	2
2	1	58	1
3	1	2	1
4	2	3	2
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6	3	6	1
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8	4	56	2
9	4	58	1
10	4	69	1
11	5	58	1
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32	22	25	1
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34	24	25	2
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41	31	61	1
42	32	33	1
43	32	62	2
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45	33	35	2
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51	37	65	2
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53	38	64	1
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55	39	40	1
56	39	41	1
57	39	42	1
58	42	43	1
59	42	67	2
60	43	44	1
61	43	47	1
62	44	45	1

63	44	51	1
64	44	46	1
65	47	48	1
66	47	50	1
67	47	49	1
68	50	68	2
69	51	53	1
70	51	54	1
71	51	52	1
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102	92	99	1
103	95	100	1
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105	98	100	1
106	98	102	1
107	98	103	1
108	100	104	1
109	100	105	1

@<TRIPOS>SUBSTRUCTURE

1 NBC 1 TEMP

0 **** **** 0 ROOT

@<TRIPOS>HEADTAIL

N1' 1

C' 1

@<TRIPOS>RESIDUECONNECT

1 N1' C' 0 0 0 0

frcmod.pna

PNA frcmod file: frcmod.pna
 MASS

BOND

CQ-NA	502.0	1.324	by analogy to CQ-NC, parm99.dat
CM-c	349.7	1.487	SOURCE1 from gaff.dat
n3-Zn	50.4	2.089	ZAFF
c3-N*	337.0	1.475	JCC,7,(1986),230; ADE,CYT,GUA,THY,URA

ANGL

C -CT-N*	50.00	110.13	FF cst by analogy to CT-CT-N*; Eq.val. from QM geo.opt.
CQ-NA-C	70.0	118.60	by analogy to CA-NC-CQ, parm99.dat
H5-CQ-NA	50.0	115.45	by analogy to H5-CQ-NC, parm99.dat
NC-CQ-NA	70.0	129.10	by analogy to NC-CQ-NC, parm99.dat
CQ-NA-H	50.0	118.00	by analogy to CA-NA-H, parm99.dat
CM-c -o	68.670	123.44	SOURCE3 from gaff.dat
c -CM-C	62.650	120.000	SOURCE3 from gaff.dat
c -CM-CM	64.640	120.140	SOURCE3 from gaff.dat
n -c -CM	68.470	115.140	SOURCE4 from gaff.dat
N*-c3-h1	50.0	109.50	parm99
N*-c3-c3	67.530	112.140	SOURCE4
C-N*-c3	70.0	117.60	parm99
CM-N*-c3	70.0	121.20	parm99

DIHEDRAL

X -CQ-NA-X	4	6.00	180.0	2.0	by analogy to X -CA-NA-X, parm99.dat
CM-CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
CM-CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
C -CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
C -CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
n -c -CM-C	4	4.000	180.0	2.	optimized by Junmei Wang, Jan-2013
CM-CM-c -n	4	4.000	180.0	2.	optimized by Junmei Wang, Jan-2013
X-c3-N*-X	6	0.00	0.0	2.	JCC,7,(1986),230

NONB

8.6.3. Model2

L1D

@<TRIPOS>MOLECULE

L1D

74 77 1 0 0

SMALL

resp

@<TRIPOS>ATOM

1	C4"	5.5680	3.0780	-1.0530	C	1	L1D	0.875570
2	C5"	5.7560	2.9470	0.3900	CM	1	L1D	-0.352481
3	C6"	5.2680	3.9210	1.1940	CM	1	L1D	-0.134303
4	C2"	4.3940	5.2530	-0.6280	C	1	L1D	0.931158
5	H3"	4.7430	4.3730	-2.4310	H	1	L1D	0.463365
6	H6"	5.3740	3.8670	2.2730	H4	1	L1D	0.284295
7	C7"	6.4960	1.7410	0.9060	c3	1	L1D	-0.511818
8	N2L	5.7850	0.4990	0.5960	n3	1	L1D	-0.512400
9	C0L	6.5040	-0.6840	1.0560	c3	1	L1D	-0.328426
10	H0L1	7.5260	-0.7590	0.6360	h1	1	L1D	0.212106
11	H0L2	6.6410	-0.5870	2.1450	h1	1	L1D	0.212106
12	C7L	5.7590	-1.9720	0.7660	ca	1	L1D	0.208201
13	C6L	4.3890	-1.9980	0.6020	ca	1	L1D	-0.410828
14	C8L	6.4860	-3.1910	0.6890	ca	1	L1D	-0.389291
15	C5L	3.6960	-3.2140	0.3580	ca	1	L1D	0.203371
16	H6L	3.8320	-1.0680	0.6580	ha	1	L1D	0.193653
17	C9L	5.8460	-4.3860	0.4630	ca	1	L1D	-0.308743
18	H8L	7.5670	-3.1690	0.8080	ha	1	L1D	0.263920
19	C4L	2.2860	-3.2530	0.1840	ca	1	L1D	-0.532370
20	C10L	4.4380	-4.4360	0.2900	ca	1	L1D	0.044448
21	H9L	6.4150	-5.3110	0.4040	ha	1	L1D	0.272120
22	C3L	1.6230	-4.4380	-0.0530	ca	1	L1D	0.302742
23	H4L	1.7280	-2.3230	0.2360	ha	1	L1D	0.303871
24	C11L	3.7350	-5.6450	0.0450	ca	1	L1D	-0.291310
25	C12L	2.3710	-5.6450	-0.1260	ca	1	L1D	-0.429242
26	H11L	4.2950	-6.5750	-0.0150	ha	1	L1D	0.268923
27	H12L	1.8500	-6.5800	-0.3210	ha	1	L1D	0.272918
28	C2L	0.1150	-4.5140	-0.2110	c3	1	L1D	-0.297705
29	H2L1	-0.1300	-5.3010	-0.9380	h1	1	L1D	0.197035
30	H2L2	-0.3590	-4.8010	0.7330	h1	1	L1D	0.197035
31	N1L	-0.5020	-3.2640	-0.6130	n	1	L1D	-0.440216
32	C7	-1.5380	-2.7150	0.0690	c	1	L1D	0.758291
33	C5	-2.0580	-1.4180	-0.4830	CM	1	L1D	-0.570754
34	C4	-1.5170	-0.7330	-1.6510	C	1	L1D	0.887496
35	C6	-3.1180	-0.8630	0.1610	CM	1	L1D	0.040649
36	H6	-3.5350	-1.3800	1.0190	H4	1	L1D	0.216458
37	C2	-3.2600	1.0410	-1.3200	C	1	L1D	0.921597
38	H3	-1.8380	0.9650	-2.7710	H	1	L1D	0.439934
39	C8'	-4.8990	0.8010	0.4620	CT	1	L1D	-0.847203
40	H8'1	-4.8420	0.5680	1.5290	H1	1	L1D	0.322992
41	H8'2	-4.9280	1.8860	0.3390	H1	1	L1D	0.322992
42	C7'	-6.1440	0.1600	-0.1830	C	1	L1D	0.939704
43	N4'	-7.3590	0.4730	0.3500	N	1	L1D	-0.062888
44	C3'	-7.6090	1.5200	1.3450	CT	1	L1D	-0.437516
45	H3'1	-8.4790	2.0930	1.0080	H1	1	L1D	0.233708
46	H3'2	-6.7710	2.2200	1.3780	H1	1	L1D	0.233708

47	C5'	-8.5430	-0.1400	-0.2750	CT	1	L1D	-0.870227
48	H5'1	-8.2730	-1.1440	-0.6050	H1	1	L1D	0.324431
49	H5'2	-9.3410	-0.1990	0.4650	H1	1	L1D	0.324431
50	C'	-9.0550	0.7300	-1.4350	C	1	L1D	0.825499
51	C2'	-7.8670	0.9520	2.7560	CT	1	L1D	0.168043
52	H2'1	-6.9860	0.4110	3.1080	H1	1	L1D	0.125461
53	H2'2	-8.6810	0.2190	2.7110	H1	1	L1D	0.125461
54	N1'	-8.2130	1.9860	3.7120	N	1	L1D	-0.824348
55	H1'1	-9.1470	2.3740	3.6760	H	1	L1D	0.481077
56	O2"	3.8130	6.2360	-1.0560	O	1	L1D	-0.814395
57	O4"	5.9400	2.2690	-1.8950	O	1	L1D	-0.773254
58	N3"	4.8960	4.2470	-1.4370	NA	1	L1D	-0.683505
59	H2L	5.6820	0.4510	-0.4170	hn	1	L1D	0.400919
60	H1L	-0.1730	-2.7680	-1.4380	hn	1	L1D	0.391910
61	O7	-2.0480	-3.2100	1.0760	o	1	L1D	-0.765352
62	O4	-0.5780	-1.0980	-2.3550	O	1	L1D	-0.784547
63	N3	-2.1760	0.4680	-1.9550	NA	1	L1D	-0.595513
64	O2	-3.7880	2.0750	-1.6860	O	1	L1D	-0.791260
65	N1	-3.6880	0.3280	-0.1880	N*	1	L1D	0.178296
66	O7'	-6.0150	-0.6140	-1.1310	O	1	L1D	-0.771219
67	O1'	-9.9490	1.5530	-1.2560	O	1	L1D	-0.748132
68	N1"	4.6110	5.0420	0.7340	N*	1	L1D	0.257124
69	CC1	4.1000	6.0710	1.6410	CT	1	L1D	-0.899509
70	HC1	4.5560	7.0370	1.4120	H1	1	L1D	0.293632
71	HC2	3.0180	6.1700	1.5310	H1	1	L1D	0.293632
72	HC3	4.3410	5.7850	2.6650	H1	1	L1D	0.293632
73	H7"1	7.5220	1.7510	0.4890	h1	1	L1D	0.268316
74	H7"2	6.6010	1.8190	1.9950	h1	1	L1D	0.268316

@<TRIPOS>BOND

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28	17	20	1
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30	19	22	1
31	19	23	1
32	20	24	1
33	22	25	1

```

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75  69  70  1
76  69  71  1
77  69  72  1

```

@<TRIPOS>SUBSTRUCTURE

```

1 L1D          1 TEMP          0 ***** 0 ROOT

```

@<TRIPOS>HEADTAIL

N1' 1

C' 1

@<TRIPOS>RESIDUECONNECT

```

1 N1' C' 0 0 0 0

```

frcmod.pna

PNA frcmod file: frcmod.pna
 MASS

BOND

```

CQ-NA          502.0    1.324
CM-c           349.7    1.487

```

by analogy to CQ-NC, parm99.dat
 SOURCE1 from gaff.dat

n3-Zn	50.4	2.089	ZAFF		
c3-N*	337.0	1.475	JCC, 7, (1986), 230;	ADE, CYT, GUA, THY, URA	
CM-c3	317.0	1.510	by analogy	parm99	
ANGL					
C -CT-N*	50.00	110.13	FF cst by analogy to	CT-CT-N*;	Eq.val. from
QM geo.opt.					
CQ-NA-C	70.0	118.60	by analogy to	CA-NC-CQ,	parm99.dat
H5-CQ-NA	50.0	115.45	by analogy to	H5-CQ-NC,	parm99.dat
NC-CQ-NA	70.0	129.10	by analogy to	NC-CQ-NC,	parm99.dat
CQ-NA-H	50.0	118.00	by analogy to	CA-NA-H,	parm99.dat
CM-c -o	68.670	123.44	SOURCE3	from	gaff.dat
c -CM-C	62.650	120.000	SOURCE3	from	gaff.dat
c -CM-CM	64.640	120.140	SOURCE3	from	gaff.dat
n -c -CM	68.470	115.140	SOURCE4	from	gaff.dat
N*-c3-h1	50.0	109.50	parm99		
N*-c3-c3	67.530	112.140	SOURCE4		
C-N*-c3	70.0	117.60	parm99		
CM-N*-c3	70.0	121.20	parm99		
CM-CM-c3	70.0	119.70	parm99		
CM-c3-n3	66.18	112.13	SOURCE4	387	1.2309
CM-c3-h1	46.78	110.95	SOURCE3	12	1.1170
C -CM-c3	70.0	119.70	by	parm99	
DIHEDRAL					
X -CQ-NA-X	4	6.00	180.0	2.0	by analogy to X -CA-NA-X,
parm99.dat					
CM-CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
CM-CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
C -CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
C -CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
n -c -CM-C	4	4.000	180.0	2.	optimized by Junmei Wang,
Jan-2013					
CM-CM-c -n	4	4.000	180.0	2.	optimized by Junmei Wang,
Jan-2013					
X -c3-N*-X	6	0.00	0.0	2.	JCC, 7, (1986), 230
X -c3-CM-X	6	0.000	0.000	2.	JCC, 7, (1986), 230

NONB

8.6.4. Model3

DCL

@<TRIPOS>MOLECULE

DCL

71 76 1 0 0

SMALL

resp

@<TRIPOS>ATOM

1	C5	4.1580	0.4530	-1.0520	CM	1	DCL	-0.678993
2	N1	6.4860	-0.1270	-1.1960	N*	1	DCL	0.353895
3	C2	6.2210	-1.4180	-1.6720	C	1	DCL	0.884243
4	C6	5.4740	0.7650	-0.9340	CM	1	DCL	-0.045393
5	N3	4.8670	-1.7160	-1.7360	NA	1	DCL	-0.692795
6	O2	7.0980	-2.1970	-1.9860	O	1	DCL	-0.767500
7	C4	3.7640	-0.8910	-1.4790	C	1	DCL	0.995462

8	O4	2.6250	-1.3120	-1.6130	O	1	DCL	-0.790335
9	H3	4.6480	-2.6530	-2.0580	H	1	DCL	0.458389
10	C5"	-8.0960	-1.3320	0.0790	CM	1	DCL	-0.638314
11	N1"	-10.1930	-2.5010	0.0790	N*	1	DCL	0.234916
12	C2"	-9.6020	-3.7370	-0.1940	C	1	DCL	0.896040
13	C6"	-9.4500	-1.3570	0.2050	CM	1	DCL	0.035812
14	N3"	-8.2210	-3.6750	-0.3190	NA	1	DCL	-0.727057
15	O2"	-10.2440	-4.7640	-0.3110	O	1	DCL	-0.780540
16	C4"	-7.3700	-2.5670	-0.2090	C	1	DCL	1.009499
17	O4"	-6.1620	-2.6970	-0.3520	O	1	DCL	-0.796925
18	H3"	-7.7660	-4.5580	-0.5190	H	1	DCL	0.468883
19	C12L	-0.1570	4.4830	0.1420	ca	1	DCL	-0.172488
20	C3L	0.0310	3.0950	-0.1210	ca	1	DCL	-0.092917
21	C4L	-1.0700	2.2570	-0.1290	ca	1	DCL	-0.422894
22	C11L	-1.4120	4.9860	0.3870	ca	1	DCL	-0.440297
23	C10L	-2.5550	4.1440	0.3850	ca	1	DCL	0.191989
24	C5L	-2.3770	2.7480	0.1200	ca	1	DCL	0.193024
25	C6L	-3.5130	1.8990	0.1140	ca	1	DCL	-0.428723
26	C7L	-4.7840	2.3890	0.3580	ca	1	DCL	-0.092585
27	C8L	-4.9500	3.7800	0.6230	ca	1	DCL	-0.172110
28	C9L	-3.8680	4.6250	0.6340	ca	1	DCL	-0.438412
29	CC1	-11.6530	-2.4920	0.2200	CT	1	DCL	-0.966796
30	C8'	7.8960	0.2800	-1.1490	CT	1	DCL	-0.962252
31	C13L	-5.9620	1.5150	0.3500	c2	1	DCL	0.396335
32	C2L	1.3850	2.5900	-0.3750	c2	1	DCL	0.398493
33	C0L	-6.0660	0.1570	0.1020	c2	1	DCL	-0.771309
34	NA1"	-7.3940	-0.1140	0.2280	na	1	DCL	1.002196
35	NA2"	-8.0840	1.0240	0.5400	n2	1	DCL	-0.257207
36	NA3"	-7.2220	1.9910	0.6100	n2	1	DCL	-0.429186
37	NA3	2.4790	3.4180	-0.3820	n2	1	DCL	-0.428986
38	NA2	3.5510	2.7350	-0.6330	n2	1	DCL	-0.261580
39	NA1	3.1730	1.4310	-0.7940	na	1	DCL	1.058193
40	C1L	1.8260	1.3060	-0.6400	c2	1	DCL	-0.797263
41	H6	5.7700	1.7620	-0.6310	H4	1	DCL	0.199919
42	H6"	-9.9940	-0.4460	0.4180	H4	1	DCL	0.179932
43	H12L	0.7120	5.1320	0.1460	ha	1	DCL	0.187421
44	H4L	-0.9500	1.1940	-0.3290	ha	1	DCL	0.280602
45	H11L	-1.5440	6.0470	0.5870	ha	1	DCL	0.292821
46	H6L	-3.3630	0.8410	-0.0880	ha	1	DCL	0.282571
47	H8L	-5.9500	4.1540	0.8140	ha	1	DCL	0.187303
48	H9L	-4.0070	5.6850	0.8360	ha	1	DCL	0.292054
49	HC1	-11.9750	-1.4720	0.4340	H1	1	DCL	0.316690
50	HC2	-11.9530	-3.1550	1.0340	H1	1	DCL	0.316690
51	HC3	-12.1200	-2.8430	-0.7030	H1	1	DCL	0.316690
52	H8'1	7.9230	1.3260	-0.8300	H1	1	DCL	0.344292
53	H8'2	8.3260	0.2120	-2.1500	H1	1	DCL	0.344292
54	H0L	-5.3590	-0.6180	-0.1400	h4	1	DCL	0.371286
55	H1L1	1.3360	0.3530	-0.7440	h4	1	DCL	0.381215
56	C7'	8.8460	-0.5630	-0.2720	C	1	DCL	0.832004
57	O7'	9.9100	-0.9170	-0.7690	O	1	DCL	-0.734164
58	N4'	8.5320	-0.8440	1.0310	N	1	DCL	0.044965
59	C3'	7.4410	-0.2250	1.8000	CT	1	DCL	-0.585281
60	H3'1	7.8600	0.1380	2.7450	H1	1	DCL	0.277921
61	H3'2	7.0870	0.6570	1.2610	H1	1	DCL	0.277921
62	C2'	6.2470	-1.1430	2.1120	CT	1	DCL	0.009452
63	H2'1	5.7970	-1.4990	1.1790	H1	1	DCL	0.150075
64	H2'2	5.5030	-0.5580	2.6600	H1	1	DCL	0.150075
65	N1'	6.5910	-2.2810	2.9360	N	1	DCL	-0.822702
66	C5'	9.5480	-1.6190	1.7530	CT	1	DCL	-0.652017
67	H5'1	9.3000	-1.5790	2.8160	H1	1	DCL	0.285724
68	H5'2	10.5270	-1.1580	1.6050	H1	1	DCL	0.285724
69	C'	9.5520	-3.1080	1.3610	C	1	DCL	0.751758

70 O'	8.5320	-3.7950	1.4750	O	1 DCL	-0.797801
71 H1'1	7.1310	-3.0250	2.4920	H	1 DCL	0.538828

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59	38	39	1

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62 56 57 1
63 56 58 1
64 58 59 1
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72 65 71 1
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@<TRIPOS>SUBSTRUCTURE

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1 DCL          1 TEMP          0 ***** 0 ROOT

```

@<TRIPOS>HEADTAIL

N1' 1

C' 1

@<TRIPOS>RESIDUECONNECT

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```

frcmod.pna

PNA frcmod file: frcmod.pna

MASS

BOND

CQ-NA	502.0	1.324	by analogy to CQ-NC, parm99.dat
CM-c	349.7	1.487	SOURCE1 from gaff.dat
n3-Zn	50.4	2.089	ZAFF
c3-N*	337.0	1.475	JCC,7, (1986),230; ADE,CYT,GUA,THY,URA
CM-c3	317.0	1.510	by analogy parm99
c2-CM	357.2	1.4800	SOUECE3 1
CM-na	422.0	1.385	JCC,7, (1986),230; HIS

ANGL

C -CT-N*	50.00	110.13	FF cst by analogy to CT-CT-N*; Eq.val. from
QM geo.opt.			
CQ-NA-C	70.0	118.60	by analogy to CA-NC-CQ, parm99.dat
H5-CQ-NA	50.0	115.45	by analogy to H5-CQ-NC, parm99.dat
NC-CQ-NA	70.0	129.10	by analogy to NC-CQ-NC, parm99.dat
CQ-NA-H	50.0	118.00	by analogy to CA-NA-H, parm99.dat
CM-c -o	68.670	123.44	SOURCE3 from gaff.dat
c -CM-C	62.650	120.000	SOURCE3 from gaff.dat
c -CM-CM	64.640	120.140	SOURCE3 from gaff.dat
n -c -CM	68.470	115.140	SOURCE4 from gaff.dat
N*-c3-h1	50.0	109.50	parm99
N*-c3-c3	67.530	112.140	SOURCE4
C-N*-c3	70.0	117.60	parm99
CM-N*-c3	70.0	121.20	parm99
CM-CM-c3	70.0	119.70	parm99
CM-c3-n3	66.18	112.13	SOURCE4 387 1.2309
CM-c3-h1	46.78	110.95	SOURCE3 12 1.1170
C -CM-c3	70.0	119.70	by parm99
c2-c2-CM	66.880	117.000	SOURCE3 1

c2-c -ca	67.170	116.780	SOURCE3	1	
c2-CM-C	67.930	120.700	SOURCE3	1	
c2-CM-CM	64.690	120.600	SOURCE3		
ha-c2-CM	50.040	120.940	SOURCE3	254	1.3150
ca-c2-n2	71.29	126.01	SOURCE3	1	0.0000
C -CM-na	69.83	121.38	SOURCE3	26	6.9463
CM-CM-na	70.0	121.20			
CM-na-c2	67.80	110.37	SOURCE3	6	0.5121
CM-na-n2	65.80	125.00	SOURCE3	1	
DIHEDRAL					
X -CQ-NA-X	4	6.00	180.0	2.0	by analogy to X -CA-NA-X,
parm99.dat					
CM-CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
CM-CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
C -CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
C -CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
n -c -CM-C	4	4.000	180.0	2.	optimized by Junmei Wang,
Jan-2013					
CM-CM-c -n	4	4.000	180.0	2.	optimized by Junmei Wang,
Jan-2013					
X -c3-CM-X	6	0.000	0.000	2.	JCC,7, (1986),230
X -c2-CM-X	4	26.600	180.000	2.000	c2=c2 double bond,
interp.l.bsd.on C6H6					
X -CM-na-X	4	2.500	180.000	2.000	

NONB

8.6.5. Model4

ABI

@<TRIPOS>MOLECULE

ABI

71 74 1 0 0

SMALL

resp

@<TRIPOS>ATOM

1	HC1	9.8980	-5.4250	-0.2580	H1	1	ABI	0.299385
2	HC2	10.3610	-4.1810	0.9380	H1	1	ABI	0.299385
3	HC3	9.4220	-5.6160	1.4390	H1	1	ABI	0.299385
4	H7"	8.1880	-0.1910	0.6860	ha	1	ABI	0.212467
5	H8'1	-6.9310	0.2930	-1.8650	H1	1	ABI	0.377267
6	H8'2	-6.9290	-1.4670	-1.6590	H1	1	ABI	0.377267
7	H7	-3.1490	2.8670	-0.9480	ha	1	ABI	0.217686
8	H1L	-0.4940	1.4740	-0.3900	ha	1	ABI	0.252844
9	H0L	5.2460	-0.0360	-0.0950	ha	1	ABI	0.261679
10	H11L	2.6820	6.7870	-0.1360	ha	1	ABI	0.282344
11	H12L	0.3160	6.1050	-0.4820	ha	1	ABI	0.237686
12	H4L	1.4600	1.9660	-0.3280	ha	1	ABI	0.299116
13	H9L	5.0740	6.1440	0.2150	ha	1	ABI	0.282363
14	H6L	3.7720	1.3450	-0.0020	ha	1	ABI	0.294156
15	H8L	6.7930	4.3640	0.4610	ha	1	ABI	0.237926
16	H6	-5.0880	1.5410	-1.1290	H4	1	ABI	0.284436
17	H6"	9.2090	-2.3000	0.7740	H4	1	ABI	0.282277
18	CC1	9.5850	-4.8880	0.6410	CT	1	ABI	-0.905467

19	C7"	7.2200	-0.5970	0.3910	c2	1	ABI	-0.097566
20	C8'	-6.6660	-0.5250	-1.1870	CT	1	ABI	-1.024424
21	O13L	7.6450	2.1350	0.6840	o	1	ABI	-0.681160
22	C7	-2.4990	2.0110	-0.7680	c2	1	ABI	-0.121851
23	C1L	-1.1900	2.2780	-0.5820	c2	1	ABI	-0.581275
24	O2L	-1.5200	4.6090	-0.8350	o	1	ABI	-0.679453
25	C0L	6.2280	0.2990	0.2040	c2	1	ABI	-0.601812
26	C2L	-0.7290	3.6880	-0.6310	c	1	ABI	0.727816
27	C13L	6.5130	1.7410	0.3980	c	1	ABI	0.729389
28	C11L	2.4090	5.7350	-0.1780	ca	1	ABI	-0.391046
29	C12L	1.1010	5.3660	-0.3680	ca	1	ABI	-0.270639
30	C3L	0.7250	3.9920	-0.4200	ca	1	ABI	0.032919
31	C4L	1.7040	3.0230	-0.2850	ca	1	ABI	-0.448024
32	C5L	3.0670	3.3740	-0.0940	ca	1	ABI	0.282537
33	C10L	3.4300	4.7570	-0.0340	ca	1	ABI	0.184025
34	C9L	4.7950	5.0940	0.1670	ca	1	ABI	-0.391404
35	C6L	4.0770	2.3860	0.0400	ca	1	ABI	-0.443389
36	C7L	5.4030	2.7360	0.2340	ca	1	ABI	0.030775
37	C8L	5.7500	4.1160	0.3010	ca	1	ABI	-0.270148
38	H3	-2.7990	-2.5650	-0.3440	H	1	ABI	0.465101
39	O4	-1.2360	-0.6830	-0.3210	O	1	ABI	-0.749021
40	C4	-2.4310	-0.5430	-0.5220	C	1	ABI	0.900570
41	O2	-5.2610	-2.8010	-0.6520	O	1	ABI	-0.764019
42	N3	-3.2630	-1.6830	-0.5350	NA	1	ABI	-0.701221
43	C6	-4.5020	0.6440	-0.9710	CM	1	ABI	-0.123054
44	C2	-4.6230	-1.7570	-0.7240	C	1	ABI	0.907322
45	N1	-5.2180	-0.5260	-1.0120	N*	1	ABI	0.289300
46	C5	-3.1540	0.7150	-0.7570	CM	1	ABI	-0.250384
47	H3"	5.2750	-4.6740	-0.4930	H	1	ABI	0.469041
48	O4"	4.8430	-2.2620	-0.3930	O	1	ABI	-0.754807
49	C4"	5.9340	-2.7540	-0.1440	C	1	ABI	0.906914
50	O2"	7.2730	-6.1190	-0.0880	O	1	ABI	-0.771815
51	N3"	6.1000	-4.1490	-0.2240	NA	1	ABI	-0.717334
52	C6"	8.2820	-2.7840	0.4830	CM	1	ABI	-0.116985
53	C2"	7.2320	-4.9060	0.0110	C	1	ABI	0.909228
54	N1"	8.3500	-4.1450	0.3800	N*	1	ABI	0.287088
55	C5"	7.1580	-2.0380	0.2480	CM	1	ABI	-0.281863
56	C7'	-7.3660	-0.2750	0.1680	C	1	ABI	0.933011
57	O7'	-6.8650	0.5350	0.9390	O	1	ABI	-0.730295
58	N4'	-8.5110	-0.9630	0.4500	N	1	ABI	-0.100153
59	C3'	-9.2040	-1.8240	-0.5190	CT	1	ABI	-0.403446
60	H3'1	-10.2820	-1.6700	-0.3880	H1	1	ABI	0.210189
61	H3'2	-8.9840	-1.5090	-1.5410	H1	1	ABI	0.210189
62	C2'	-8.8620	-3.3190	-0.3610	CT	1	ABI	0.240285
63	H2'1	-7.7820	-3.4520	-0.4490	H1	1	ABI	0.102330
64	H2'2	-9.1460	-3.6540	0.6470	H1	1	ABI	0.102330
65	N1'	-9.5290	-4.1110	-1.3760	N	1	ABI	-0.877097
66	H1'1	-10.5430	-4.1020	-1.3960	H	1	ABI	0.480588
67	C5'	-9.2550	-0.6230	1.6760	CT	1	ABI	-0.570402
68	H5'1	-9.8900	0.2520	1.4990	H1	1	ABI	0.288810
69	H5'2	-9.9070	-1.4740	1.8990	H1	1	ABI	0.288810
70	C'	-8.4740	-0.2770	2.9590	C	1	ABI	0.622967
71	O1'	-8.7820	0.7300	3.5790	O	1	ABI	-0.747267

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73 67 70 1
74 70 71 1
@<TRIPOS>SUBSTRUCTURE
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@<TRIPOS>HEADTAIL
N1' 1
C' 1
@<TRIPOS>RESIDUECONNECT
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```

frcmod.pna

PNA frcmod file: frcmod.pna
 MASS

BOND

CQ-NA	502.0	1.324	by analogy to CQ-NC, parm99.dat
CM-c	349.7	1.487	SOURCE1 from gaff.dat
n3-Zn	50.4	2.089	ZAFF
c3-N*	337.0	1.475	JCC, 7, (1986), 230; ADE, CYT, GUA, THY, URA
CM-c3	317.0	1.510	by analogy parm99
c2-CM	357.2	1.4800	SOUECE3 1

ANGL

C -CT-N*	50.00	110.13	FF cst by analogy to CT-CT-N*; Eq.val. from
QM geo.opt.			
CQ-NA-C	70.0	118.60	by analogy to CA-NC-CQ, parm99.dat
H5-CQ-NA	50.0	115.45	by analogy to H5-CQ-NC, parm99.dat
NC-CQ-NA	70.0	129.10	by analogy to NC-CQ-NC, parm99.dat
CQ-NA-H	50.0	118.00	by analogy to CA-NA-H, parm99.dat
CM-c -o	68.670	123.44	SOURCE3 from gaff.dat
c -CM-C	62.650	120.000	SOURCE3 from gaff.dat
c -CM-CM	64.640	120.140	SOURCE3 from gaff.dat
n -c -CM	68.470	115.140	SOURCE4 from gaff.dat
N*-c3-h1	50.0	109.50	parm99
N*-c3-c3	67.530	112.140	SOURCE4
C-N*-c3	70.0	117.60	parm99
CM-N*-c3	70.0	121.20	parm99
CM-CM-c3	70.0	119.70	parm99
CM-c3-n3	66.18	112.13	SOURCE4 387 1.2309
CM-c3-h1	46.78	110.95	SOURCE3 12 1.1170
C -CM-c3	70.0	119.70	by parm99
c2-c2-CM	66.880	117.000	SOURCE3 1
c2-c -ca	67.170	116.780	SOURCE3 1
c2-CM-C	67.930	120.700	SOURCE3 1
c2-CM-CM	64.690	120.600	SOURCE3
ha-c2-CM	50.040	120.940	SOURCE3 254 1.3150

DIHEDRAL

X -CQ-NA-X	4	6.00	180.0	2.0	by analogy to X -CA-NA-X,
parm99.dat					
CM-CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
CM-CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
C -CM-c -o	1	2.175	180.0	-2.	Junmei et al, 1999
C -CM-c -o	1	0.30	0.0	3.	Junmei et al, 1999
n -c -CM-C	4	4.000	180.0	2.	optimized by Junmei Wang,

Jan-2013

CM-CM-c -n	4	4.000	180.0	2.	optimized by Junmei Wang, Jan-2013
X -c3-CM-X	6	0.000	0.000	2.	JCC,7, (1986),230
X -c2-CM-X	4	26.600	180.000	2.000	c2=c2 double bond, intrpol.bsd.on C6H6

NONB

8.7. *Ab initio* study on model4 analogue

8.7.1. Model4 in vacuum

HETATM	1	H	0	8.868	-3.569	-0.625	H
HETATM	2	H	0	9.028	-2.258	0.579	H
HETATM	3	H	0	8.505	-3.891	1.080	H
HETATM	4	H	0	5.896	1.039	0.438	H
HETATM	5	H	0	-5.896	1.038	-0.439	H
HETATM	6	H	0	-2.988	0.438	0.252	H
HETATM	7	H	0	2.988	0.438	-0.251	H
HETATM	8	H	0	-1.247	6.360	-0.082	H
HETATM	9	H	0	-3.379	5.087	-0.218	H
HETATM	10	H	0	-1.209	1.383	-0.069	H
HETATM	11	H	0	1.247	6.360	0.083	H
HETATM	12	H	0	1.209	1.383	0.068	H
HETATM	13	H	0	3.379	5.087	0.219	H
HETATM	14	H	0	-7.426	-0.738	-0.477	H
HETATM	15	H	0	7.426	-0.737	0.474	H
HETATM	16	C	0	8.453	-3.139	0.290	C
HETATM	17	C	0	5.056	0.399	0.167	C
HETATM	18	O	0	4.782	3.150	0.459	O
HETATM	19	C	0	-5.056	0.399	-0.168	C
HETATM	20	C	0	-3.862	1.011	-0.024	C
HETATM	21	O	0	-4.782	3.150	-0.460	O
HETATM	22	C	0	3.862	1.011	0.025	C
HETATM	23	C	0	-3.776	2.477	-0.228	C
HETATM	24	C	0	3.776	2.477	0.227	C
HETATM	25	C	0	-1.245	5.272	-0.078	C
HETATM	26	C	0	-2.425	4.576	-0.150	C
HETATM	27	C	0	-2.440	3.152	-0.136	C
HETATM	28	C	0	-1.238	2.468	-0.063	C
HETATM	29	C	0	-0.000	3.160	0.000	C
HETATM	30	C	0	-0.000	4.592	0.000	C
HETATM	31	C	0	1.245	5.272	0.079	C
HETATM	32	C	0	1.238	2.468	0.063	C
HETATM	33	C	0	2.440	3.151	0.136	C
HETATM	34	C	0	2.425	4.576	0.150	C
HETATM	35	H	0	-4.204	-4.034	0.735	H
HETATM	36	O	0	-3.168	-1.816	0.585	O
HETATM	37	C	0	-4.354	-2.012	0.371	C
HETATM	38	O	0	-6.517	-4.918	0.402	O
HETATM	39	N	0	-4.872	-3.316	0.477	N
HETATM	40	C	0	-6.647	-1.441	-0.200	C
HETATM	41	C	0	-6.167	-3.758	0.282	C
HETATM	42	N	0	-7.060	-2.738	-0.073	N
HETATM	43	C	0	-5.363	-1.009	-0.004	C
HETATM	44	H	0	4.204	-4.035	-0.733	H

HETATM	45	O	0	3.168	-1.816	-0.583	O
HETATM	46	C	0	4.354	-2.012	-0.370	C
HETATM	47	O	0	6.517	-4.918	-0.401	O
HETATM	48	N	0	4.872	-3.316	-0.476	N
HETATM	49	C	0	6.647	-1.441	0.198	C
HETATM	50	C	0	6.167	-3.758	-0.282	C
HETATM	51	N	0	7.061	-2.737	0.072	N
HETATM	52	C	0	5.363	-1.009	0.003	C
HETATM	53	C	0	-8.453	-3.139	-0.292	C
HETATM	54	H	0	-8.504	-3.895	-1.079	H
HETATM	55	H	0	-8.870	-3.565	0.624	H
HETATM	56	H	0	-9.027	-2.259	-0.585	H
END							

8.7.2. Linear analogue of Model4 in vacuum

HETATM	1	H	0	11.543	-1.904	0.886	H
HETATM	2	H	0	10.438	-2.993	-0.000	H
HETATM	3	H	0	11.543	-1.903	-0.886	H
HETATM	4	H	0	6.014	-1.938	-0.000	H
HETATM	5	H	0	-6.014	-1.938	-0.000	H
HETATM	6	H	0	-5.079	0.962	0.000	H
HETATM	7	H	0	5.079	0.962	0.000	H
HETATM	8	H	0	-1.253	3.227	-0.000	H
HETATM	9	H	0	-3.363	2.001	-0.000	H
HETATM	10	H	0	-1.256	-1.745	0.000	H
HETATM	11	H	0	1.253	3.227	-0.000	H
HETATM	12	H	0	1.256	-1.745	-0.000	H
HETATM	13	H	0	3.363	2.001	-0.000	H
HETATM	14	H	0	-8.319	-2.364	0.000	H
HETATM	15	H	0	8.319	-2.364	-0.000	H
HETATM	16	C	0	10.913	-2.011	-0.000	C
HETATM	17	C	0	6.145	-0.856	-0.000	C
HETATM	18	O	0	3.630	-2.034	-0.000	O
HETATM	19	C	0	-6.145	-0.856	0.000	C
HETATM	20	C	0	-5.016	-0.116	0.000	C
HETATM	21	O	0	-3.630	-2.034	0.000	O
HETATM	22	C	0	5.016	-0.116	-0.000	C
HETATM	23	C	0	-3.703	-0.805	-0.000	C
HETATM	24	C	0	3.703	-0.805	-0.000	C
HETATM	25	C	0	-1.246	2.140	-0.000	C
HETATM	26	C	0	-2.433	1.444	-0.000	C
HETATM	27	C	0	-2.446	0.019	-0.000	C
HETATM	28	C	0	-1.238	-0.660	-0.000	C
HETATM	29	C	0	0.000	0.027	-0.000	C
HETATM	30	C	0	0.000	1.461	-0.000	C
HETATM	31	C	0	1.246	2.140	-0.000	C
HETATM	32	C	0	1.238	-0.660	-0.000	C
HETATM	33	C	0	2.446	0.019	-0.000	C
HETATM	34	C	0	2.433	1.444	-0.000	C
HETATM	35	H	0	-9.554	2.239	-0.000	H
HETATM	36	O	0	-7.114	1.992	0.000	O
HETATM	37	C	0	-7.879	1.039	0.000	C
HETATM	38	O	0	-11.481	0.649	0.000	O
HETATM	39	N	0	-9.268	1.266	-0.000	N
HETATM	40	C	0	-8.536	-1.300	0.000	C
HETATM	41	C	0	-10.301	0.347	0.000	C
HETATM	42	N	0	-9.866	-0.985	0.000	N

HETATM	43	C	0	-7.518	-0.387	0.000	C
HETATM	44	H	0	9.554	2.239	0.000	H
HETATM	45	O	0	7.114	1.992	0.000	O
HETATM	46	C	0	7.879	1.039	0.000	C
HETATM	47	O	0	11.481	0.649	0.000	O
HETATM	48	N	0	9.268	1.266	0.000	N
HETATM	49	C	0	8.536	-1.300	-0.000	C
HETATM	50	C	0	10.301	0.347	0.000	C
HETATM	51	N	0	9.866	-0.985	0.000	N
HETATM	52	C	0	7.518	-0.387	0.000	C
HETATM	53	C	0	-10.913	-2.011	0.000	C
HETATM	54	H	0	-11.543	-1.903	0.886	H
HETATM	55	H	0	-11.543	-1.904	-0.886	H
HETATM	56	H	0	-10.438	-2.993	0.000	H
END							

8.7.3. PES scan

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generated by VMD

H	11.542610	-1.903560	0.886190
H	10.438060	-2.992710	-0.000150
H	11.542580	-1.903400	-0.886340
H	6.013820	-1.937900	-0.000060
H	-6.013820	-1.937900	-0.000010
H	-5.078510	0.962270	0.000010
H	5.078510	0.962270	0.000010
H	-1.252530	3.227170	-0.000130
H	-3.363070	2.000880	-0.000090
H	-1.255840	-1.745470	0.000000
H	1.252530	3.227170	-0.000150
H	1.255840	-1.745470	-0.000040
H	3.363070	2.000880	-0.000140
H	-8.318880	-2.363730	0.000040
H	8.318880	-2.363730	-0.000060
C	10.912850	-2.010700	-0.000070
C	6.145290	-0.855720	-0.000020
O	3.630270	-2.034090	-0.000120
C	-6.145290	-0.855720	0.000000
C	-5.016160	-0.116180	0.000010
O	-3.630270	-2.034090	0.000030
C	5.016160	-0.116180	-0.000030
C	-3.702580	-0.804700	-0.000000
C	3.702580	-0.804700	-0.000080
C	-1.246410	2.139950	-0.000110
C	-2.432810	1.443970	-0.000080
C	-2.445500	0.018690	-0.000040
C	-1.238140	-0.659870	-0.000030
C	0.000000	0.027320	-0.000060
C	0.000000	1.461350	-0.000100
C	1.246400	2.139950	-0.000120
C	1.238140	-0.659870	-0.000060
C	2.445500	0.018690	-0.000080
C	2.432810	1.443970	-0.000110
H	-9.554090	2.239140	-0.000010
O	-7.113690	1.991540	0.000010
C	-7.878930	1.039450	0.000010
O	-11.480560	0.649090	0.000070
N	-9.267580	1.266150	-0.000010

C	-8.536430	-1.300440	0.000030
C	-10.300730	0.347200	0.000040
N	-9.866310	-0.984990	0.000050
C	-7.517530	-0.386510	0.000020
H	9.554090	2.239140	0.000230
O	7.113690	1.991540	0.000110
C	7.878930	1.039450	0.000100
O	11.480570	0.649090	0.000110
N	9.267580	1.266150	0.000190
C	8.536430	-1.300440	-0.000000
C	10.300730	0.347200	0.000110
N	9.866310	-0.984990	0.000030
C	7.517530	-0.386510	0.000030
C	-10.912850	-2.010700	0.000120
H	-11.542560	-1.903450	0.886410
H	-11.542630	-1.903510	-0.886120
H	-10.438070	-2.992710	0.000130

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generated by VMD

H	11.562535	-1.909432	0.664069
H	10.432876	-2.984544	-0.207638
H	11.509879	-1.878929	-1.107419
H	6.010105	-1.935086	-0.059218
H	-6.024895	-1.926991	0.241249
H	-5.057913	0.945289	-0.085641
H	5.072131	0.963326	0.014764
H	-1.255451	3.212644	0.363910
H	-3.366883	1.981674	0.354621
H	-1.256990	-1.754205	0.108290
H	1.248176	3.216818	0.290904
H	1.253608	-1.749342	0.039001
H	3.358725	1.995501	0.177644
H	-8.330005	-2.343455	0.168376
H	8.314178	-2.358075	-0.134559
C	10.906809	-2.002123	-0.204832
C	6.140617	-0.852883	-0.045281
O	3.627877	-2.034862	-0.006637
C	-6.145649	-0.852951	0.098599
C	-5.010260	-0.123967	0.062561
O	-3.639709	-2.038301	0.313308
C	5.011219	-0.115129	0.000376
C	-3.705075	-0.813072	0.209360
C	3.699073	-0.805878	0.027473
C	-1.249471	2.127311	0.299916
C	-2.435273	1.430418	0.288966
C	-2.446761	0.007323	0.212885
C	-1.240173	-0.669558	0.155321
C	-0.002672	0.019090	0.157361
C	-0.003395	1.451347	0.230753
C	1.242467	2.131051	0.234634
C	1.235356	-0.665074	0.092579
C	2.442058	0.014635	0.096896
C	2.428826	1.438029	0.169561
H	-9.521233	2.229643	-0.448962
O	-7.086403	1.976682	-0.291007
C	-7.859368	1.033721	-0.218484
O	-11.459863	0.657146	-0.355952
N	-9.244064	1.262786	-0.317931
C	-8.537544	-1.287340	0.028643
C	-10.284307	0.353815	-0.260014
N	-9.862734	-0.970083	-0.077903
C	-7.511582	-0.383141	-0.029551

H	9.545877	2.245529	-0.091930
O	7.106631	1.995129	-0.025033
C	7.872188	1.044104	-0.063893
O	11.472579	0.657899	-0.175022
N	9.260121	1.272349	-0.100339
C	8.530962	-1.294686	-0.122882
C	10.293433	0.354733	-0.145994
N	9.860065	-0.977781	-0.156198
C	7.511890	-0.382059	-0.077718
C	-10.917870	-1.984330	-0.004574
H	-11.581774	-1.774574	0.837415
H	-11.510248	-1.974858	-0.922358
H	-10.452671	-2.962264	0.125701

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generated by VMD

H	11.564872	-1.920660	0.385697
H	10.402583	-2.974864	-0.468556
H	11.445558	-1.847413	-1.381267
H	5.990149	-1.931088	-0.116298
H	-6.050785	-1.891751	0.490593
H	-5.010286	0.915433	-0.094130
H	5.055054	0.964498	0.049497
H	-1.255373	3.197792	0.689924
H	-3.371496	1.962560	0.692584
H	-1.266009	-1.761322	0.292404
H	1.243844	3.204519	0.541478
H	1.240685	-1.751546	0.139963
H	3.349988	1.988548	0.329539
H	-8.349317	-2.299431	0.289139
H	8.288795	-2.351156	-0.293934
C	10.876833	-1.992633	-0.459878
C	6.121225	-0.849277	-0.086233
O	3.612668	-2.034578	0.023433
C	-6.144361	-0.839724	0.220182
C	-4.996713	-0.130379	0.179522
O	-3.667434	-2.020805	0.709213
C	4.994316	-0.113629	0.017574
C	-3.714430	-0.811943	0.481676
C	3.684495	-0.806489	0.080930
C	-1.252713	2.115348	0.588067
C	-2.438814	1.418963	0.583156
C	-2.451646	-0.000706	0.463372
C	-1.248821	-0.677778	0.361960
C	-0.011322	0.011597	0.347142
C	-0.009717	1.441224	0.463099
C	1.236169	2.120979	0.451988
C	1.224100	-0.669345	0.226056
C	2.430545	0.010743	0.214316
C	2.419975	1.431175	0.330745
H	-9.426795	2.188998	-0.896720
O	-7.013661	1.938455	-0.532922
C	-7.803335	1.013867	-0.418381
O	-11.389146	0.647411	-0.783168
N	-9.173927	1.240081	-0.642913
C	-8.531181	-1.262877	0.023640
C	-10.228373	0.349095	-0.566972
N	-9.840441	-0.950154	-0.212997
C	-7.490683	-0.377012	-0.055296
H	9.523285	2.250913	-0.200602
O	7.088428	1.998159	-0.043331
C	7.851433	1.048498	-0.132663
O	11.444517	0.666203	-0.394033

N	9.236987	1.278037	-0.218911
C	8.506360	-1.288179	-0.268051
C	10.267275	0.362048	-0.325209
N	9.833222	-0.970097	-0.347451
C	7.490275	-0.377157	-0.162887
C	-10.912106	-1.944771	-0.114826
H	-11.643213	-1.634405	0.635234
H	-11.422809	-2.041469	-1.075769
H	-10.473832	-2.902302	0.169365

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generated by VMD

H	11.536103	-1.951524	0.194675
H	10.351790	-2.984182	-0.655672
H	11.381438	-1.844047	-1.567787
H	5.953333	-1.928209	-0.184694
H	-6.071867	-1.839372	0.695814
H	-4.962651	0.898644	-0.073289
H	5.035370	0.967245	0.060700
H	-1.241833	3.205326	0.960559
H	-3.367445	1.973998	0.983763
H	-1.285300	-1.743169	0.434261
H	1.250109	3.206712	0.730540
H	1.213710	-1.736175	0.195019
H	3.343792	1.989889	0.422506
H	-8.354798	-2.259652	0.377030
H	8.245248	-2.354755	-0.421042
C	10.830770	-2.004375	-0.637990
C	6.090112	-0.847750	-0.136708
O	3.579662	-2.024940	0.004297
C	-6.138692	-0.815633	0.326518
C	-4.982996	-0.118901	0.293275
O	-3.701827	-1.957246	1.079144
C	4.969326	-0.109815	0.008844
C	-3.728030	-0.777338	0.731118
C	3.658545	-0.798975	0.090983
C	-1.248179	2.126624	0.824769
C	-2.437004	1.434960	0.833679
C	-2.458525	0.020082	0.671954
C	-1.263823	-0.661013	0.523400
C	-0.024194	0.024863	0.486581
C	-0.013041	1.450801	0.641965
C	1.235094	2.126065	0.611720
C	1.204074	-0.656602	0.310330
C	2.412669	0.019219	0.281346
C	2.411844	1.435923	0.436167
H	-9.320211	2.115601	-1.247705
O	-6.936155	1.889675	-0.713545
C	-7.738975	0.980567	-0.571522
O	-11.297923	0.593894	-1.128816
N	-9.091253	1.190813	-0.899609
C	-8.511213	-1.248820	0.014104
C	-10.155292	0.312102	-0.815718
N	-9.800488	-0.953741	-0.329870
C	-7.460900	-0.375400	-0.073320
H	9.503080	2.239525	-0.273344
O	7.071379	1.994368	-0.064438
C	7.827607	1.043244	-0.188681
O	11.412062	0.650367	-0.538570
N	9.211893	1.268362	-0.302775
C	8.468356	-1.293360	-0.381170
C	10.235229	0.350073	-0.448718
N	9.794563	-0.979577	-0.485061

C	7.459176	-0.380061	-0.236514
C	-10.883851	-1.934106	-0.217219
H	-11.662189	-1.556270	0.449665
H	-11.328336	-2.116379	-1.198517
H	-10.473323	-2.862757	0.181275
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generated by VMD			
H	11.486932	-2.014568	-0.036404
H	10.270168	-3.007380	-0.888401
H	11.287019	-1.847268	-1.789607
H	5.895320	-1.924897	-0.274947
H	-6.095401	-1.757506	0.910244
H	-4.910124	0.888119	-0.053808
H	5.015460	0.968687	0.098261
H	-1.215368	3.238671	1.210322
H	-3.354996	2.021345	1.257201
H	-1.315630	-1.698290	0.580273
H	1.266924	3.224875	0.902521
H	1.172345	-1.703393	0.255056
H	3.339906	1.999689	0.503006
H	-8.356378	-2.204021	0.493412
H	8.176081	-2.365529	-0.582727
C	10.759435	-2.033225	-0.851223
C	6.045018	-0.848314	-0.191641
O	3.526947	-2.005301	-0.025544
C	-6.130517	-0.775453	0.437515
C	-4.968140	-0.089569	0.406865
O	-3.746491	-1.840198	1.454724
C	4.936291	-0.104997	0.008498
C	-3.746944	-0.705676	0.980395
C	3.620524	-0.783608	0.099090
C	-1.236025	2.163860	1.047748
C	-2.430206	1.481846	1.073219
C	-2.467996	0.072275	0.877286
C	-1.285591	-0.617634	0.685669
C	-0.041424	0.059752	0.624418
C	-0.013320	1.481535	0.812323
C	1.239262	2.147341	0.759941
C	1.175117	-0.626767	0.395723
C	2.388230	0.039904	0.346179
C	2.404384	1.452511	0.533264
H	-9.183393	1.984819	-1.624500
O	-6.840531	1.812807	-0.915667
C	-7.657220	0.923250	-0.735717
O	-11.176404	0.485428	-1.477993
N	-8.983974	1.099751	-1.170923
C	-8.481833	-1.235012	0.020846
C	-10.057936	0.235576	-1.066151
N	-9.744176	-0.975082	-0.433387
C	-7.421585	-0.375937	-0.086951
H	9.486742	2.207825	-0.304429
O	7.058625	1.979601	-0.043181
C	7.801131	1.025995	-0.219988
O	11.371368	0.610354	-0.673582
N	9.184501	1.241257	-0.360916
C	8.411632	-1.308498	-0.510957
C	10.193962	0.318686	-0.564756
N	9.738243	-1.004398	-0.636713
C	7.416211	-0.390982	-0.308939
C	-10.840567	-1.936727	-0.289525
H	-11.646867	-1.500320	0.304724
H	-11.238932	-2.199242	-1.272236

H	-10.457870	-2.830128	0.205574
56			
generated by VMD			
H	11.390501	-2.184673	-0.281498
H	10.146338	-3.042139	-1.234798
H	11.197776	-1.820410	-2.005399
H	5.802499	-1.903040	-0.504267
H	-6.103592	-1.622231	1.164922
H	-4.870404	0.892306	-0.060462
H	5.012090	0.953818	0.203640
H	-1.167469	3.347423	1.303524
H	-3.322340	2.156849	1.440835
H	-1.356102	-1.592214	0.708302
H	1.295997	3.300630	0.878368
H	1.111353	-1.629353	0.265957
H	3.334259	2.046675	0.393996
H	-8.339573	-2.137788	0.693801
H	8.070762	-2.375841	-0.858442
C	10.663789	-2.092646	-1.092014
C	5.984888	-0.847965	-0.299443
O	3.435703	-1.947406	-0.211667
C	-6.117173	-0.707199	0.571764
C	-4.954926	-0.022562	0.512556
O	-3.782695	-1.609866	1.844178
C	4.899311	-0.098332	-0.014236
C	-3.762442	-0.554106	1.215745
C	3.563045	-0.741790	0.005416
C	-1.209563	2.272448	1.146155
C	-2.410396	1.606312	1.225304
C	-2.474640	0.197442	1.035405
C	-1.311658	-0.510855	0.802896
C	-0.062241	0.150374	0.682667
C	-0.007532	1.572474	0.861273
C	1.249638	2.222118	0.748552
C	1.134241	-0.552671	0.404608
C	2.353036	0.097600	0.302936
C	2.395442	1.510980	0.478711
H	-9.067243	1.746274	-1.971140
O	-6.763861	1.685682	-1.127233
C	-7.586271	0.818184	-0.879562
O	-11.061088	0.258586	-1.739632
N	-8.889215	0.926659	-1.400730
C	-8.443050	-1.236123	0.098516
C	-9.964817	0.072803	-1.243057
N	-9.681224	-1.045878	-0.447261
C	-7.380765	-0.387848	-0.062496
H	9.516218	2.097912	-0.079349
O	7.082272	1.914791	0.158408
C	7.796327	0.964551	-0.122820
O	11.353365	0.495754	-0.625234
N	9.185633	1.152816	-0.241551
C	8.337616	-1.340726	-0.670524
C	10.167628	0.228851	-0.547299
N	9.672996	-1.064092	-0.763864
C	7.369547	-0.422023	-0.366605
C	-10.780919	-1.991989	-0.239910
H	-11.625899	-1.485290	0.231838
H	-11.112674	-2.397657	-1.198591
H	-10.428570	-2.800394	0.401997
56			
generated by VMD			
H	11.294396	-2.312974	-0.464550

H	10.016449	-3.097900	-1.435300
H	11.086329	-1.868820	-2.167843
H	5.709028	-1.882935	-0.620764
H	-6.117898	-1.477838	1.352397
H	-4.822178	0.889290	-0.086695
H	4.998381	0.951705	0.243987
H	-1.109265	3.445127	1.503131
H	-3.283509	2.290409	1.665275
H	-1.406063	-1.472108	0.776978
H	1.341011	3.363992	1.012397
H	1.043256	-1.541456	0.264313
H	3.342282	2.085307	0.442028
H	-8.327491	-2.054105	0.842857
H	7.961647	-2.396274	-1.015128
C	10.558773	-2.168073	-1.259123
C	5.920486	-0.844907	-0.363470
O	3.345902	-1.884895	-0.301030
C	-6.102425	-0.636637	0.658444
C	-4.938271	0.043222	0.579324
O	-3.834336	-1.369167	2.147175
C	4.857038	-0.083992	-0.029151
C	-3.783504	-0.398165	1.397283
C	3.505125	-0.694368	-0.028528
C	-1.176179	2.375908	1.317398
C	-2.387126	1.729941	1.412022
C	-2.483485	0.328983	1.184765
C	-1.342679	-0.394369	0.900073
C	-0.083823	0.247124	0.763169
C	0.003867	1.661876	0.980888
C	1.270568	2.290577	0.854126
C	1.090834	-0.470001	0.433993
C	2.319704	0.159261	0.321636
C	2.395267	1.565837	0.536417
H	-8.925651	1.483613	-2.294988
O	-6.664986	1.535352	-1.340850
C	-7.498899	0.699392	-1.031699
O	-10.928598	0.022440	-1.984355
N	-8.775552	0.738390	-1.623453
C	-8.402444	-1.229769	0.140721
C	-9.857672	-0.096616	-1.417166
N	-9.613202	-1.111333	-0.481050
C	-7.333484	-0.400552	-0.068730
H	9.528343	1.996296	-0.034326
O	7.092509	1.863405	0.218204
C	7.779529	0.910228	-0.115476
O	11.318910	0.376480	-0.674762
N	9.172157	1.068862	-0.238676
C	8.256570	-1.378748	-0.779269
C	10.127433	0.136130	-0.597883
N	9.597934	-1.132031	-0.871059
C	7.315136	-0.451614	-0.422147
C	-10.721077	-2.033821	-0.217813
H	-11.591396	-1.479410	0.140315
H	-10.998979	-2.558464	-1.135074
H	-10.403356	-2.753215	0.537979
56			
generated by VMD			
H	11.165831	-2.431311	-0.702036
H	9.845806	-3.149990	-1.667907
H	10.926145	-1.919215	-2.381953
H	5.589408	-1.866551	-0.702822
H	-6.157228	-1.303771	1.538817

H	-4.739594	0.857157	-0.098518
H	4.967806	0.949565	0.282908
H	-1.053106	3.551016	1.723284
H	-3.246070	2.438441	1.922876
H	-1.471159	-1.343366	0.905177
H	1.381536	3.428571	1.169329
H	0.960906	-1.453907	0.337382
H	3.340070	2.120411	0.519763
H	-8.333075	-1.938181	0.951711
H	7.818684	-2.415827	-1.175608
C	10.414391	-2.240101	-1.471527
C	5.832058	-0.844053	-0.412858
O	3.236941	-1.829607	-0.311581
C	-6.084781	-0.559148	0.745233
C	-4.910867	0.103594	0.660328
O	-3.931637	-1.077353	2.483057
C	4.795652	-0.071927	-0.023916
C	-3.820076	-0.224826	1.607816
C	3.430117	-0.651364	-0.009142
C	-1.146777	2.486656	1.521662
C	-2.368340	1.864125	1.637275
C	-2.500062	0.470612	1.386523
C	-1.384646	-0.270389	1.055349
C	-0.114663	0.347045	0.901389
C	0.008941	1.754867	1.142237
C	1.284793	2.359988	0.993093
C	1.035301	-0.387360	0.527299
C	2.273686	0.218776	0.392860
C	2.385273	1.618929	0.631565
H	-8.672719	1.171554	-2.644886
O	-6.482396	1.337955	-1.550346
C	-7.345367	0.551188	-1.195763
O	-10.708944	-0.232677	-2.291115
N	-8.577703	0.517613	-1.875336
C	-8.349588	-1.210460	0.146577
C	-9.680620	-0.281785	-1.640890
N	-9.512004	-1.171170	-0.569979
C	-7.259736	-0.416602	-0.090783
H	9.515795	1.894762	-0.057488
O	7.083093	1.811095	0.243002
C	7.738871	0.855490	-0.142536
O	11.250834	0.259427	-0.802801
N	9.132199	0.985067	-0.289981
C	8.143890	-1.415791	-0.907072
C	10.055646	0.045007	-0.708477
N	9.488622	-1.197686	-1.020481
C	7.234132	-0.481589	-0.491271
C	-10.646230	-2.049194	-0.269462
H	-11.526740	-1.452222	-0.020747
H	-10.881965	-2.666727	-1.139235
H	-10.379911	-2.685855	0.575224
56			
generated by VMD			
H	11.023061	-2.561355	-0.884480
H	9.681513	-3.184041	-1.886461
H	10.802689	-1.953413	-2.534871
H	5.480412	-1.845752	-0.769373
H	-6.210670	-1.094899	1.687839
H	-4.635711	0.775512	-0.152025
H	4.942783	0.949222	0.320442
H	-0.996422	3.642311	1.928193
H	-3.207832	2.569810	2.145123

H	-1.538101	-1.212049	0.955053
H	1.427837	3.480799	1.340610
H	0.880772	-1.361269	0.360479
H	3.346012	2.149312	0.620075
H	-8.371660	-1.750889	1.076934
H	7.685894	-2.429393	-1.304471
C	10.279379	-2.304883	-1.642759
C	5.752223	-0.839610	-0.449617
O	3.137382	-1.768230	-0.334973
C	-6.070387	-0.476732	0.800311
C	-4.874544	0.142949	0.693990
O	-4.048254	-0.763557	2.736086
C	4.741195	-0.057095	-0.017189
C	-3.856850	-0.058953	1.750928
C	3.362571	-0.604936	0.001408
C	-1.116339	2.585645	1.701528
C	-2.348783	1.986128	1.824362
C	-2.514723	0.603479	1.538047
C	-1.426395	-0.148328	1.149945
C	-0.143492	0.444586	0.998366
C	0.016508	1.840309	1.281996
C	1.303821	2.420828	1.132182
C	0.982895	-0.303306	0.582717
C	2.233148	0.278260	0.448664
C	2.381358	1.666830	0.730654
H	-8.409360	0.790470	-2.955692
O	-6.275637	1.060430	-1.772647
C	-7.183821	0.360101	-1.355726
O	-10.509768	-0.489601	-2.515473
N	-8.378846	0.257959	-2.092904
C	-8.319777	-1.152310	0.172997
C	-9.519988	-0.467456	-1.806712
N	-9.440226	-1.189781	-0.607214
C	-7.191729	-0.432120	-0.115902
H	9.507262	1.797321	-0.068125
O	7.079202	1.760006	0.276932
C	7.704382	0.803794	-0.155064
O	11.188343	0.148789	-0.902514
N	9.097456	0.905874	-0.325252
C	8.040151	-1.447557	-1.007056
C	9.990090	-0.040317	-0.793782
N	9.387389	-1.257027	-1.139283
C	7.161097	-0.507722	-0.541030
C	-10.619031	-1.982096	-0.246071
H	-11.492658	-1.332558	-0.155850
H	-10.820998	-2.727414	-1.018971
H	-10.426885	-2.478763	0.705853

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generated by VMD

H	10.916286	-2.557835	-1.076633
H	9.514832	-3.256622	-1.936655
H	10.567876	-2.058557	-2.741313
H	5.369055	-1.870067	-0.682879
H	-6.316017	-0.867461	1.787568
H	-4.467225	0.627331	-0.139490
H	4.875640	0.974012	0.293632
H	-0.959013	3.664366	2.268998
H	-3.189817	2.630301	2.471476
H	-1.622449	-1.123970	1.069099
H	1.461393	3.467319	1.675689
H	0.794214	-1.310914	0.483382
H	3.342914	2.126401	0.884443

H	-8.453477	-1.494144	1.067036
H	7.545849	-2.473284	-1.306595
C	10.115880	-2.357900	-1.792737
C	5.652491	-0.848989	-0.426969
O	3.046219	-1.767402	-0.167663
C	-6.056972	-0.397165	0.838210
C	-4.819690	0.140361	0.761150
O	-4.244262	-0.468207	2.991417
C	4.661891	-0.049205	0.020875
C	-3.918958	0.059649	1.933770
C	3.288196	-0.597988	0.134409
C	-1.103037	2.619388	2.005014
C	-2.347266	2.042748	2.116441
C	-2.546046	0.676414	1.777726
C	-1.482970	-0.077630	1.330397
C	-0.184595	0.488802	1.204996
C	0.010217	1.866281	1.548279
C	1.310253	2.422090	1.416843
C	0.921362	-0.265807	0.749345
C	2.183797	0.292427	0.628290
C	2.366838	1.662833	0.971934
H	-7.959655	0.414520	-3.273190
O	-5.927271	0.744704	-1.937160
C	-6.914057	0.164222	-1.514993
O	-10.170961	-0.674356	-2.863143
N	-8.042121	0.015947	-2.344079
C	-8.280852	-1.041268	0.095868
C	-9.249469	-0.598595	-2.070942
N	-9.325508	-1.138456	-0.778546
C	-7.086539	-0.431376	-0.180432
H	9.408637	1.813427	-0.377049
O	7.003338	1.786307	0.101043
C	7.609139	0.811928	-0.317807
O	11.050620	0.131612	-1.222740
N	8.990481	0.909214	-0.567583
C	7.910375	-1.477201	-1.076507
C	9.861367	-0.055854	-1.038462
N	9.247662	-1.289998	-1.289944
C	7.052619	-0.518064	-0.610111
C	-10.582173	-1.805120	-0.426237
H	-11.413116	-1.099038	-0.494216
H	-10.773171	-2.630615	-1.115675
H	-10.501863	-2.185132	0.593077

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generated by VMD

H	10.916286	-2.557835	-1.076633
H	9.514832	-3.256622	-1.936655
H	10.567876	-2.058557	-2.741313
H	5.369055	-1.870067	-0.682879
H	-6.316017	-0.867461	1.787568
H	-4.467225	0.627331	-0.139490
H	4.875640	0.974012	0.293632
H	-0.959013	3.664366	2.268998
H	-3.189817	2.630301	2.471476
H	-1.622449	-1.123970	1.069099
H	1.461393	3.467319	1.675689
H	0.794214	-1.310914	0.483382
H	3.342914	2.126401	0.884443
H	-8.453477	-1.494144	1.067036
H	7.545849	-2.473284	-1.306595
C	10.115880	-2.357900	-1.792737
C	5.652491	-0.848989	-0.426969

O	3.046219	-1.767402	-0.167663
C	-6.056972	-0.397165	0.838210
C	-4.819690	0.140361	0.761150
O	-4.244262	-0.468207	2.991417
C	4.661891	-0.049205	0.020875
C	-3.918958	0.059649	1.933770
C	3.288196	-0.597988	0.134409
C	-1.103037	2.619388	2.005014
C	-2.347266	2.042748	2.116441
C	-2.546046	0.676414	1.777726
C	-1.482970	-0.077630	1.330397
C	-0.184595	0.488802	1.204996
C	0.010217	1.866281	1.548279
C	1.310253	2.422090	1.416843
C	0.921362	-0.265807	0.749345
C	2.183797	0.292427	0.628290
C	2.366838	1.662833	0.971934
H	-7.959655	0.414520	-3.273190
O	-5.927271	0.744704	-1.937160
C	-6.914057	0.164222	-1.514993
O	-10.170961	-0.674356	-2.863143
N	-8.042121	0.015947	-2.344079
C	-8.280852	-1.041268	0.095868
C	-9.249469	-0.598595	-2.070942
N	-9.325508	-1.138456	-0.778546
C	-7.086539	-0.431376	-0.180432
H	9.408637	1.813427	-0.377049
O	7.003338	1.786307	0.101043
C	7.609139	0.811928	-0.317807
O	11.050620	0.131612	-1.222740
N	8.990481	0.909214	-0.567583
C	7.910375	-1.477201	-1.076507
C	9.861367	-0.055854	-1.038462
N	9.247662	-1.289998	-1.289944
C	7.052619	-0.518064	-0.610111
C	-10.582173	-1.805120	-0.426237
H	-11.413116	-1.099038	-0.494216
H	-10.773171	-2.630615	-1.115675
H	-10.501863	-2.185132	0.593077

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generated by VMD

H	10.752528	-2.739484	-1.196435
H	9.338100	-3.301784	-2.131718
H	10.461488	-2.096634	-2.822493
H	5.240221	-1.827651	-0.820634
H	-6.383453	-0.517070	1.898384
H	-4.340913	0.457890	-0.161593
H	4.857290	0.955618	0.360532
H	-0.908585	3.841133	2.244310
H	-3.149327	2.845248	2.535184
H	-1.696278	-0.942417	1.102136
H	1.492742	3.589264	1.599215
H	0.705294	-1.187775	0.476545
H	3.329118	2.202445	0.778516
H	-8.521059	-1.143536	1.184859
H	7.397321	-2.481496	-1.460838
C	9.976971	-2.446484	-1.907926
C	5.564505	-0.841817	-0.486851
O	2.921608	-1.671625	-0.289685
C	-6.043695	-0.265065	0.893074
C	-4.772509	0.181570	0.792166
O	-4.372021	0.029767	3.135514

C	4.603294	-0.033019	0.006509
C	-3.950106	0.306900	2.017661
C	3.205663	-0.527110	0.066057
C	-1.077695	2.794319	2.003474
C	-2.327422	2.241536	2.160224
C	-2.559454	0.874057	1.847339
C	-1.526916	0.099548	1.364053
C	-0.219641	0.636939	1.206597
C	0.009768	2.014123	1.529559
C	1.316240	2.543516	1.359307
C	0.859852	-0.143581	0.731466
C	2.130116	0.387962	0.577596
C	2.347726	1.758160	0.900886
H	-7.664742	-0.228383	-3.414468
O	-5.687189	0.257152	-2.043900
C	-6.733885	-0.161669	-1.577116
O	-9.965933	-1.076789	-2.935526
N	-7.824465	-0.415624	-2.430425
C	-8.266031	-0.916901	0.154421
C	-9.085028	-0.887462	-2.116594
N	-9.267878	-1.134288	-0.748059
C	-7.018300	-0.450567	-0.162604
H	9.433185	1.637859	-0.197208
O	7.021918	1.684519	0.248787
C	7.590518	0.716867	-0.233026
O	11.011938	-0.050390	-1.144544
N	8.978357	0.769925	-0.459073
C	7.801942	-1.522202	-1.154298
C	9.812853	-0.196687	-0.989049
N	9.149073	-1.380109	-1.338326
C	6.980488	-0.562008	-0.628508
C	-10.584659	-1.639113	-0.349228
H	-11.361403	-0.929645	-0.643340
H	-10.784966	-2.594142	-0.840660
H	-10.594240	-1.770944	0.733484

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generated by VMD

H	10.613051	-2.877159	-1.231827
H	9.174441	-3.419854	-2.141395
H	10.313986	-2.247424	-2.861544
H	5.126849	-1.839643	-0.795385
H	-6.445934	-0.185131	1.907274
H	-4.224625	0.330505	-0.132916
H	4.821476	0.965425	0.356290
H	-0.853774	4.000700	2.282224
H	-3.118757	3.059215	2.597611
H	-1.778444	-0.758154	1.139659
H	1.538091	3.680699	1.633922
H	0.617217	-1.074691	0.520530
H	3.332910	2.243485	0.809205
H	-8.560463	-0.847380	1.157919
H	7.260418	-2.548508	-1.456264
C	9.834448	-2.575938	-1.936499
C	5.477330	-0.857551	-0.477480
O	2.818641	-1.623833	-0.241191
C	-6.027637	-0.130996	0.901433
C	-4.732162	0.243222	0.819413
O	-4.511780	0.517111	3.175341
C	4.540818	-0.021493	0.018025
C	-3.993690	0.563855	2.063458
C	3.133237	-0.482805	0.099678
C	-1.051821	2.957463	2.048060

C	-2.313854	2.438035	2.215278
C	-2.584220	1.076438	1.905822
C	-1.576967	0.276317	1.408882
C	-0.255079	0.775672	1.250987
C	0.012476	2.146731	1.571563
C	1.332071	2.639972	1.395753
C	0.801120	-0.034745	0.773353
C	2.084740	0.461919	0.613907
C	2.340158	1.826129	0.934610
H	-7.356271	-0.801264	-3.454147
O	-5.451303	-0.156456	-2.047217
C	-6.546709	-0.444059	-1.593065
O	-9.724007	-1.462778	-3.005810
N	-7.590114	-0.803622	-2.467125
C	-8.226337	-0.818423	0.125603
C	-8.890772	-1.163994	-2.169707
N	-9.175793	-1.155962	-0.796574
C	-6.938352	-0.464201	-0.174848
H	9.403436	1.538139	-0.265571
O	7.000107	1.643108	0.211840
C	7.540590	0.657974	-0.266928
O	10.931706	-0.194994	-1.214554
N	8.926108	0.677700	-0.511626
C	7.690239	-1.595196	-1.165761
C	9.731895	-0.312937	-1.041645
N	9.037602	-1.485196	-1.368731
C	6.897207	-0.611151	-0.640290
C	-10.539903	-1.530966	-0.414004
H	-11.258539	-0.853328	-0.880519
H	-10.756886	-2.548201	-0.748396
H	-10.626113	-1.472154	0.671718

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generated by VMD

H	10.511412	-2.913450	-1.224081
H	9.034096	-3.575904	-1.979510
H	10.106266	-2.477491	-2.893676
H	5.049644	-1.920117	-0.539612
H	-6.513734	0.093183	1.815941
H	-4.109000	0.248298	-0.068133
H	4.743126	1.005438	0.256861
H	-0.794520	4.078800	2.502628
H	-3.104043	3.212679	2.729518
H	-1.865345	-0.605832	1.192587
H	1.604419	3.678993	1.928375
H	0.535655	-1.002720	0.647164
H	3.366661	2.196391	1.114801
H	-8.586746	-0.589627	0.972078
H	7.150635	-2.664337	-1.263614
C	9.687502	-2.705218	-1.910736
C	5.393546	-0.899215	-0.372564
O	2.761193	-1.661563	0.072694
C	-6.009179	-0.008447	0.854377
C	-4.696604	0.309745	0.838985
O	-4.672675	0.914755	3.142206
C	4.469263	-0.023830	0.076010
C	-4.054770	0.782799	2.088519
C	3.081112	-0.494670	0.303130
C	-1.025319	3.050536	2.234676
C	-2.309916	2.575984	2.351060
C	-2.623246	1.235166	1.993095
C	-1.632276	0.411303	1.499619
C	-0.287931	0.861684	1.398104

C	0.021599	2.211932	1.766501
C	1.364197	2.656765	1.646289
C	0.750596	0.023565	0.929564
C	2.055147	0.476103	0.814740
C	2.352725	1.818955	1.185056
H	-6.985822	-1.250469	-3.468797
O	-5.181315	-0.478179	-1.995637
C	-6.324751	-0.657994	-1.609132
O	-9.412950	-1.755040	-3.154588
N	-7.304322	-1.102660	-2.517173
C	-8.163201	-0.721198	-0.018572
C	-8.642090	-1.369171	-2.294556
N	-9.044493	-1.153537	-0.968718
C	-6.838446	-0.459915	-0.244680
H	9.263429	1.571799	-0.714366
O	6.894168	1.692677	-0.092672
C	7.426818	0.666447	-0.486259
O	10.769809	-0.236656	-1.551189
N	8.792809	0.679974	-0.823586
C	7.574312	-1.675731	-1.118291
C	9.586984	-0.353196	-1.285445
N	8.902200	-1.567755	-1.424159
C	6.792610	-0.649361	-0.661283
C	-10.453109	-1.420703	-0.665433
H	-11.094987	-0.792489	-1.287195
H	-10.689206	-2.467259	-0.872025
H	-10.630159	-1.202669	0.388542

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generated by VMD

H	10.390401	-3.099792	-1.096316
H	8.899596	-3.747789	-1.837393
H	10.004927	-2.709481	-2.781852
H	4.955503	-1.939636	-0.471566
H	-6.548151	0.417755	1.712130
H	-4.059952	0.214801	-0.052852
H	4.723488	1.017548	0.227645
H	-0.726048	4.298095	2.392630
H	-3.068841	3.509984	2.599836
H	-1.935135	-0.367651	1.135399
H	1.665308	3.809306	1.856074
H	0.455826	-0.852753	0.628535
H	3.386980	2.255547	1.089087
H	-8.596328	-0.363439	0.888391
H	7.038100	-2.762999	-1.160824
C	9.575634	-2.892869	-1.794200
C	5.325962	-0.923371	-0.336070
O	2.670105	-1.598808	0.112663
C	-6.001265	0.144657	0.808966
C	-4.683262	0.438605	0.803171
O	-4.756886	1.439711	2.961585
C	4.423535	-0.009716	0.079512
C	-4.089251	1.113191	1.982067
C	3.022233	-0.435798	0.313647
C	-0.988506	3.274702	2.135432
C	-2.289281	2.845469	2.240258
C	-2.644467	1.511705	1.892893
C	-1.673875	0.646875	1.427327
C	-0.314040	1.050128	1.339023
C	0.035897	2.394533	1.692843
C	1.394079	2.791865	1.585109
C	0.701679	0.169982	0.898257
C	2.022226	0.576637	0.795002

C	2.359686	1.914139	1.149565
H	-6.792602	-1.830155	-3.272778
O	-5.044603	-0.839306	-1.864014
C	-6.210612	-0.927220	-1.514293
O	-9.245249	-2.227442	-3.007184
N	-7.154448	-1.509992	-2.380920
C	-8.127221	-0.675648	-0.039272
C	-8.508540	-1.709087	-2.187747
N	-8.970929	-1.255053	-0.944469
C	-6.786645	-0.480528	-0.236280
H	9.261307	1.431548	-0.743289
O	6.894690	1.635108	-0.132915
C	7.400763	0.583019	-0.491793
O	10.721486	-0.442944	-1.514096
N	8.767360	0.549457	-0.825160
C	7.487640	-1.781809	-1.046324
C	9.535188	-0.519204	-1.249324
N	8.818832	-1.719262	-1.350238
C	6.732193	-0.720441	-0.626500
C	-10.400108	-1.436932	-0.676568
H	-10.991771	-0.910327	-1.428935
H	-10.656716	-2.498137	-0.715871
H	-10.622484	-1.038828	0.314300

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generated by VMD

H	10.296026	-3.273384	-0.889184
H	8.798899	-3.917633	-1.620831
H	9.939881	-2.947151	-2.594663
H	4.880121	-1.956515	-0.401321
H	-6.572243	0.705391	1.562857
H	-4.035999	0.210902	-0.067866
H	4.714215	1.029347	0.186705
H	-0.659845	4.503889	2.235273
H	-3.033696	3.793423	2.395719
H	-1.990195	-0.137496	1.010675
H	1.725120	3.931404	1.752638
H	0.388647	-0.704174	0.556248
H	3.410552	2.311474	1.043175
H	-8.598448	-0.198253	0.810416
H	6.951803	-2.859340	-1.019469
C	9.494727	-3.077773	-1.605611
C	5.275024	-0.945838	-0.296608
O	2.592418	-1.533440	0.118338
C	-5.998651	0.287279	0.734999
C	-4.679260	0.573504	0.722953
O	-4.817677	1.923142	2.676742
C	4.390401	0.005106	0.070880
C	-4.116438	1.430154	1.794160
C	2.974831	-0.376220	0.295834
C	-0.950897	3.487422	1.981306
C	-2.266600	3.102476	2.060877
C	-2.659953	1.778154	1.715833
C	-1.706329	0.874628	1.287950
C	-0.332762	1.232904	1.223968
C	0.053595	2.569305	1.569875
C	1.425957	2.920274	1.487548
C	0.662462	0.313603	0.817738
C	1.997738	0.675297	0.738787
C	2.371141	2.005473	1.083852
H	-6.669982	-2.330173	-2.991471
O	-4.961944	-1.125464	-1.706112
C	-6.139632	-1.152218	-1.386105

O	-9.133073	-2.672714	-2.748151
N	-7.058536	-1.867654	-2.176654
C	-8.101434	-0.659027	-0.037339
C	-8.419846	-2.029411	-1.999474
N	-8.919198	-1.377959	-0.863072
C	-6.753965	-0.500557	-0.218852
H	9.274719	1.292138	-0.733987
O	6.906031	1.577919	-0.165613
C	7.390074	0.500502	-0.476537
O	10.697656	-0.647327	-1.407582
N	8.759728	0.420009	-0.788471
C	7.424560	-1.885888	-0.936711
C	9.506206	-0.683568	-1.157930
N	8.761292	-1.868761	-1.221461
C	6.690351	-0.789993	-0.571248
C	-10.357996	-1.508991	-0.618157
H	-10.920822	-1.095219	-1.458249
H	-10.624442	-2.562792	-0.509437
H	-10.606647	-0.967998	0.295790

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generated by VMD

H	10.149799	-3.546568	-0.584114
H	8.679520	-4.088768	-1.442006
H	9.941606	-3.161438	-2.301713
H	4.795664	-1.950843	-0.426370
H	-6.591646	1.037261	1.327094
H	-4.032973	0.189536	-0.107451
H	4.747154	1.027736	0.215610
H	-0.610822	4.802728	1.721735
H	-3.005436	4.158574	1.882017
H	-2.026703	0.115421	0.808668
H	1.766366	4.145804	1.306583
H	0.336614	-0.531235	0.420884
H	3.421781	2.442812	0.739415
H	-8.606953	-0.012486	0.753010
H	6.855743	-2.949624	-0.927189
C	9.416600	-3.289736	-1.352418
C	5.233921	-0.964213	-0.275182
O	2.510059	-1.426911	-0.000946
C	-6.003984	0.445884	0.624333
C	-4.683338	0.720864	0.574089
O	-4.855684	2.473644	2.168992
C	4.377268	0.025709	0.053653
C	-4.135275	1.793569	1.438416
C	2.932740	-0.284631	0.182126
C	-0.922562	3.778677	1.530423
C	-2.247784	3.430594	1.611191
C	-2.670045	2.095787	1.346949
C	-1.729193	1.140944	1.009539
C	-0.346990	1.463697	0.942162
C	0.066155	2.811843	1.199253
C	1.447098	3.124686	1.112887
C	0.631371	0.495226	0.617704
C	1.976607	0.818518	0.537587
C	2.376087	2.161357	0.792329
H	-6.613137	-2.928886	-2.446456
O	-4.925334	-1.472172	-1.422384
C	-6.109458	-1.427176	-1.128553
O	-9.082218	-3.208331	-2.185410
N	-7.015662	-2.298523	-1.761308
C	-8.095221	-0.648855	0.038051
C	-8.381140	-2.416667	-1.581308

N	-8.899853	-1.531586	-0.626215
C	-6.743763	-0.534367	-0.146168
H	9.362141	1.070087	-0.406338
O	6.978301	1.469238	0.010348
C	7.426809	0.373282	-0.289081
O	10.726433	-0.929210	-1.024393
N	8.808275	0.227012	-0.512427
C	7.370554	-2.003441	-0.794316
C	9.520673	-0.907488	-0.854617
N	8.722165	-2.050684	-0.992059
C	6.670355	-0.877484	-0.454540
C	-10.343966	-1.604705	-0.388755
H	-10.887495	-1.383484	-1.310390
H	-10.618310	-2.609250	-0.058727
H	-10.607999	-0.877382	0.380054

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generated by VMD

H	10.111968	-3.628240	-0.272097
H	8.633402	-4.227057	-1.076419
H	9.896517	-3.379958	-2.013877
H	4.762278	-1.980761	-0.265545
H	-6.614395	1.276445	1.027942
H	-4.023384	0.171107	-0.144528
H	4.739316	1.038252	0.149670
H	-0.569166	4.953079	1.453708
H	-2.984910	4.370742	1.507397
H	-2.052428	0.277038	0.595891
H	1.808710	4.228594	1.165044
H	0.301315	-0.432742	0.325209
H	3.444285	2.470891	0.716347
H	-8.616521	0.135373	0.597829
H	6.815819	-3.033589	-0.675833
C	9.376118	-3.428531	-1.054547
C	5.209593	-0.989981	-0.184070
O	2.474694	-1.399418	0.075279
C	-6.008958	0.566138	0.464392
C	-4.685150	0.823291	0.408209
O	-4.897739	2.836276	1.646237
C	4.360792	0.030137	0.062559
C	-4.154224	2.030489	1.085542
C	2.911664	-0.254492	0.198022
C	-0.897209	3.932897	1.269045
C	-2.232670	3.619109	1.292956
C	-2.678326	2.288933	1.039701
C	-1.745166	1.302608	0.777271
C	-0.353963	1.591698	0.762961
C	0.081590	2.935098	1.006666
C	1.472404	3.211639	0.978563
C	0.612917	0.591276	0.508413
C	1.968110	0.879868	0.481073
C	2.389908	2.218199	0.722675
H	-6.542746	-3.311509	-1.958505
O	-4.879625	-1.690302	-1.168697
C	-6.071608	-1.594344	-0.921830
O	-9.018863	-3.544039	-1.720634
N	-6.962436	-2.567455	-1.411643
C	-8.086889	-0.619847	0.025841
C	-8.332877	-2.653599	-1.251810
N	-8.875547	-1.609913	-0.489990
C	-6.730289	-0.538839	-0.136712
H	9.359680	0.988692	-0.426909
O	6.976998	1.440202	-0.059580

C	7.416380	0.321175	-0.275593
O	10.707659	-1.063328	-0.888069
N	8.797934	0.145585	-0.476089
C	7.339347	-2.085342	-0.607524
C	9.501050	-1.017641	-0.729507
N	8.691933	-2.160099	-0.789340
C	6.648238	-0.930799	-0.356728
C	-10.325789	-1.642301	-0.283092
H	-10.842222	-1.606914	-1.245176
H	-10.610725	-2.564856	0.228027
H	-10.610899	-0.780709	0.321997

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generated by VMD

H	10.099829	-3.666275	0.151464
H	8.622169	-4.330136	-0.601951
H	9.893999	-3.578910	-1.606933
H	4.752614	-1.997328	-0.076616
H	-6.632129	1.491816	0.604904
H	-4.018954	0.106771	-0.136525
H	4.744330	1.046676	0.074080
H	-0.548362	5.086164	1.002353
H	-2.972638	4.538639	0.962541
H	-2.055765	0.386886	0.341528
H	1.830219	4.316001	0.867807
H	0.289671	-0.364304	0.212715
H	3.457166	2.514020	0.590948
H	-8.637403	0.305312	0.352624
H	6.805151	-3.094577	-0.354626
C	9.368790	-3.536392	-0.649818
C	5.206132	-1.006020	-0.071745
O	2.461350	-1.373664	0.147599
C	-6.017697	0.663033	0.252694
C	-4.686525	0.881737	0.212203
O	-4.916661	3.117027	0.963160
C	4.360548	0.036744	0.068171
C	-4.160493	2.202593	0.632311
C	2.906550	-0.226374	0.194814
C	-0.882637	4.061610	0.857138
C	-2.221993	3.766711	0.832176
C	-2.677947	2.429815	0.635752
C	-1.747330	1.418493	0.477612
C	-0.352720	1.689964	0.507343
C	0.092279	3.038697	0.696887
C	1.486916	3.295050	0.719816
C	0.608891	0.664237	0.352702
C	1.968230	0.933430	0.371443
C	2.399501	2.277148	0.559857
H	-6.530095	-3.661261	-1.241229
O	-4.867966	-1.920425	-0.767598
C	-6.067253	-1.756877	-0.605554
O	-9.018591	-3.805595	-1.072264
N	-6.956911	-2.806376	-0.900908
C	-8.099965	-0.567258	-0.004881
C	-8.334606	-2.838129	-0.790295
N	-8.887073	-1.640165	-0.317779
C	-6.736008	-0.541755	-0.115431
H	9.375717	0.918275	-0.400667
O	6.990090	1.414429	-0.119015
C	7.424686	0.278357	-0.230415
O	10.715485	-1.174002	-0.661019
N	8.808337	0.077599	-0.388109
C	7.334786	-2.147266	-0.357125

C	9.506704	-1.107402	-0.528666
N	8.689941	-2.245795	-0.505100
C	6.648280	-0.971091	-0.219667
C	-10.345126	-1.607738	-0.176339
H	-10.818666	-1.809440	-1.139834
H	-10.670581	-2.370623	0.534960
H	-10.638651	-0.619758	0.180705

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generated by VMD

H	10.078094	-3.632574	0.757989
H	8.619473	-4.377295	0.043886
H	9.924972	-3.752690	-1.003781
H	4.750357	-1.988921	0.173791
H	-6.639862	1.606674	-0.002257
H	-4.021380	0.047128	0.035001
H	4.754152	1.051394	-0.040164
H	-0.546145	5.178929	0.252440
H	-2.969912	4.632639	0.182218
H	-2.050222	0.435771	0.087874
H	1.832723	4.395448	0.288835
H	0.290072	-0.326995	0.125675
H	3.458662	2.571073	0.271950
H	-8.651558	0.407133	-0.097806
H	6.803217	-3.116264	0.090908
C	9.371827	-3.596488	-0.074615
C	5.209200	-1.005084	0.073978
O	2.457473	-1.339983	0.246020
C	-6.024120	0.707083	-0.007361
C	-4.688649	0.896853	0.037318
O	-4.917444	3.252590	0.095031
C	4.365507	0.048510	0.062444
C	-4.160075	2.280899	0.087913
C	2.906948	-0.195672	0.174108
C	-0.880141	4.144642	0.216413
C	-2.219118	3.850084	0.177890
C	-2.676217	2.499766	0.129954
C	-1.745052	1.476403	0.123361
C	-0.350760	1.748456	0.162459
C	0.094724	3.109268	0.209474
C	1.489164	3.364732	0.247953
C	0.610055	0.710401	0.156138
C	1.969494	0.978093	0.186865
C	2.401256	2.333906	0.234368
H	-6.540838	-3.865622	-0.143799
O	-4.872833	-2.069984	-0.044032
C	-6.075413	-1.859741	-0.076084
O	-9.036722	-3.941381	-0.226799
N	-6.968258	-2.945848	-0.132608
C	-8.112989	-0.535151	-0.105378
C	-8.350012	-2.936744	-0.177195
N	-8.903070	-1.649406	-0.159804
C	-6.745131	-0.550023	-0.062442
H	9.398046	0.856095	-0.357049
O	7.008125	1.388149	-0.207640
C	7.439085	0.245895	-0.169632
O	10.732187	-1.255606	-0.327216
N	8.825534	0.024371	-0.260768
C	7.338420	-2.177341	-0.008767
C	9.520540	-1.170693	-0.238873
N	8.696675	-2.296169	-0.103682
C	6.655255	-0.991435	-0.033193
C	-10.365466	-1.569001	-0.205643

H	-10.738201	-2.031725	-1.122516
H	-10.796278	-2.097685	0.647751
H	-10.657677	-0.518532	-0.176098

8.8. *Ab initio* study on TACN and TACNA

8.8.1. 6-31G* optimized structure of Zn²⁺ in vacuum

```
HETATM    1 Zn          0          0.000   0.000   0.000          Zn
END
```

8.8.2. 6-31G* optimized structure of TACN in vacuum

```
HETATM    1 C          0          0.000   2.105  -0.040          C
HETATM    2 N          0          1.172   1.348  -0.472          N
HETATM    3 C          0          1.755   0.402   0.483          C
HETATM    4 C          0          1.823  -1.052  -0.040          C
HETATM    5 H          0          1.193   0.426   1.425          H
HETATM    6 H          0          2.782   0.708   0.737          H
HETATM    7 N          0          0.582  -1.689  -0.472          N
HETATM    8 H          0          2.281  -1.671   0.743          H
HETATM    9 H          0          2.512  -1.085  -0.895          H
HETATM   10 C          0         -0.529  -1.721   0.483          C
HETATM   11 C          0         -1.823  -1.052  -0.040          C
HETATM   12 H          0         -0.227  -1.246   1.425          H
HETATM   13 H          0         -0.778  -2.763   0.737          H
HETATM   14 N          0         -1.754   0.341  -0.472          N
HETATM   15 H          0         -2.588  -1.140   0.743          H
HETATM   16 H          0         -2.196  -1.633  -0.895          H
HETATM   17 C          0         -1.225   1.319   0.483          C
HETATM   18 H          0         -2.004   2.055   0.737          H
HETATM   19 H          0         -0.965   0.820   1.425          H
HETATM   20 H          0         -0.316   2.718  -0.895          H
HETATM   21 H          0          0.307   2.811   0.743          H
HETATM   22 H          0         -1.264   0.410  -1.360          H
HETATM   23 H          0          0.277  -1.300  -1.360          H
HETATM   24 H          0          0.988   0.890  -1.360          H
END
```

8.8.3. 6-31G* optimized structure of [Zn(TACNA)]²⁺ complex in vacuum

```
HETATM    1 C          0          1.413   1.649  -0.519          C
HETATM    2 N          0          0.000   1.700   0.048          N
HETATM    3 C          0         -1.107   1.460  -0.972          C
HETATM    4 C          0         -2.134   0.399  -0.519          C
HETATM    5 H          0         -0.637   1.172  -1.915          H
HETATM    6 H          0         -1.639   2.398  -1.159          H
HETATM    7 N          0         -1.472  -0.850   0.048          N
HETATM    8 H          0         -2.770   0.135  -1.371          H
```

HETATM	9	H	0	-2.784	0.796	0.268	H
HETATM	10	C	0	-0.711	-1.689	-0.972	C
HETATM	11	C	0	0.722	-2.048	-0.519	C
HETATM	12	H	0	-0.696	-1.138	-1.915	H
HETATM	13	H	0	-1.257	-2.618	-1.159	H
HETATM	14	N	0	1.472	-0.850	0.048	N
HETATM	15	H	0	1.268	-2.467	-1.371	H
HETATM	16	H	0	0.702	-2.809	0.268	H
HETATM	17	C	0	1.818	0.229	-0.972	C
HETATM	18	H	0	2.896	0.220	-1.159	H
HETATM	19	H	0	1.334	-0.034	-1.915	H
HETATM	20	H	0	2.081	2.013	0.268	H
HETATM	21	H	0	1.502	2.332	-1.371	H
HETATM	22	H	0	2.323	-1.183	0.507	H
HETATM	23	H	0	-2.186	-1.421	0.507	H
HETATM	24	H	0	-0.137	2.603	0.507	H
HETATM	25	Zn	0	0.000	0.000	1.228	Zn
END							

8.8.4. 6-31G* optimized structure of TACNA in vacuum

HETATM	1	C	0	-2.097	2.057	-0.290	C
HETATM	2	N	0	-0.938	1.705	0.548	N
HETATM	3	C	0	-1.238	0.939	1.772	C
HETATM	4	C	0	-0.396	-0.351	1.883	C
HETATM	5	H	0	-2.304	0.687	1.794	H
HETATM	6	H	0	-1.057	1.541	2.674	H
HETATM	7	N	0	-0.404	-1.139	0.659	N
HETATM	8	H	0	-0.741	-0.921	2.768	H
HETATM	9	H	0	0.643	-0.068	2.081	H
HETATM	10	C	0	-1.665	-1.781	0.294	C
HETATM	11	C	0	-2.051	-1.489	-1.180	C
HETATM	12	H	0	-2.449	-1.403	0.959	H
HETATM	13	H	0	-1.636	-2.872	0.457	H
HETATM	14	N	0	-1.960	-0.088	-1.584	N
HETATM	15	H	0	-3.067	-1.860	-1.366	H
HETATM	16	H	0	-1.388	-2.058	-1.844	H
HETATM	17	C	0	-2.837	0.839	-0.864	C
HETATM	18	H	0	-3.617	1.227	-1.537	H
HETATM	19	H	0	-3.379	0.331	-0.053	H
HETATM	20	H	0	-1.710	2.661	-1.120	H
HETATM	21	H	0	-2.838	2.680	0.243	H
HETATM	22	H	0	-0.998	0.197	-1.423	H
HETATM	23	H	0	-0.448	2.561	0.791	H
HETATM	24	C	0	0.806	-1.897	0.399	C
HETATM	25	C	0	1.989	-1.073	-0.144	C
HETATM	26	H	0	0.594	-2.653	-0.365	H
HETATM	27	H	0	1.181	-2.449	1.277	H
HETATM	28	O	0	3.098	-1.588	-0.257	O
HETATM	29	N	0	1.699	0.204	-0.513	N
HETATM	30	C	0	2.729	1.097	-1.026	C
HETATM	31	H	0	0.777	0.578	-0.283	H
HETATM	32	C	0	3.463	1.873	0.072	C
HETATM	33	H	0	2.255	1.788	-1.733	H
HETATM	34	H	0	3.441	0.482	-1.583	H
HETATM	35	H	0	2.768	2.489	0.656	H
HETATM	36	H	0	4.221	2.536	-0.362	H
HETATM	37	H	0	3.964	1.178	0.754	H
END							

8.8.5. 6-31G* optimized structure of [Zn(TACNA)]²⁺ complex in vacuum

HETATM	1	C	0	3.139	-1.442	0.045	C
HETATM	2	N	0	1.828	-1.481	0.777	N
HETATM	3	C	0	1.762	-0.601	2.000	C
HETATM	4	C	0	0.502	0.288	2.037	C
HETATM	5	H	0	2.669	0.007	2.035	H
HETATM	6	H	0	1.773	-1.221	2.901	H
HETATM	7	N	0	0.250	0.989	0.740	N
HETATM	8	H	0	0.594	1.003	2.866	H
HETATM	9	H	0	-0.376	-0.335	2.241	H
HETATM	10	C	0	1.283	2.008	0.378	C
HETATM	11	C	0	1.795	1.839	-1.067	C
HETATM	12	H	0	2.110	1.921	1.086	H
HETATM	13	H	0	0.894	3.025	0.497	H
HETATM	14	N	0	2.157	0.413	-1.387	N
HETATM	15	H	0	2.657	2.499	-1.220	H
HETATM	16	H	0	1.025	2.135	-1.787	H
HETATM	17	C	0	3.376	-0.093	-0.659	C
HETATM	18	H	0	4.205	-0.210	-1.362	H
HETATM	19	H	0	3.687	0.668	0.061	H
HETATM	20	H	0	3.113	-2.256	-0.688	H
HETATM	21	H	0	3.974	-1.640	0.727	H
HETATM	22	H	0	2.324	0.357	-2.392	H
HETATM	23	H	0	1.656	-2.448	1.055	H
HETATM	24	C	0	-1.151	1.438	0.584	C
HETATM	25	C	0	-2.020	0.352	-0.087	C
HETATM	26	H	0	-1.170	2.312	-0.077	H
HETATM	27	H	0	-1.587	1.749	1.541	H
HETATM	28	O	0	-1.472	-0.576	-0.769	O
HETATM	29	N	0	-3.326	0.449	0.024	N
HETATM	30	C	0	-4.319	-0.443	-0.632	C
HETATM	31	H	0	-3.704	1.216	0.573	H
HETATM	32	C	0	-5.553	-0.614	0.247	C
HETATM	33	H	0	-3.818	-1.393	-0.822	H
HETATM	34	H	0	-4.578	0.003	-1.599	H
HETATM	35	H	0	-5.306	-1.083	1.205	H
HETATM	36	H	0	-6.271	-1.258	-0.269	H
HETATM	37	H	0	-6.053	0.342	0.437	H
HETATM	38	Zn	0	0.490	-0.659	-0.634	Zn
END							

8.8.6. 6-31G* optimized structure of [Zn(H₂O)₆]²⁺ complex in vacuum

HETATM	1	Zn	0	0.000	0.000	0.000	Zn
HETATM	2	O	0	0.000	1.740	1.231	O
HETATM	3	H	0	0.554	2.530	1.112	H
HETATM	4	H	0	-0.552	1.893	2.017	H
HETATM	5	O	0	1.507	-0.870	1.231	O
HETATM	6	H	0	1.916	-0.468	2.017	H
HETATM	7	H	0	1.915	-1.745	1.112	H
HETATM	8	O	0	-1.507	-0.870	1.231	O
HETATM	9	H	0	-1.364	-1.425	2.017	H
HETATM	10	H	0	-2.468	-0.786	1.112	H
HETATM	11	O	0	1.507	0.870	-1.231	O

HETATM	12	H	0	2.468	0.786	-1.112	H
HETATM	13	H	0	1.364	1.425	-2.017	H
HETATM	14	O	0	-1.507	0.870	-1.231	O
HETATM	15	H	0	-1.916	0.468	-2.017	H
HETATM	16	H	0	-1.915	1.745	-1.112	H
HETATM	17	O	0	0.000	-1.740	-1.231	O
HETATM	18	H	0	0.552	-1.893	-2.017	H
HETATM	19	H	0	-0.554	-2.530	-1.112	H
END							

8.8.7. 6-31G* optimized structure of water in vacuum

HETATM	1	O	0	0.000	0.000	0.120	O
HETATM	2	H	0	0.000	0.761	-0.479	H
HETATM	3	H	0	0.000	-0.761	-0.479	H
END							

8.8.8. 6-31G* optimized structure of [Zn(TACN)(H₂O)]²⁺ complex in vacuum

HETATM	1	C	0	-0.823	0.085	2.156	C
HETATM	2	N	0	-0.299	-1.044	1.312	N
HETATM	3	C	0	-1.343	-1.732	0.469	C
HETATM	4	C	0	-0.918	-1.878	-1.003	C
HETATM	5	H	0	-2.275	-1.168	0.549	H
HETATM	6	H	0	-1.549	-2.728	0.872	H
HETATM	7	N	0	-0.330	-0.610	-1.555	N
HETATM	8	H	0	-1.784	-2.196	-1.595	H
HETATM	9	H	0	-0.150	-2.653	-1.105	H
HETATM	10	C	0	-1.321	0.512	-1.733	C
HETATM	11	C	0	-0.835	1.841	-1.126	C
HETATM	12	H	0	-2.270	0.206	-1.285	H
HETATM	13	H	0	-1.518	0.668	-2.797	H
HETATM	14	N	0	-0.252	1.663	0.250	N
HETATM	15	H	0	-1.669	2.551	-1.106	H
HETATM	16	H	0	-0.046	2.278	-1.748	H
HETATM	17	C	0	-1.260	1.295	1.311	C
HETATM	18	H	0	-1.420	2.145	1.980	H
HETATM	19	H	0	-2.217	1.102	0.822	H
HETATM	20	H	0	-0.015	0.367	2.840	H
HETATM	21	H	0	-1.670	-0.247	2.768	H
HETATM	22	H	0	0.191	2.544	0.512	H
HETATM	23	H	0	0.086	-0.829	-2.461	H
HETATM	24	H	0	0.122	-1.729	1.941	H
HETATM	25	Zn	0	0.994	-0.024	-0.013	Zn
HETATM	26	O	0	3.037	-0.039	-0.027	O
HETATM	27	H	0	3.623	0.739	0.021	H
HETATM	28	H	0	3.613	-0.825	-0.086	H
END							

8.8.9. 6-31G* optimized structure of [Zn(TACNA)(H₂O)]²⁺ complex in vacuum

HETATM	1	C	0	-3.236	0.759	0.848	C
HETATM	2	N	0	-1.931	0.407	1.491	N
HETATM	3	C	0	-1.807	-1.032	1.900	C

HETATM	4	C	0	-0.494	-1.673	1.418	C
HETATM	5	H	0	-2.671	-1.580	1.516	H
HETATM	6	H	0	-1.852	-1.117	2.990	H
HETATM	7	N	0	-0.208	-1.397	-0.018	N
HETATM	8	H	0	-0.527	-2.753	1.618	H
HETATM	9	H	0	0.341	-1.262	1.997	H
HETATM	10	C	0	-1.172	-2.028	-0.965	C
HETATM	11	C	0	-1.695	-1.028	-2.010	C
HETATM	12	H	0	-2.001	-2.444	-0.389	H
HETATM	13	H	0	-0.717	-2.878	-1.485	H
HETATM	14	N	0	-2.147	0.256	-1.385	N
HETATM	15	H	0	-2.508	-1.496	-2.579	H
HETATM	16	H	0	-0.904	-0.770	-2.724	H
HETATM	17	C	0	-3.389	0.128	-0.548	C
HETATM	18	H	0	-4.233	0.605	-1.054	H
HETATM	19	H	0	-3.643	-0.931	-0.463	H
HETATM	20	H	0	-3.267	1.852	0.777	H
HETATM	21	H	0	-4.083	0.451	1.473	H
HETATM	22	H	0	-2.329	0.914	-2.142	H
HETATM	23	H	0	-1.827	0.994	2.319	H
HETATM	24	C	0	1.207	-1.635	-0.358	C
HETATM	25	C	0	2.060	-0.403	-0.018	C
HETATM	26	H	0	1.288	-1.788	-1.441	H
HETATM	27	H	0	1.601	-2.538	0.126	H
HETATM	28	O	0	1.514	0.739	0.022	O
HETATM	29	N	0	3.354	-0.580	0.168	N
HETATM	30	C	0	4.350	0.492	0.404	C
HETATM	31	H	0	3.713	-1.530	0.140	H
HETATM	32	C	0	5.677	0.158	-0.272	C
HETATM	33	H	0	4.475	0.604	1.486	H
HETATM	34	H	0	3.921	1.417	0.016	H
HETATM	35	H	0	6.105	-0.775	0.115	H
HETATM	36	H	0	6.397	0.955	-0.064	H
HETATM	37	H	0	5.568	0.074	-1.357	H
HETATM	38	Zn	0	-0.520	0.825	-0.076	Zn
HETATM	39	O	0	-0.221	2.902	-0.283	O
HETATM	40	H	0	0.701	3.209	-0.237	H
HETATM	41	H	0	-0.781	3.677	-0.452	H

END

8.8.10.6-31G* optimized structure of Zn²⁺ in PCM water

HETATM	1	Zn	0	0.000	0.000	0.000	Zn
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END

8.8.11.6-31G* optimized structure of TACN in PCM water

HETATM	1	C	0	0.000	2.106	-0.040	C
HETATM	2	N	0	1.178	1.353	-0.474	N
HETATM	3	C	0	1.751	0.399	0.484	C
HETATM	4	C	0	1.824	-1.053	-0.040	C
HETATM	5	H	0	1.185	0.423	1.423	H
HETATM	6	H	0	2.778	0.698	0.747	H
HETATM	7	N	0	0.583	-1.697	-0.474	N
HETATM	8	H	0	2.285	-1.667	0.744	H
HETATM	9	H	0	2.509	-1.081	-0.898	H

HETATM	10	C	0	-0.530	-1.716	0.484	C
HETATM	11	C	0	-1.824	-1.053	-0.040	C
HETATM	12	H	0	-0.226	-1.238	1.423	H
HETATM	13	H	0	-0.785	-2.755	0.747	H
HETATM	14	N	0	-1.761	0.343	-0.474	N
HETATM	15	H	0	-2.586	-1.146	0.744	H
HETATM	16	H	0	-2.191	-1.633	-0.898	H
HETATM	17	C	0	-1.221	1.317	0.484	C
HETATM	18	H	0	-1.994	2.057	0.747	H
HETATM	19	H	0	-0.959	0.815	1.423	H
HETATM	20	H	0	-0.318	2.714	-0.898	H
HETATM	21	H	0	0.301	2.813	0.744	H
HETATM	22	H	0	-1.262	0.409	-1.359	H
HETATM	23	H	0	0.276	-1.297	-1.359	H
HETATM	24	H	0	0.985	0.888	-1.359	H
END							

8.8.12.6-31G* optimized structure of [Zn(TACN)]²⁺ complex in PCM water

HETATM	1	C	0	-0.490	-2.079	0.535	C
HETATM	2	N	0	-1.333	-0.982	-0.010	N
HETATM	3	C	0	-1.802	0.019	0.991	C
HETATM	4	C	0	-1.557	1.467	0.530	C
HETATM	5	H	0	-1.292	-0.172	1.938	H
HETATM	6	H	0	-2.871	-0.110	1.181	H
HETATM	7	N	0	-0.186	1.647	-0.019	N
HETATM	8	H	0	-1.742	2.144	1.373	H
HETATM	9	H	0	-2.254	1.727	-0.272	H
HETATM	10	C	0	0.911	1.554	0.989	C
HETATM	11	C	0	2.045	0.617	0.537	C
HETATM	12	H	0	0.488	1.209	1.935	H
HETATM	13	H	0	1.333	2.545	1.181	H
HETATM	14	N	0	1.518	-0.662	-0.014	N
HETATM	15	H	0	2.720	0.439	1.383	H
HETATM	16	H	0	2.622	1.089	-0.264	H
HETATM	17	C	0	0.891	-1.573	0.987	C
HETATM	18	H	0	1.537	-2.438	1.163	H
HETATM	19	H	0	0.812	-1.045	1.940	H
HETATM	20	H	0	-0.373	-2.810	-0.271	H
HETATM	21	H	0	-0.981	-2.579	1.378	H
HETATM	22	H	0	2.278	-1.147	-0.486	H
HETATM	23	H	0	-0.140	2.547	-0.491	H
HETATM	24	H	0	-2.136	-1.394	-0.480	H
HETATM	25	Zn	0	0.002	-0.013	-1.414	Zn
END							

8.8.13.6-31G* optimized structure of TACNA in PCM water

HETATM	1	C	0	-2.016	2.078	-0.314	C
HETATM	2	N	0	-0.873	1.700	0.536	N
HETATM	3	C	0	-1.213	0.965	1.770	C
HETATM	4	C	0	-0.417	-0.351	1.905	C
HETATM	5	H	0	-2.287	0.749	1.786	H
HETATM	6	H	0	-1.017	1.576	2.661	H
HETATM	7	N	0	-0.429	-1.143	0.681	N
HETATM	8	H	0	-0.799	-0.908	2.781	H

HETATM	9	H	0	0.627	-0.102	2.124	H
HETATM	10	C	0	-1.698	-1.761	0.304	C
HETATM	11	C	0	-2.059	-1.476	-1.176	C
HETATM	12	H	0	-2.479	-1.359	0.957	H
HETATM	13	H	0	-1.694	-2.851	0.474	H
HETATM	14	N	0	-1.923	-0.079	-1.598	N
HETATM	15	H	0	-3.085	-1.819	-1.366	H
HETATM	16	H	0	-1.406	-2.074	-1.825	H
HETATM	17	C	0	-2.785	0.876	-0.884	C
HETATM	18	H	0	-3.552	1.278	-1.564	H
HETATM	19	H	0	-3.342	0.384	-0.075	H
HETATM	20	H	0	-1.608	2.669	-1.143	H
HETATM	21	H	0	-2.743	2.721	0.213	H
HETATM	22	H	0	-0.959	0.179	-1.394	H
HETATM	23	H	0	-0.367	2.549	0.776	H
HETATM	24	C	0	0.772	-1.907	0.416	C
HETATM	25	C	0	1.948	-1.096	-0.157	C
HETATM	26	H	0	0.549	-2.679	-0.330	H
HETATM	27	H	0	1.159	-2.443	1.300	H
HETATM	28	O	0	3.047	-1.640	-0.321	O
HETATM	29	N	0	1.681	0.188	-0.482	N
HETATM	30	C	0	2.701	1.087	-1.008	C
HETATM	31	H	0	0.765	0.577	-0.228	H
HETATM	32	C	0	3.455	1.850	0.087	C
HETATM	33	H	0	2.209	1.790	-1.688	H
HETATM	34	H	0	3.400	0.486	-1.596	H
HETATM	35	H	0	2.766	2.454	0.689	H
HETATM	36	H	0	4.198	2.522	-0.358	H
HETATM	37	H	0	3.975	1.154	0.753	H
END							

8.8.14.6-31G* optimized structure of [Zn(TACNA)]²⁺ complex in PCM water

HETATM	1	C	0	3.231	-1.288	0.323	C
HETATM	2	N	0	1.919	-1.257	1.023	N
HETATM	3	C	0	1.787	-0.205	2.074	C
HETATM	4	C	0	0.484	0.604	1.947	C
HETATM	5	H	0	2.655	0.455	2.019	H
HETATM	6	H	0	1.810	-0.663	3.067	H
HETATM	7	N	0	0.228	1.058	0.555	N
HETATM	8	H	0	0.518	1.453	2.644	H
HETATM	9	H	0	-0.361	-0.028	2.238	H
HETATM	10	C	0	1.224	2.039	0.044	C
HETATM	11	C	0	1.757	1.657	-1.347	C
HETATM	12	H	0	2.047	2.102	0.758	H
HETATM	13	H	0	0.794	3.045	-0.006	H
HETATM	14	N	0	2.201	0.237	-1.398	N
HETATM	15	H	0	2.576	2.338	-1.611	H
HETATM	16	H	0	0.972	1.769	-2.100	H
HETATM	17	C	0	3.421	-0.067	-0.593	C
HETATM	18	H	0	4.270	-0.257	-1.255	H
HETATM	19	H	0	3.680	0.815	-0.003	H
HETATM	20	H	0	3.246	-2.206	-0.272	H
HETATM	21	H	0	4.065	-1.334	1.035	H
HETATM	22	H	0	2.368	-0.011	-2.371	H
HETATM	23	H	0	1.754	-2.172	1.438	H
HETATM	24	C	0	-1.166	1.471	0.343	C
HETATM	25	C	0	-2.047	0.297	-0.103	C
HETATM	26	H	0	-1.200	2.204	-0.470	H

HETATM	27	H	0	-1.593	1.955	1.230	H
HETATM	28	O	0	-1.542	-0.754	-0.579	O
HETATM	29	N	0	-3.360	0.461	-0.014	N
HETATM	30	C	0	-4.351	-0.501	-0.520	C
HETATM	31	H	0	-3.708	1.341	0.350	H
HETATM	32	C	0	-5.617	-0.484	0.331	C
HETATM	33	H	0	-3.880	-1.485	-0.508	H
HETATM	34	H	0	-4.581	-0.253	-1.563	H
HETATM	35	H	0	-5.401	-0.766	1.367	H
HETATM	36	H	0	-6.339	-1.197	-0.078	H
HETATM	37	H	0	-6.087	0.506	0.330	H
HETATM	38	Zn	0	0.516	-0.874	-0.592	Zn
END							

8.8.15.6-31G* optimized structure of $[\text{Zn}(\text{H}_2\text{O})_6]^{2+}$ complex in PCM water

HETATM	1	Zn	0	0.000	0.000	0.000	Zn
HETATM	2	O	0	0.671	1.718	1.077	O
HETATM	3	H	0	1.613	1.901	0.911	H
HETATM	4	H	0	0.546	1.788	2.039	H
HETATM	5	O	0	1.152	-1.432	1.085	O
HETATM	6	H	0	1.274	-1.353	2.046	H
HETATM	7	H	0	0.846	-2.342	0.924	H
HETATM	8	O	0	-1.822	-0.295	1.079	O
HETATM	9	H	0	-1.814	-0.443	2.040	H
HETATM	10	H	0	-2.462	0.420	0.920	H
HETATM	11	O	0	1.822	0.295	-1.079	O
HETATM	12	H	0	2.462	-0.420	-0.920	H
HETATM	13	H	0	1.814	0.443	-2.040	H
HETATM	14	O	0	-1.152	1.432	-1.085	O
HETATM	15	H	0	-1.274	1.353	-2.046	H
HETATM	16	H	0	-0.846	2.342	-0.924	H
HETATM	17	O	0	-0.671	-1.718	-1.077	O
HETATM	18	H	0	-0.546	-1.788	-2.039	H
HETATM	19	H	0	-1.613	-1.901	-0.911	H
END							

8.8.16.6-31G* optimized structure of water in PCM water

HETATM	1	O	0	0.000	0.000	0.121	O
HETATM	2	H	0	0.000	0.759	-0.483	H
HETATM	3	H	0	0.000	-0.759	-0.483	H
END							

8.8.17.6-31G* optimized structure of $[\text{Zn}(\text{TACN})(\text{H}_2\text{O})]^{2+}$ complex in PCM water

HETATM	1	C	0	0.028	0.031	0.028	C
HETATM	2	N	0	0.060	0.063	1.514	N
HETATM	3	C	0	1.425	0.020	2.113	C
HETATM	4	C	0	1.632	1.109	3.180	C
HETATM	5	H	0	2.162	0.128	1.314	H
HETATM	6	H	0	1.606	-0.956	2.573	H
HETATM	7	N	0	1.133	2.434	2.724	N
HETATM	8	H	0	2.695	1.153	3.446	H

HETATM	9	H	0	1.069	0.857	4.083	H
HETATM	10	C	0	1.926	3.065	1.630	C
HETATM	11	C	0	1.040	3.561	0.475	C
HETATM	12	H	0	2.656	2.339	1.265	H
HETATM	13	H	0	2.497	3.913	2.019	H
HETATM	14	N	0	0.025	2.547	0.082	N
HETATM	15	H	0	1.680	3.838	-0.371	H
HETATM	16	H	0	0.487	4.452	0.786	H
HETATM	17	C	0	0.579	1.331	-0.581	C
HETATM	18	H	0	0.347	1.343	-1.649	H
HETATM	19	H	0	1.668	1.355	-0.500	H
HETATM	20	H	0	-1.019	-0.101	-0.262	H
HETATM	21	H	0	0.599	-0.818	-0.367	H
HETATM	22	H	0	-0.653	2.990	-0.534	H
HETATM	23	H	0	1.103	3.061	3.526	H
HETATM	24	H	0	-0.491	-0.716	1.867	H
HETATM	25	Zn	0	-0.872	1.996	2.006	Zn
HETATM	26	O	0	-2.864	2.247	2.683	O
HETATM	27	H	0	-3.485	2.437	1.957	H
HETATM	28	H	0	-3.212	1.452	3.124	H

END

8.8.18.6-31G* optimized structure of [Zn(TACNA)(H₂O)]²⁺ complex in PCM water

HETATM	1	C	0	3.212	0.594	-1.110	C
HETATM	2	N	0	1.891	0.108	-1.591	N
HETATM	3	C	0	1.772	-1.378	-1.683	C
HETATM	4	C	0	0.484	-1.908	-1.033	C
HETATM	5	H	0	2.651	-1.829	-1.218	H
HETATM	6	H	0	1.783	-1.691	-2.730	H
HETATM	7	N	0	0.241	-1.327	0.313	N
HETATM	8	H	0	0.526	-3.005	-0.990	H
HETATM	9	H	0	-0.371	-1.638	-1.661	H
HETATM	10	C	0	1.248	-1.735	1.331	C
HETATM	11	C	0	1.807	-0.534	2.109	C
HETATM	12	H	0	2.057	-2.265	0.824	H
HETATM	13	H	0	0.819	-2.449	2.041	H
HETATM	14	N	0	2.230	0.568	1.202	N
HETATM	15	H	0	2.638	-0.876	2.739	H
HETATM	16	H	0	1.038	-0.121	2.768	H
HETATM	17	C	0	3.434	0.267	0.375	C
HETATM	18	H	0	4.293	0.840	0.735	H
HETATM	19	H	0	3.691	-0.788	0.500	H
HETATM	20	H	0	3.219	1.678	-1.258	H
HETATM	21	H	0	4.034	0.168	-1.699	H
HETATM	22	H	0	2.416	1.389	1.774	H
HETATM	23	H	0	1.714	0.516	-2.506	H
HETATM	24	C	0	-1.147	-1.532	0.751	C
HETATM	25	C	0	-2.068	-0.503	0.093	C
HETATM	26	H	0	-1.205	-1.358	1.830	H
HETATM	27	H	0	-1.494	-2.556	0.562	H
HETATM	28	O	0	-1.629	0.643	-0.185	O
HETATM	29	N	0	-3.326	-0.862	-0.134	N
HETATM	30	C	0	-4.354	0.016	-0.709	C
HETATM	31	H	0	-3.605	-1.802	0.123	H
HETATM	32	C	0	-5.357	0.508	0.335	C
HETATM	33	H	0	-4.860	-0.551	-1.495	H
HETATM	34	H	0	-3.832	0.852	-1.177	H
HETATM	35	H	0	-5.874	-0.329	0.814	H

HETATM	36	H	0	-6.109	1.139	-0.151	H
HETATM	37	H	0	-4.858	1.099	1.110	H
HETATM	38	Zn	0	0.488	0.890	-0.119	Zn
HETATM	39	O	0	-0.114	2.910	0.194	O
HETATM	40	H	0	-1.075	2.890	0.345	H
HETATM	41	H	0	0.269	3.394	0.945	H
END							

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