

University of Parma Research Repository

Pepsinogen II in gastritis and Helicobacter pylori infection

This is the peer reviewd version of the followng article:

Original

Pepsinogen II in gastritis and Helicobacter pylori infection / Di Mario, F.; Crafa, P.; Barchi, A.; Franzoni, L.; Franceschi, M.; Russo, M.; Bricca, L.; Brozzi, L.; Rodriguez Castro, K.; Rugge, M. - In: HELICOBACTER. -ISSN 1083-4389. - (2022), p. e12872. [10.1111/hel.12872]

Availability: This version is available at: 11381/2915248 since: 2022-02-03T09:02:28Z

Publisher: John Wiley and Sons Inc

Published DOI:10.1111/hel.12872

Terms of use:

Anyone can freely access the full text of works made available as "Open Access". Works made available

Publisher copyright

note finali coverpage

(Article begins on next page)

PEPSINOGEN II in GASTRITIS and *HELICOBACTER PYLORI* INFECTION

Francesco DI MARIO¹, Pellegrino CRAFA, Alberto BARCHI, Lorella FRANZONI, Marilisa FRANCESCHI, Antonio ANTICO, Piera PANOZZO, Michele RUSSO, Ludovica BRICCA⁴, L BROZZI, Kryzia RODRIGUEZ CASTRO, Massimo RUGGE^{4,5}

1 Department of Medicine and Surgery, University of Parma; Parma - Italy

4 Department of Medicine – DIMED; Surgical Pathology and Cytopathology Unit, University of Padova; Padova - Italy 5. Registro Tumori del Veneto (RTV); Azienda Zero; Padova - Italy

AUTHORS' CONTRIBUTIONS:

Crafa P, Di Mario F., Rugge M.: study design, writing of the manuscript, supervision; Brozzi L, Franceschi M.: clinical data collection and analysis; Antico A., Panozzo M.P.: laboratory data collection and analysis; Barchi A., Bricca L., Franzoni L., Rodriguez Castro K, Russo M.: data collection and analysis of the literature.

All authors critically revised the manuscript, approved the final version, and agree to be accountable for all aspects of the work. None of the authors have any conflicts of interest to disclose.

RUNNING TITLE: Pepsinogen II in non-neoplastic gastric diseases

KEYWORDS: Pepsinogen II, Helicobacter pylori, Gastritis, Dyspepsia

WORD COUNT (excluding abstract): 2533 including tables

FIGURES: 1

TABLES: 4

Corresponding author:

Francesco Di Mario, MD, Department of Medicine and Surgery, University of Parma, Via Gramsci, n. 14, 43126, Parma, Italy, Tel.: +39 3318133870, e-mail: <u>francesco.dimario@unipr.it</u>

Background and Aim: In the gastric mucosa, pepsinogen II (PgII) is produced/secreted by glands in the mucus-secreting antral and cardia compartments, but also by the chief cells and the oxyntic glands. Increasing PgII serum levels are associated with the whole spectrum of gastric inflammatory diseases, including gastritis induced by *Helicobacter pylori* (*H. pylori*). This review critically addresses the clinical value of PgII serology for assessing gastric mucosal inflammation, and as a marker of *H. pylori* status, in both naïve-*H. pylori*-positive patients and after eradication therapy.

Results: A search in PubMed/Scopus records yielded 39 out of 1,190 published scientific studies meeting the selection criteria for this study. In the studies considered, PgII levels were significantly associated with non-atrophic gastric inflammatory lesions (p-values: 0.025-0.0001). *H. pylori*-positive patients had significantly higher PgII levels than *H. pylori*-negative individuals (p-values: 0.05-0.0001). While a significant drop in serum PgII levels is consistently reported in *H. pylori*-eradicated patients (p-values: from 0.05 to 0.0001), inconsistencies in the related negative and positive predictive values significantly lower the clinical reliability of PgII testing by comparison with other available non-invasive tests.

Conclusions: PgII serology may provide clinically useful information on gastric inflammatory diseases, particularly if they are non-atrophic. PgII serology is inconsistent, however, for the purposes of distinguishing patients whose *H. pylori* eradication therapy is successful from those who remain infected.

INTRODUCTION

Acid secretion is the distinctive function of the gastric mucosa¹. Hydrochloric acid is secreted by parietal cells (also known as oxyntic cells) that are located mainly (95%) in the oxyntic mucosa (fundus and body regions), and partly in the mucus-secreting antral mucosa $(50\%)^{2,3}$

Pepsinogens are aspartic proteinases synthesized/secreted by the gastric chief cells. They have a major role in the digestive process⁴. Pepsinogen I ([PgI]; pepsinogen A) is only produced/secreted by the chief cells of the oxyntic glands. As both parietal cells and chief cells are located within the oxyntic glands, PgI is considered a reliable proxy of the stomach's acid-secreting capacity, and ultimately as a consistent surrogate marker of maximal acid output. Pepsinogen II (also known as progastricsin [PGC], or pepsinogen C) is produced/secreted by glands in the mucus-secreting antral and cardia compartments, but also by the chief cells mainly included in the oxyntic glands. ^{5, 6} Both pepsinogens are autocatalytically activated when the acidity level drops below pH 5, leading to the exposure of active site, and they are rapidly inactivated by the post-pyloric alkaline pH. While both pepsinogens are excreted mainly into the stomach lumen, very small amounts (about 1%) spread into the bloodstream, enabling their serological detection. A large body of literature has examined the reliability of serological assessments of PgI and PgII as a proxy of the gastric gland function, and on the clinical value of PgI, PgII and the PgI/II ratio as markers of gastric mucosa inflammation and atrophy (the latter consistently believed to be associated with gastric cancer risk).

This comprehensive review focuses on the clinical value of PgII as a serological marker of gastric inflammatory disease, and *H. pylori* gastritis, in particular. The following issues are critically addressed: (a) PgII serology in the setting of gastritis (be it atrophic or non-atrophic); (b) PgII and *H. pylori* status; and (c) PgII before and after successful or unsuccessful *H. pylori* eradication treatment.

MATERIALS & METHODS

A search for the available literature electively addressing PgII serology was conducted in the PubMed and Scopus databases up to December 31st 2020. The search included the following keywords: pepsinogen II, PgII, pepsinogen C, PgC, with "and" as a Boolean operator, in relation to atrophic gastritis, non-atrophic gastritis, gastritis activity, *H. pylori* status, *H. pylori* eradication, "and" PgII sensitivity, PgII specificity, and cut-off values. All scientific articles including the above-mentioned keywords were considered, whatever the method used to measure PgII levels (radioimmunoassay, ELISA or chemiluminescence). For each study, sensitivity and specificity values (with 95% confidence intervals [95% CI]) were both checked, together with positive and negative predictive values. Only scientific articles in English were considered. Studies lacking essential information that could not be obtained directly from the authors were also excluded.

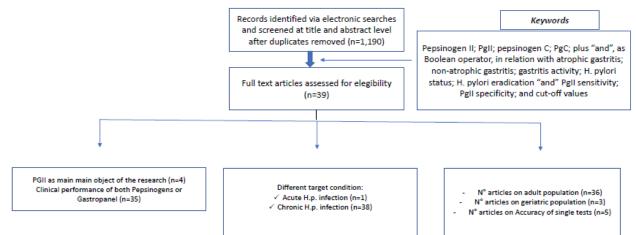
RESULTS

After removing duplicates, 1,190 records were identified, among which there were 35 full-text articles that included the sensitivity, specificity and cut-off values for the relationship between PgII levels and: i) gastritis; ii) *H. pylori* status; and iii) *H. pylori* eradication. In all but three of the studies ^{7, 8, 9}, PgII was tested together with PgI and/or in

conjunction with other serological variables (anti-*Hp* antibodies, gastrin 17 [Gastropanel]) (Fig. 1).

Fig. 1

Flow chart of literature search and selection



Nella tavola PGII va corretto in PgII

- *PgII serology in patients with gastric inflammatory lesions (with or without mucosal atrophy)*

Table 1 summarizes the results obtained by 9 studies in which PgII levels were tested in different gastritis phenotypes (non-atrophic *versus* atrophic), and compared with those of healthy controls. One further study only considered patients with atrophic gastritis (not those with non-atrophic gastritis or healthy controls).

Table 1: Pepsinogen II (PgII) in normal gastric mucosa, and in non-atrophic and atrophic gastritis phenotypes († atrophic and non-atrophic gastritis collapsed together; ‡ diagnosis established endoscopically; nr= not reported)

Author (reference)	Year	Patients (n.)	PgII (micrograms/L)				p value
			Normal mucosa	Non- atrophic gastritis	Atrophic gastritis	IM	
Broutet ¹³	2003	222	8.7	12.5	14.5		0.0001
Germanà 11	2005	287	$9.0{\pm}8.0$	15.7 ± 10.4	13.9 ± 10.1		nr
Sun ¹⁰	2007	841	7.4	12.8	13.0		nr
Haj-	2008	294	$6.6{\pm}2.8$	13.9 ± 9			nr
Sheykholeslami ²⁷							
He‡ ⁸	2011	1200	6.6	12.4	11.9	11.2	0.0001
Syrianen ⁶³	2017		$9.5 {\pm} 4.6$	17.3±10.1			nr
Han‡ ⁶⁴	2018	107			13.9±6.7 "limited atrophy"		0.025

					11.2±5.5 advanced atrophy	
Crafa ¹²	2020	266	5.8 ± 2.4	14.1±8.2	7.3 ± 3.6	nr
Kumar ²⁸	2020	210	12.77 ± 7.53	13.97 ± 8.14		
Miftahussurur ²⁶	2020	1206	8.5	14.2	2 †	nr

In all cases, PgII levels were significantly higher in patients with non-atrophic gastritis (range: 12.4 to 17.3 <u>micrograms</u> /L) than in healthy controls (range: 5.8 to 12.7 <u>micrograms</u>/L). In three of the five studies comparing atrophic *versus* non-atrophic gastritis ^{8, 10, 11, 12, 13} higher PgII levels were associated with non-atrophic gastritis. In a large study by He and coworkers⁸, the status of the gastric mucosa was only assessed endoscopically¹⁴: patients with both non-atrophic and atrophic gastritis showed significantly higher PgII serum levels than controls ¹⁵. Taken together, the studies considered suggest that PgII is consistently associated with gastric inflammatory lesions (more than with atrophy). Such an association is in keeping with the results obtained by Kuipers *et al.*¹⁶, who found a significant direct association between the amount of inflammatory infiltrate (as assessed histologically) and PgII serum levels.

- PgII serology by H. pylori status

Table 2 shows the relationship between serum PgII levels and *H. pylori* status. Seventeen studies (4 from Japan ^{17, 18,19, 20}; 2 from China ^{8, 21}; 1 from Korea ²²; 5 from Europe ^{7, 11, 12, 13, 23}; 1 from the USA ²⁴; 1 from South America ²⁵; 1 on populations of South and Southeast Asia ²⁶; 1 from Iran ²⁷; and one from India ²⁸) tested the clinical usefulness of PgII as a marker of active *H. pylori* infection.

Table 2: PgII levels by *H. pylori* status († gastric cancer patients; ‡ two weeks after acute *H. pylori* infection; Hp-*ve*: *H. pylori* negative patients; Hp+*ve*: *H. pylori* positive patients; nr: not reported in the publication)

				-	
Author (reference)	Year	Patients	PgII in	PgII in	p values
		(n.)	Hp- <i>ve</i> patients	Hp+ <i>ve</i> patients	
			(<u>micrograms</u> /L)	(<u>micrograms</u> /L)	
Broutet ¹³	2003	266	8.45	15.5	0.001
Di Mario ⁷	2004	313	8.6 ± 3.7	16.8 ± 7.4	< 0.001
Germanà 11	2005	287	8.9 ± 7.9	15.9 ± 1.0	< 0.001
Nurgalieva ²⁴	2006	20	9.1±8.5 ‡	42±40 ‡	0.001
Kim ²²	2007	1,485	8.0 ± 5.8	17.5 ± 10.5	< 0.001
Haj-Sheykholeslami	2008	481	9.8 ± 9	15.0 ± 9.9	nr
27					
He ⁸	2011	2,022	6.6	14.0	< 0.0001
Huang ²¹	2016	2,814	11.50 ± 7.45	18.09 ± 8.68	< 0.05
Kumar ²⁸	2016	168	13.97 ± 8.14 †	18.78 ± 12.63 †	0.034
Osumi ¹⁷	2017	650		$24.7{\pm}12.1$	
Kawamura ¹⁸	2019	45	24.7±13.1	16.3 ± 8.6	0.08
Kikuchi ¹⁹	2019		6.70		
Syrianen ²³	2019	80	9.5 ± 4.6	17.3 ± 10.1	< 0.001
Fernandez-Botran ²⁵	2020	203	8.2	17.8	< 0.0001

Miftahussurur ²⁶	2020	1,309	9.3	17.9	< 0.001
Okuda ²⁰	2020	187	9.5	17.0	< 0.01
Crafa ¹²	2020	246	$5.2{\pm}2.4$	14.1 ± 8.2	< 0.001

In all but one ²⁹ of the studies, *H. pylori*-positive patients had significantly higher PgII levels (range: 6.6 to 13.9 <u>micrograms</u>/L in Hp-*ve* individuals versus 14.0 to 24.7 <u>micrograms</u>/L in Hp+*ve* cases). These features were basically consistent in various epidemiological contexts.

Focusing on the relationship between PgII and CagA status, in keeping with results previously obtained by Broutet et al¹³, Okuda *et al* recently confirmed that serum PgII levels were significantly higher in CagA-seropositive infection (PgII levels: 9.0 <u>micrograms</u>/L in CagA-negative individuals *versus* 15.3 <u>micrograms</u>/L in CagA-positive ones; p = 0.001). This difference retained a borderline significance even when cancer patients were considered²⁵.

- PgII serology in H. pylori eradication

Table 3 summarizes the results of the studies addressing the changes (if any) in patients' serum PgII levels after they had been given successful *versus* unsuccessful *H. pylori* eradication therapy.

Table 3: PgII levels before and after successful or unsuccessful *H. pylori* eradication therapy († patients cured of their *H. pylori* infection; ‡ Patients not cured of their *H. pylori* infection; nr= not reported in the publication)

Author	Year	Patients	Serum	p value		
(reference)		(n.)	BeforeAfter eradication therapyeradicationsuccessfulunsuccessful			
			therapy	Successiui	unsuccessiui	
Hunter ²⁹	1993	nr	13. 3±0.8	7.9±0.7	13.6 ± 1.7	0.001
Tanaka ³⁰	2004	9	21.9±3	10.3±2		0.0001
Plebani 31	1992	49	Increased	Decreased	Unchanged	0.001
Pilotto 32	1996	88	21.58 ± 1.97	14.34 ± 1.75	21.12 ± 3.07	0.001
Plebani 33	1996	192†	18±1	8±1		0.001
		97‡	13±1		13±1	ns
Di Mario 7	2004	70	17.5 ± 7.5	$8.2{\pm}2.0$	15.6 ± 5.9	< 0.001
Gatta ³⁴	2011	228	17	8	16	< 0.001
Osumi ¹⁷	2017	650	24.7±12.1	7.5 ± 3.5	23.3±12.2	< 0.05

Serum PgII levels (tested between 1 and 2 months after completing the eradication therapy) dropped significantly only in patients whose treatment was successful, whereas the other patients' PgII levels remained much the same as before. Notably, these results were consistent whatever the method used to measure PgII (radioimmunoassay or ELISA).

Table 4: Cut-off values, sensitivity, and specificity for PgII in assessing the success of *H. pylori* eradication therapy († population over>65 years old; nr = not reported in the publication; ‡ according to Youden's Index)

Author	Year	Cut-off	Sensitivity %	Specificity %	Accuracy %
(reference)					
Hunter ²⁹	1993	-25%	82	62	nr
Pilotto ^{† 32}	1996	5.1 micrograms/L	59	62	60
		‡			
Di Mario 7	2004	-25%	93	91	93
He ⁸	2011	10.25	71.6	70.1	70,7
		<u>micrograms</u> /L			
Gatta ³⁴	2011	-22.7%	100	96.6	98
Osumi ¹⁷	2017	-25%	93.1	93.8	93.2

As for the PgII cut-off values for judging the success of *H. pylori* eradication therapy, most studies suggested a 23-25% decrease from a patient's PgII levels before the treatment. In an adult population, He⁸ established a cut-off at 10.25 μ g /L, significantly higher than the one proposed by Pilotto *et al.* in a geriatric population based on Youden's index (5.1 μ g /L)³². Depending on the cut-off adopted, the test's specificity ranged between 62% and 96.6%. Two studies reporting the positive (PPV) and negative (NPV) predictive values for PgII levels as an indicator of H. pylori eradication generated inconsistent results (PPV = 98-67.6%; NPV = 71-74%)^{7,8}.

DISCUSSION

Based on what was known about functional gastric physiopathology, IM Samloff suggested in the early 1980s that serum pepsinogens (I and II) could be useful serological markers of various kinds of gastric mucosal damage ³⁴. At around the same time, Ichinose and coworkers proposed a radioimmunoassay method for measuring serum pepsinogens driven by the same biological rationale³⁵. The promise of such non-invasive diagnostic procedures prompted a (possibly excessive) enthusiasm for the idea of a "serological biopsy". ³⁴

Over the years before *H. pylori* was discovered, PgII serology had been suggested as a non-invasive test potentially capable of providing clinically useful information on a variety of gastric diseases, including non-atrophic and atrophic gastritis, gastric polyps, and gastric cancer (GC). ^{10, 16, 28, 36, 37, 38, 39, 40, 41, 42}

After the discovery of *H. pylori*, further studies proposed testing pepsinogens (PgI and PgII) to identify patients harboring (advanced) atrophic gastritis, which had been widely recognized as the *H. pylori*-triggered field of cancerization in the so-called "epidemic" gastric cancer. A functional test for exploring the "efficiency" of the gastric mucosa thus came to be inappropriately perceived as revealing a "cancer marker".

Far more studies on the clinical impact of testing serum pepsinogens have focused on PgI (with 5,953 records, as opposed to 1,117 records for PgII), while PgII is largely tested "in support" of PgI values (as in the PgI/PgII ratio), as a marker of oxyntic atrophy ^{43, 44, 45}. This review aims to restore serum PgII testing to its original (appropriate) clinicalbiological role, examining both its clinical usefulness and its operative limitations.

In 2004, DY Graham ⁴⁶ unequivocally documented seroconversion (from anti-Hpnegative to anti-Hp-positive, with a prevalence of 94% in 4 weeks), and the onset of gastric mucosal inflammation in a model of acute *H. pylori* infection in a group of volunteers. A significant increase in serum levels of PgII (but not PgI) was also documented within the first two weeks of these volunteers becoming infected (PgII 9.1±8.5 μ g /L at the baseline, 18±16 after 7 days [p=0.008], and 42±40 after 14 days [p=0.001]). These results are consistent with those obtained by Lorente and coworkers in an in vitro study showing that *H. pylori* (irrespective of the strain involved) promotes PgII secretion from human peptic cells via an intracellular pathway mediated by calcium and nitric oxide. ⁴⁷

The present review likewise documented significantly higher serum PgII levels in gastritis patients who were infected with *H. pylori* (from 14.1 to 24.7 μ g /L) than in those who were not (below 10 μ g /L in all but three studies) ^{8, 10, 17, 27, 35, 49}. As expected (given the association between *H. pylori* infection and mucosal inflammation), serum PgII levels were always significantly higher in the patients carrying the infection, regardless of the laboratory method used (colorimetric, radioimmunoassay, ELISA, chemiluminescence) . ^{10, 36, 37, 49, 50, 51, 52} The increase in PgII levels also correlated with the amounts of all the types of inflammatory cells involved in the histological phenotype of gastritis. ¹⁶

PgII has also been proposed as a marker of successful *H. pylori* eradication therapy ever since 1992 (years before more sensitive/specific tests become available) ³¹. Judging from the findings of the present review (Table 4), PgII serum levels drop significantly after *H. pylori* eradication therapy, but the reliability of serological testing as an indicator of successful eradication suffers from inconsistencies in its sensitivity and specificity. As a cut-off that can be safely assumed to indicate that eradication therapy has been successful, most studies suggest a 25% drop in PgII levels after the treatment ^{7, 17, 29}. That said, the sensitivity and specificity values can vary considerably in adult populations (from 82% to 100%, and from 62% to 96.6%, respectively), reflecting the numerous situations that might potentially influence the results (Table 4). Much the same can be said of the results that emerged when PgII was tested in a selected population of geriatric patients. ^{32, 53} The variability seen in how PgII levels respond to the eradication effort may plausibly have to do with the fact that the efficacy of the treatment in down-modulating or clearing the inflammatory lesions (and lowering PgII levels as a consequence) may not necessarily reflect a successful eradication of the infection ^{54, 55,56}. On the other hand, the successful eradication of *H. pylori* may not always be associated with patients' inflammatory lesions disappearing (and their serum PgII levels dropping)^{57, 58, 59, 60, 61}. Il giallo si avvolge su se stesso e dovrebbe essere più esplicito... vorrebbe dre qualcosa, ma senza arrivare al punto dolente ...in più a prima frase è lunga

Overall, the findings of the present review do not alter the skeptical conclusion reached by M. Tarek Al-Assi in 1999, who wrote: "Despite a significant fall in serum markers of *H. pylori* infection in the groups of successfully treated individuals, no marker (PgII among others) tested in this study approached the reported accuracy of the urea breath test"⁶².

In conclusion, this review supports the reliability of PgII as a marker of gastric mucosal inflammation, particularly in non-atrophic gastritis caused by *H. pylori* infection. A drop in PgII levels of at least 25% (within 2 months after completing eradication therapy) supports a down-modulation of a patient's inflammatory lesions. On the other hand, PgII is a considerably less reliable indicator of the success of *H. pylori* eradication therapy. Its clinical usefulness is further lessened by the availability of more sensitive and specific (and even less invasive) test methods. When coupled with PgI levels (and the PgI/PgII ratio, as in the ABC classification ⁴⁴ and MAPS ⁴⁵), then PgII levels are useful as a marker of both gastric inflammation and *H. pylori* infection. Mi pare che così non vada perchè non è mai stato discusso il Rapporto.

Forse meglio togliere la frase o rimodularla The value of PgII in association with PgI testing ... ma toglierei anche questo

REFERENCES

1) Hunt RH, Camilleri M, Crowe SE, et al. The stomach in health and disease Gut. 201; 64(10):1650-68.

2) Schubert ML Functional anatomy and physiology of gastric secretion. Curr. Opin. Gastroenterol. 2015; 31(6):479-85

3) Engevik AC, Kaji I, Goldenring JR. The Physiology of the Gastric Parietal Cell Physiol Rev. 2020 Apr 1;100(2):573-602.

4) Gritti I, Banfi G, Roi G S. Pepsinogens: physiology, pharmacology pathophysiology and exercise Pharmacol Res. 2000; 41(3):265-81

5) Kageyama T. Pepsinogens, progastricsins, and prochymosins: structure, function, evolution, and development Cell Mol Life Sci. 2002; 59(2):288-306

6) Foster C, Aktar A Kopf D, Zhang P, Guttentag S. Pepsinogen C: a type 2 cell-specific protease Am J Physiol Lung Cell Mol Physiol. 2004; 286(2):L382-7

7) Di Mario F, Moussa AM, Cavallaro LG, et al. Clinical usefulness of serum pepsinogen II in the management of Helicobacter pylori infection. Digestion. 2004; 70(3):167–72.

8) He CY, Sun PL, Gong YH, Xu Q, Dong NN, Yuan Y. Serum pepsinogen II: a neglected but useful biomarker to differentiate between diseased and normal stomachs. J Gastroenterol Hepatol 2011; 26: 1039–1046

9) Gatta L, Di Mario F, Vaira D, et al. Quantification of serum levels of pepsinogens and gastrin to assess eradication of Helicobacter pylori. Clinical Gastroenterology and Hepatology. 2011; 9(5):440–2.

10) Sun LP, Gong HY, Wang L, Yuan Y. Serum pepsinogen levels and their influencing factors: a population-based study in 6990 Chinese from North China. World J Gastroenterol 2007; 28; 13(48): 6562-6567

11) Germaná B, Di Mario F, Cavallaro LG, et al. Clinical usefulness of serum pepsinogens I and II, gastrin-17 and anti-Helicobacter pylori antibodies in the management of dyspeptic patients in primary care. Digestive and Liver Disease. 2005; 37(7):501–8.

12) Crafa P, Franceschi M, Rodriguez Castro KI et al. Functional dyspepsia. Acta Biomed. 2020; 9:91(3).

13) Broutet N, Plebani M, Sakarovitch C, Sipponen P, Megraud F and the Eurohepygast Study Group Pepsinogen A, pepsinogen C, and gastrin as markers of atrophic chronic gastritis in European dyspeptics. Br J Cancer 2003 Apr 22;88(8):1239-47

14) Sugano K, Tack J, Kuipers EJ et al.; faculty members of Kyoto Global Consensus Conference. Kyoto global consensus report on Helicobacter pylori gastritis. Gut 2015; 64: 1353–67

15) Han YM, Chung SJ, Choi JM, Lee C, Kim JS. Long-term outcome of group D patients with negative serum anti-Helicobacter pylori antibody and positive serum pepsinogen test

in healthy Koreans: Long-term outcome of group D patients. J Dig Dis. 2018;19(9):529–39.

16) Kuipers EJ, Uyterlinde AM, Pena AS, Roosendaal R, Pals G and Nelis GF: Long-term sequelae of Helicobacter pylori gastritis. Lancet 1995; 345: 1525-1528.

17) Osumi H, Fujisaki J, Suganuma T, et al. A significant increase in the pepsinogen I/II ratio is a reliable biomarker for successful Helicobacter pylori eradication. PLoS One. 2017; 12(8):e0183980.

18) Kawamura Y, Funaki Y, Yoshimine T, et al. Characteristics and predictive factor of Helicobacter pylori-associated functional dyspepsia in Japanese patients. Digestion. 2019; 100(4):277–85.

19) Kikuchi S, Kato M, Mabe K, et al. Optimal criteria and diagnostic ability of serum pepsinogen values for Helicobacter pylori infection. J. Epidemiol. 2019; 29(4):147–54.

20) Okuda M, Lin Y, Mabe K, et al. Serum pepsinogen values in Japanese junior high school students with reference to Helicobacter pylori infection. J. Epidemiol. 2020; 30(1):30–6.

21) Huang RG, Xiao HL, Zhou B et al. Serum pepsinogen levels are correlated with age, sex and the level of Helicobacter pylori infection in healthy individuals. Am J Med Sci. 2016; 352(5):481-486.

22) Kim HY, Kim N, Kang JM, et al. Clinical meaning of pepsinogen test and Helicobacter pylori serology in the health check-up population in Korea. Eur J Gastroenterol Hepatol. 2009; 21(6):606-12

23) Syrjänen K, Eskelinen M, Peetsalu A, et al. GastroPanel® Biomarker Assay: the most comprehensive test for Helicobacter pylori infection and its clinical sequelae. A critical review. Anticancer Res. 2019; 39(3):1091–104

24) Nurgalieva ZZ, Opekun AR, Graham DY. Problem of distinguishing false-positive tests from acute or transient Helicobacter pylori infections. Helicobacter. 2006;11(2):69–74.

25) Fernandez-Botran R, Wellmann IA, Une C, et al. Seroprevalence of Helicobacter pylori/CagA antibodies in Guatemalan gastric cancer patients: association of seropositivity with increased plasma levels of pepsinogens but not soluble urokinase plasminogen activator receptor. Am J Trop Med Hyg. 2020; 103(1):260-265.

26) Miftahussurur M, Waskito LA, Aftab H, et al. Serum pepsinogens as a gastric cancer and gastritis biomarker in South and Southeast Asian populations. PLoS ONE. 9 2020;15(4):e0230064.

27) Haj-Sheykholeslami A, Rakhshani N, Amirzargar A, et al. Serum pepsinogen I, pepsinogen II, and gastrin 17 in relatives of gastric cancer patients: comparative study with type and severity of gastritis. Clin Gastroenterol Hepatol. 2008; 6(2):174-9

28) Kumar S, Kumari N, Mittal RD, Ghoshal UC. Pepsinogen-II 100 bp ins/del gene polymorphism and its elevated circulating levels are associated with gastric cancer,

particularly with Helicobacter pylori infection and intestinal metaplasia. Gastric Cancer. 2016; 19(3):808–16.

29) Hunter FM, Correa P, Fontham E, Ruiz B, Sobhan M, Samloff M. Serum pepsinogens as markers of response to therapy for Helicobacter pylori gastritis. Digest Dis Sci. 1993;38(11):2081–6.

30) Tanaka I., Tatsumi Y, Kodama T et al. Effect of Helicobacter pylori eradication on gastroesophageal function. J Gastroenterol Hepatol 2004; 19:251–257

31) Plebani M, Di Mario F, Stanghellini V, Delle Fave G. Serological tests to monitor treatment of Helicobacter pylori. Lancet. 1992; 340(8810):51-2

32) Pilotto A, Franceschi M, Leandro G, et al. The clinical usefulness of serum pepsinogens, specific IgG anti-HP antibodies and gastrin for monitoring Helicobacter pylori treatment in older people. J. Am. Geriatr. Soc. 1996; 44(6):665–70.

33) Plebani M, Basso D, Scrigner M, et al. Serum pepsinogen C: a useful marker of Helicobacter pylori eradication? J Clin Lab Anal. 1996; 10(1):1-5.

34) Samloff IM, Varis K, Ihamaki T, Siurala M, Rotter JI. Relationships among serum pepsinogen I, serum pepsinogen II, and gastric mucosal histology. A study in relatives of patients with pernicious anemia. Gastroenterology, 1982; 83 (1 Pt 2):204-9.

35) Ichinose M, Miki K, Furihata C, al. Radioimmunoassay of group II pepsinogen in human serum. Clin Chim Acta 1982; 122:61-69.

36) Benberin V, Bektayeva R, Karabayeva R, et al. Prevalence of H. pylori infection and atrophic gastritis among symptomatic and dyspeptic adults in Kazakhstan. A hospital-based screening study using a panel of serum biomarkers. Anticancer Research. 2013;8.

37) Khulusi S, Mendall MA, Patel P, Levy J, Badve S and Northfield TC. Helicobacter pylori infection density and gastric inflammation in duodenal ulcer and non-ulcer subjects. Gut 1995; 37: 319-324.

38) Uemura N, Okamoto S, Yamamoto S, Matsumura N, Yamaguchi S and Yamakido M: Helicobacter pylori infection and the development of gastric cancer. N Engl J Med 2001; 345: 829-832.

39) Valle J, Kekki M, Sipponen P, Ihamaki T and Siurala M: Longterm course and consequences of Helicobacter pylori gastritis. Results of a 32-year follow-up study. Scand J Gastroenterol 1996; 31: 546-550,.

40) Filipe MI, Munoz N, Matko I, Kato I, Pompe-Kirn V and Jutersek A: Intestinal metaplasia types and the risk of gastric cancer: a cohort study in Slovenia. Int J Cancer 1994; 57: 324-329,.

41) Ohata H, Kitauchi S, Yoshimura N, Mugitani K, Iwane M and Nakamura H: Progression of chronic atrophic gastritis associated with Helicobacter pylori infection increases risk of gastric. Int J Cancer . 2004; 109(1):138-43 42) Correa P, Haenszel W, Cuello C, Zavala D, Fontham E and Zarama G: Gastric precancerous process in a high-risk population: cohort follow-up. Cancer Res 1990; 50: 4737-4740

43) Watabe H, Mitsushima T, Yamaji Y et al. Predicting the development of gastric cancer from combining Helicobacter pylori antibodies and serum pepsinogen status: a prospective endoscopic cohort study Gut 2005; 54(6):764-8.

44) Pimentel-Nunes P, Libânio D, Marcos-Pinto R. et al. Management of epithelial precancerous conditions and lesions in the stomach (MAPS II): European Society of Gastrointestinal Endoscopy (ESGE), European Helicobacter and Microbiota Study Group (EHMSG), European Society of Pathology (ESP), and Sociedade Portuguesa de Endoscopia Digestiva (SPED) guideline update 2019 Endoscopy 2019; 51(04): 365-388

45) Dinis-Ribeiro M, Yamaki G, Miki K, Costa-Pereira A, Matsukawa M, Kurihara M. Meta-analysis on the validity of pepsinogen test for gastric carcinoma, dysplasia or chronic atrophic gastritis screening. J Med Screen 2004; 11:141–147

46) Graham D Y, Opekun A R, Osato M S, et al. Challenge model for Helicobacter pylori infection in human volunteers. Gut 2004; 53:1235–1243

47) Lorente S, Doiz O, Trinidad Serrano M, Castillo J, Lanas A. Helicobacter pylori stimulates pepsinogen secretion from isolated human peptic cells. Gut 2002; 50:13–18

48) Muhsen K., Sinnreich R., Dafna M., et al. Prevalence and determinants of serological evidence of atrophic gastritis among Arab and Jewish residents of Jerusalem: a cross-sectional study. BMJ Open 2019;9:e024689.

49) Dixon MF, Genta RM, Yardley JH, et al. Classification and grading of gastritis. The updated Sydney System. International Workshop on the Histopathology of Gastritis, Houston 1994. Am J Surg Pathol 1996; 20:1161–81.

50) Rugge M, Meggio A, Pennelli G, et al. Gastritis staging in clinical practice: the OLGA staging system. Gut 2007; 56(5):631-6

51) Pilotto A, Di Mario F, Franceschi M, et al. Cure of Helicobacter pylori infection in the elderly: effects of eradication on gastritis and serological markers. Aliment. Pharmacol & Ther. 1996; 10(6):1021–7.

52) Kadkhodaei S, Siavoshi F, Noghabi KA. Mucoid and coccoid Helicobacter pylori with fast growth and antibiotic resistance Helicobacter. 2020; 25(2):e12678

53) Zou Y, Qian X, Liu X, et al.. The effect of antibiotic resistance on Helicobacter pylori eradication efficacy: A systematic review and meta-analysis Helicobacter. 2020; 25(4):e12714

54) Brennan D, O'Morain C, McNamara D, M Smith S. Molecular Detection of Antibiotic-Resistant Helicobacter pylori Methods Mol Biol. 2021;2283:29-36

55) Josephson M, Skole K The Houston Consensus Conference on Testing for Helicobacter pylori Infection Clin Gastroenterol Hepatol. 2018; 16(12):2004-2005

56) Shiota S, Thrift AP, Green L, et al. Clinical Manifestations of Helicobacter pylori-Negative Gastritis Clin Gastroenterol Hepatol. 2017; 15(7):1037-1046.e3)

57) Rugge M, Genta RM, Di Mario F, et al. Gastric Cancer as Preventable Disease Clin Gastroenterol Hepatol. 2017; 15(12):1833-1843

58) Nordenstedt H, Graham DY, Kramer JR, et al. Helicobacter pylori-negative gastritis: prevalence and risk factors Am J Gastroenterol. 2013; 108(1):65-71)

59) Huang Q, Jia X, Chu Y, Zhang X, Ye H. Helicobacter pylori Infection in Geriatric Patients: Current Situation and Treatment Regimens Front Med (Lausanne). 2021; 30; 8:713908.

60) Al-Assi M T, Miki K, Walsh J H, Graham D P, Asaka M, Graham D Y. Noninvasive evaluation of Helicobacter pylori therapy: role of fasting or postprandial gastrin, pepsinogen I, pepsinogen II, or serum IgG antibodies Gastroenterol. 1999; 94(9):2367-72).

61) Syrjanen K. Serological Biomarker Panel (GastroPanel®): A test for non-invasive diagnosis of dyspeptic symptoms and for comprehensive detection of Helicobacter pylori infection. Biomark J Internet. 2017; 03(01). http://biomarkers.imedpub.com/serological-biomarker-panel-gastropanel-a-test-for-noninvasive-diagnosis-of-dyspeptic-symptoms-and-for-comprehensive-detection-of.php?aid=18564

63) Han YM, Chung SJ, Choi JM, Lee C, Kim JS. Long-term outcome of group D patients with negative serum anti-Helicobacter pylori antibody and positive serum pepsinogen test in healthy Koreans: Long-term outcome of group D patients. J Dig Dis. 2018;19(9):529–39.