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Survey of prevalence and seasonal variability of Listeria monocytogenes in raw cow milk from Northern Italy

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Abstract

Listeria monocytogenes is an important food-borne pathogen causing meningitis, meningo-
encephalitis and abortion. Both sporadic and epidemic human listeriosis cases are associated
with the consumption of contaminated foods. To assess the potential risk to consumer health,
the presence of L. monocytogenes was investigated using qualitative and quantitative methods
in raw milk (bulk tank milk and milk for vending machine) collected from 2010 to 2013 in
Northern Italy (Lombardy and Emilia-Romagna regions). Overall, L. monocytogenes was
detected in 145 on 8716 of raw milk samples, with a prevalence of 1.66% (95% C.I. 1.4%-
1.7%). The prevalence ranged from 0.52% (95% C.I. 0.3%-0.9%) in 2012 to 2.7% (95% C.I.
2.0%-3.8%) in 2013, but no trend of increase was observed in four-years of investigation. The
pathogen was detected from 2.2% (95% C.I. 1.9%-2.6%) of bulk tank milk and from 0.5%
(95% C.I. 0.3% - 0.8%) of milk for vending machine. A significative difference (p < 0.05) of
the prevalence data was observed between data collected in two different regions of Northern
Italy with an higher prevalence in Lombardy. In addition to the geographical area, the L.
monocytogenes presence was influenced also by the seasonal period of collection samples,
with peaks in spring and autumn. These results confirm the raw milk can be a source of
foodborne illness outbreaks if consumed without sanitizing treatments, but the low prevalence
and the low contamination levels (more than 80% of the contaminated samples contained <10
cfu ml-1 of L. monocytogeses) proving the hygienic quality of the milk produced in Northern
Italy.

Key words: Raw milk; incidence; Listeria monocytogenes; risk analysis.

1. Introduction

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51 Milk and dairy products are assumed to be implicated in 2-6% of the bacterial foodborne 52 diseases (De Buyser at al., 2001) and, in particular, the presence of foodborne pathogens in 53 raw milk and bulk tank milk has been widely reported (D'Amico and Donnelly, 2010; De Reu 54 et al., 2004; Gaya et al., 1996; Hassan et al., 2000; Jayarao and Henning, 2001; Kousta et al., 55 2010; Moshtaghi and Mohamadpour, 2007; Van Kessel et al., 2004; Waak et al., 2002). The 56 prevalence of pathogens in milk is influenced by numerous factors such as farm size, number 57 of animals on the farm, hygienic conditions, farm management practices, variation in sampling and types of samples evaluated, differences in detection methodologies used, geographical 58 59 location, and season. However, in spite of the variation, all of these surveys clearly demonstrated that milk can be a major source of foodborne pathogens of human health 60 61 significance (Oliver et al., 2005). 62 In this contest, *Listeria monocytogenes*, responsible of listeriosis, represents one of the most 63 serious food safety concerns. It has been isolated from many foods (Gianfranceschi et al., 64 2003), among which milk and cheeses (Bernini et al., 2013; Dalmasso and Jordan, 2014; 65 Manfreda et al., 2005; Pintado et al., 2005; Torres-Vitela et al., 2012) and it has been involved in numerous outbreaks occurring after consumption of contaminated milk and milk products 66 67 throughout the world (Linnan et al., 1988; Lyytikainen et al., 2000; Donnelly, 2001; Lunde'n 68 et al., 2004). In particular, dairy products contaminated with L. monocytogenes have been 69 implicated at almost the half of the reported listeriosis outbreaks in Europe (Lunden, 70 Tolvanen & Korkeala, 2004). The pathogen is widespread in nature and lives naturally in 71 plants and soil environments. Its ability to survive and grow over a wide range of 72 environmental conditions, including refrigeration temperatures, high salt concentration and 73 low pH, makes it a potential hazard in foods (Ryser, 2007). Moreover, L. monocytogenes into 74 food processing plants results in reservoirs that are difficult to eradicate: this is the case of

biofilms that are a constant issue in food processing environments (Oliver et al., 2005). So, in addition to the risk associated to direct consumption, the introduction of raw contaminated milk into dairy processing plants represents a risk to human health if milk is used unpasteurized for cheese making or in case of cross contamination (Kousta et al., 2010). Considering the threat represented by the pathogen in raw milk, a survey was conducted from January 2010 to September 2013, involving a large number of samples collected in different geographical area in the North of Italy. This research aimed to give a considerable overview of *L. monocytogenes* presence in raw cow milk, intended both for cheese making and for direct consumption, by evaluating the prevalence of the pathogen at farm level also in relation to the seasonality.

2. Materials and methods

2.1. Samples collection

A total of 8716 raw cow milk samples were collected from January 2010 to September 2013 in 942 farms located in Lombardy and Emilia-Romagna regions, Northern Italy. All the samples, consisting in 5897 samples of bulk tank milk intended for cheese making and 2819 samples of milk intended for sale in automatic vending machines, were collected into sterile containers, kept below 4 °C during transportation and analysed within 2 h after receipt. Samples were collected in the frame of Food Business Operator's self-control programs or in the frame of monitoring surveys officers of the Regional Veterinary Authority.

97 2.2. Detection and enumeration of L. monocytogenes

All samples were tested for the presence of *L. monocytogenes* on 25 ml of raw cow milk by means of qualitative methods.

The samples collected from 2010 to 2011 were examined qualitatively according to ISO 11290-1 (ISO, 1996). The samples collected from 2012 to 2013 were examined by a biomolecular method (real-time PCR) (Biorad AFNOR BRD 07/10-04/05) (AFNOR, 2004) to detect L. monocytogenes DNA. Samples testing positive were retested under a microbiological protocol according to ISO 11290-1 (ISO, 1996). Typical colonies (n = 5) presumed to be Listeria spp. were streaked from Agar Listeria acc. to Ottaviani & Agosti (ALOA) (Biolife Italiana, Teramo, Italy) supplemented with ALOA enrichment-selective supplements (Biolife Italiana) onto Tryptone Soya Yeast Extract Agar, TSYEA (Oxoid, Basingstoke, UK) and plates were incubated at 37 °C for 24 h. By following the Gram's staining, catalase reaction and tumbling motility were performed using the pure cultures obtained from TSYEA. The isolates resulted positive to phenotypic tests were inoculated on 5% sheep blood agar (Oxoid) to determine the Beta haemolytic reaction. For following confirmation, carbohydrate utilization and CAMP tests were performed. On samples found to be positive, the enumeration of L. monocytogenes was carried out according to the method described by ISO 11290-2 (ISO, 1998) to evaluate the prevalence of contamination level. 2.3. Data analyses

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The prevalence of *L. monocytogenes* in raw milk, calculated as proportion between positive samples on total sample, was expressed in percentage values. Statistical analysis was performed by Epi tools (http://epitools.ausvet.com.au): the confidence intervals (C.I.) of proportions were calculated with using the binomial exact method and the statistical significance of differences between proportions was evaluated by Chi-square (χ^2) test.

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3. Results and discussion

From 2010 to 2013, 8716 raw milk samples, intended both for cheese making and for vending machine, were collected in Northen Italy. Samples were taken from local farms within selfcontrol sampling programs and by the official veterinarians within state surveillance programs and were investigated for the presence of *L. monocytogenes*. The results are summarized in Table 1. The prevalence values in raw milk ranged from 0.52% (95% C.I. 0.3%-0.9%) in 2012 to 2.73% (95% C.I. 2.0%-3.8%) in 2013, but no trend of increase was observed in four-year investigation. Overall, L. monocytogenes was detected in 145 raw milk samples out of 8716, with a prevalence of 1.66% (95% C.I. 1.4%-2.0%). This result is mainly due (p > 0.05) to bulk tank milk contamination rather than to raw milk intended for vending machine. In fact, concerning bulk tank milk, L. monocytoges was found in 2.22% (95% C.I. 1.9%-2.6%) of samples (131/5897) in four years of survey (Table 2). These findings were in agreement with those reported in several studies carried out internationally and recently published, in which the prevalence of L. monocytogenes in raw milk has ranged from 'not detected' to 7.1%. In particular, the pathogen was "not detected" in Norway in 2011 (Jakobsen et al., 2011), and to the extent of 0.68% in New Zeland in 2012 (Hill et al., 2012), 2.12% in Turkey in 2006 (Aygun & Pehlivanlar, 2006), 2.61% in Algeria in 2007 (Hamdi et al., 2007), 5.5% in Finnish in 2013 (Ruusunen et al., 2013), 6.1% in North-West Spain in 2007 (Vilar et al., 2007) and 7.1% in USA in 2011 (Van Kessel et al., 2011). Previously, in 2005, the prevalence of L. monocytogenes in bulk tank milk has been reported to range from 1 to 12% (Oliver et al., 2005), therefore, a reduction of contamination samples seems to have been monitored in the last years. Anyway, the above data collected in different world areas underline the wide variability of the L. monocytogenes prevalence, which can be due to several factors, as geographical area, size farm, types of housings for the cattle. As further confirmation of this variability, the overall data we collected in this study show that even in the same geographical area there may be a different prevalence of *L. monocytogenes*.

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In Lombardy region, the pathogen was detected in 96 (2.6%; 95% C.I. 2.1%-3.1%) of 3721 samples analyzed and the prevalence was statistically highest (p < 0.05) compared to that found in Emilia-Romagna region, where L. monocytogenes was detected in 26 (1.2%; 95%) C.I. 0.8%-1.8%) of 2176 milk samples (data not shown). The different prevalence values can be due to the different treatment of the cattle usually practiced in the two Italian regions. In Lombardy, the most of the cattle is fed with silages, the housing of cattle is indoors and these practices, when combined with poor hygiene on the farm, may contribute to contamination of milk, in agreement with Husu et al. (1990) and Sanaa et al. (1993). In contrast with this, many researchers have identified the raw milk as a source of L. monocytogenes, but environmental and fecal contamination during the transportation and storage of milk have also been reported (Frece et al., 2010). Moreover, L. monocytogenes may also directly contaminate milk from animals with mastitis (Hird & Genigeorgis, 1990). A low prevalence of L. monocytogenes was observed over the four years survey in raw milk samples indented for vending machine (Table 2). The pathogen was detected in 0.50% (95% C.I. 0.03%-0.8%) of samples (14/2819), but the prevalence ranged from 0% (95% C.I. 0%-0.7%) in 2010 to 1.57% (95% C.I. 0.7%-3.1%) in 2011. A similar result was obtained by Bianchi et al. (2013) who detected the *L. monocytogenes* in 1.6% of the milk samples for vending machine during the monitoring survey from 2009 to 2011 in another Italian region (Piedmont region, near Lombardy). L. monocytogenes is widely extended throughout the environment (Fenlon et al., 1996). This pathogen has the ability to survive in stress conditions and it is able to grow at low temperatures in several food types as cooked meat (Daminelli et al., 2014), vegetables (Sant'Ana et al., 2012), cold-smoked salmon (Beaufort et al., 2007), milk and cheese (Schvartzman et al., 2010). To prevent the growth of the pathogen in raw milk, in Italy, where the sale and the distribution of unpacked raw milk via automatic self-service vending

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machines was authorized since 2007, the product must be maintained at constant temperature between 0 °C and 4 °C, and the customers are instructed to boil the milk before consumption. Even if the most important aspect remains the hygienic quality of the product, these practices can contribute to improve the safety of the raw milk, as shown the low prevalence of L. *monocytogenes* reported in the present study. To evaluate the seasonal influence on the L. monocytogenes presence in bulk tank milk, in Table 3 were reported the prevalence data for each year (from 2010 to 2013), broken down by each month. In four years, only in January on a total of 188 samples analyzed, milk samples positive for L. monocytogenes have never been found. Considering the seasonal variability, the L. monocytogenes prevalence was statistically lower (p < 0.05) during the winter season, with a prevalence of 0.8% (95% C.I. 0.3%-1.7%), in contrast with the spring prevalence of 3.04% (95% C.I. 2.3%-3.9%), the summer prevalence of 1.91% (95% C.I. 1.4%-2.6%) and the autumn prevalence of 2.33% (95% C.I. 1.6%-3.3%) (Table 3). Previous studies on Listeria spp. prevalence in raw milk reported some evidence of seasonal variation. Atil et al. (2011) in eastern Turkey observed a high prevalence in spring and winter; in France, Meyer-Broseta et al. (2003) reported peaks in winter. Ryser (1999) reported that seasonal variations in *Listeria* prevalence may be related to silage feeding, with higher prevalence in months when silage is fed to animals. In fact, L. monocytogenes could be present in silage, in which the pathogen can multiply if the silage has been inadequately fermented (pH above 5.0 to 5.5) (Husu, 1990). Seasonal differences in L. monocytogenes prevalence were observed in our monitoring, but more studies are needed to determine the validity and the causes of these differences. In fact, in the present study, the reduced number of samples found positive to L. monocytogenes in the winter season may be due to the lower number of samples analyzed (746 samples against over 1000 samples analyzed in each of the other seasons), or to other

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variables such as the type of cattle feed (fresh grass or silage), the hygienic conditions of breeding or the presence of cattle infected with mastitis.

The contamination levels of *L. monocytogeses* in bulk tank milk samples are reported in Figure 1 as percentage of samples with pathogen concentration variable between <1 and >1000 cfu ml⁻¹. Overall, more than 80% of the contaminated samples contained <10 cfu ml⁻¹ of *L. monocytogeses*. This trend is reflected from 2010 to 2012, while in 2013 the majority of the positive samples contained <1 cfu ml⁻¹ of the pathogen, as shown in Figure 1. It is difficult to compare the results from this study with those conducted in other countries, because most studies express the results only qualitatively. However, a study by Meyer-Broseta et al. (2003), shows that even in France, the level of contamination of bulk tank milk was very low, generally less than 1 cfu ml⁻¹ of the *L. monocytogenes*.

4. Conclusions

During 2007-2009 in Italy there was an increase of notifications of listeriosis with the most cases are reported in the Centre-North of Italy. This is probably attributable both to a real increase of listeriosis in Italy and to surveillance implementation (Pontello et al., 2012). However, statistically significant increasing trends in listeriosis notification rates from 2005 to 2009 were noted in Italy as elsewhere in Europe (EFSA/ECDC, 2011). In the present study the raw milk was found positive to the presence of *L. monocytogenes*, confirming the milk as potential source of the food borne disease, but the low prevalence and the reduced level of pathogen concentration, when present, can highlight the hygienic quality of the milk produced in Northern Italy. Furthermore, the collected data in the present study can be a useful tool for the quantitative risk assessment study for human listeriosis linked to the consumption of raw milk and cheese made from raw milk in Italy. Furthermore, the collected data in the present

223 study can be a useful tool for the quantitative risk assessment study for human listeriosis 224 linked to the consumption of raw milk and cheese made from raw milk in Italy. 225 226 **Disclosure** 227 Authors declare that no conflict of interests exists. 228 229 Acknowledgements 230 We sincerely thank Dr. Silvia Todeschi (Food Microbiology Department, IZSLER, Brescia, 231 Italy) for the technical assistance and for the informatics support development to the data 232 management. 233 234 References 235 AFNOR. (2004). Biorad AFNOR BRD 07/6-07/04. http://www.afnor validation.org/pdf/AFNOR-Validation2008.pdf. Accessed 26 June 2014. 236 237 Atil, E., Ertas, H. B., & Ozbey, G. (2011). Isolation and molecular characterization of *Listeria* 238 spp. from animals, food and environmental samples. Veterinarni Medicina, 56(8), 386-394. 239 Aygun, O., & Pehlivanlar, S. (2006). Listeria spp. in the raw milk and dairy products in 240 Antakya, Turkey. Food Control, 17, 676-679. 241 Beaufort, A., Rudelle, S., Gnanou-Besse, N., Toquin, M. T., Kerouanton, A., Bergis, H., ... & 242 Cornu, M. (2007). Prevalence and growth of *Listeria monocytogenes* in naturally 243 contaminated cold-smoked salmon. Letters in Applied Microbiology, 44, 406-411. 244 Bernini, V., Bottari, B., Dalzini, E., Sgarbi, E., Lazzi, C., Neviani, E., & Gatti, M. (2013). The 245 presence, genetic diversity and behaviour of *Listeria monocytogenes* in blue-veined cheese 246 rinds during the shelf life. Food Control, 34, 323-330.

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Table 1. Detection and prevalence of *L. monocytogenes* in raw milk collected in Northern Italy according to the year of sampling.

Year	Samples	Positive for <i>L. monocytogenes</i> (%)		
2010	1728	20 (1.16%)		
2011	3150	76 (2.41%)		
2012	2519	13 (0.52%)		
2013	1319	36 (2.73%)		
Total 8716		145 (1.66%)		

Table 2. Detection and prevalence of *L. monocytogenes* in different categories of raw milk collected in Northern Italy according to the year of sampling.

	Bulk Tank Milk		Raw Milk for vending machine		
Year	Samples	Positive for <i>L.</i> monocytogenes (%)	Samples	Positive for L. monocytogenes (%)	
2010	1176	20 (1.70%)	552	0	
2011	2639	68 (2.58%)	511	8 (1.57%)	
2012	1317	9 (0.68%)	1202	4 (0.33%)	
2013	765	34 (4.44%)	554	2 (0.36%)	
Total	5897	131 (2.22%)	2819	14 (0.50%)	

Table 3. Seasonal detection and prevalence of *L. monocytogenes* in bulk tank milk collected in Northern Italy according to the year of sampling.

C	Month —		Sample positive / Sample analyzed (%)				
Season		2010	2011	2012	2013	Total for Month	Total for Season
Winter	December	0/58 (0%)	2/106 (1.89%)	0/45 (0%)	nc	2/209 (0.96%)	6/746 (0.8%)
	January	0/47 (0%)	0/49 (0%)	0/70 (0%)	0/22 (0%)	0/188 (0%)	
	February	0/108 (0%)	1/51 (1.96%)	3/158 (1.9%)	0/32 (0%)	4/349 (1.15%)	
Spring	March	2/174 (1.15%)	7/233 (3%)	0/215 (0%)	21/212 (9.91%)	30/834 (3.6%)	56/1844 (3.04%)
	April	0/74 (0%)	8/225 (3.56%)	0/79 (0%)	12/167 (7.19%)	20/545 (3.67%)	
	May	1/112 (0.89%)	5/218 (2.29%)	0/76 (0%)	0/59 (0%)	6/465 (1.29%)	
Summer	June	4/84 (4.76%)	5/297 (1.68%)	0/80 (0%)	0/73 (0%)	9/534 (1.69%)	37/1934 (1.91%)
	July	2/116 (1.72%)	8/463 (1.73%)	5/212 (2.36%)	1/148 (0.68%)	16/939 (1.7%)	
	August	2/129 (1.55%)	10/121 (8.26%)	0/162 (0%)	0/49 (0%)	12/461 (2.6%)	
Autumn	September	6/119 (5.04%)	12/485 (2.47%)	1/154 (0.65%)	0/3 (0%)	19/761 (2.5%)	32/1373 (2.33%)
	October	3/97 (3.09%)	5/273 (1.83%)	0/35 (0%)	nc	8/405 (1.98%)	
	November	0/58 (0%)	5/118 (4.24%)	0/31 (0%)	nc	5/207 (2.42%)	

nc: data not collected.

Figure 1. Counts of *L. monocytogenes* in bulk tank milk according the year of sampling, expressed as percentages of samples containing colony numbers ranging from < 1 to > 1000 cfu ml⁻¹.

■2010 □ 2011 □ 2012 % of samples № 2013 1-10 <1 100-1000 10-100 >1000 Contamination level (cfu ml⁻¹)

*Highlights (for review)

Highlights

- 1. Survey study of *Listeria monocytogenes* in raw cow mil in Northern Italy.
- 2. Low level of milk contamination with *L. monocytogenes*.
- 3. Seasonal differences in *L. monocytogenes* prevalence (lower in winter).
- 4. Overall, more than 80% of the contaminated samples contained low L. monocytogeses concentration (<10 cfu ml⁻¹).